The Protein Hourglass: First Passage Time Distributions for Protein Thresholds

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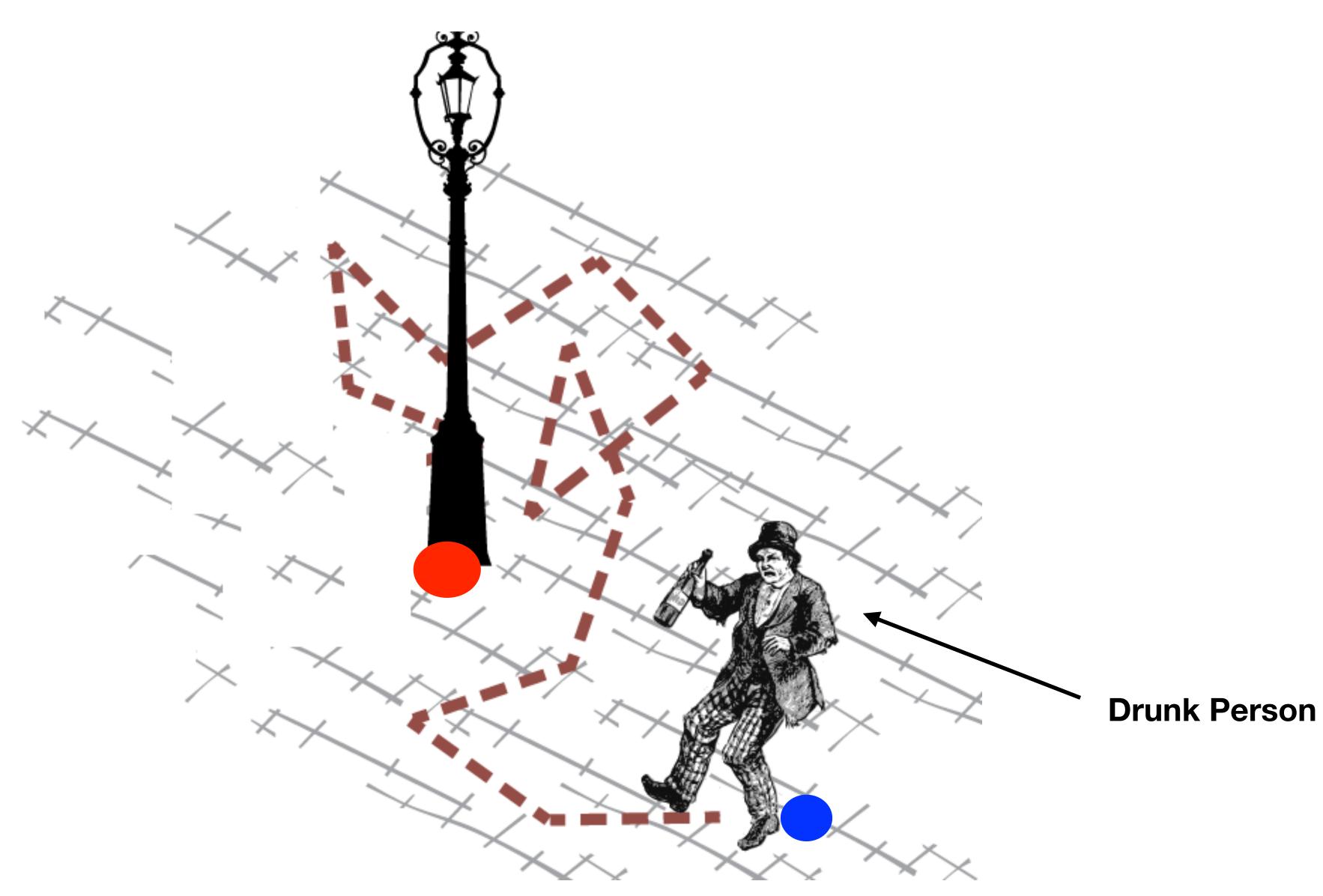
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SERB (Matrics), India

Basic idea of First Passage

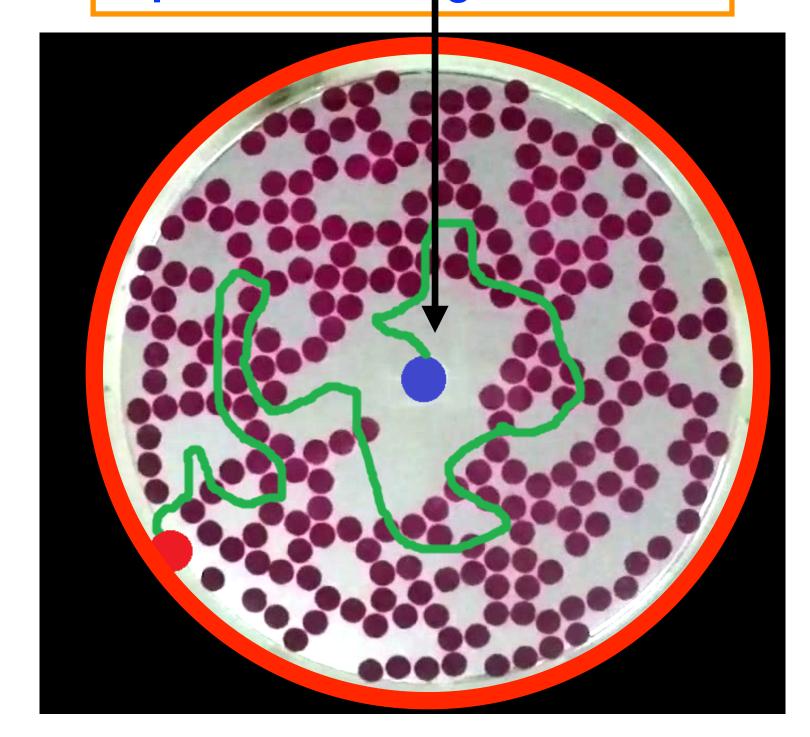
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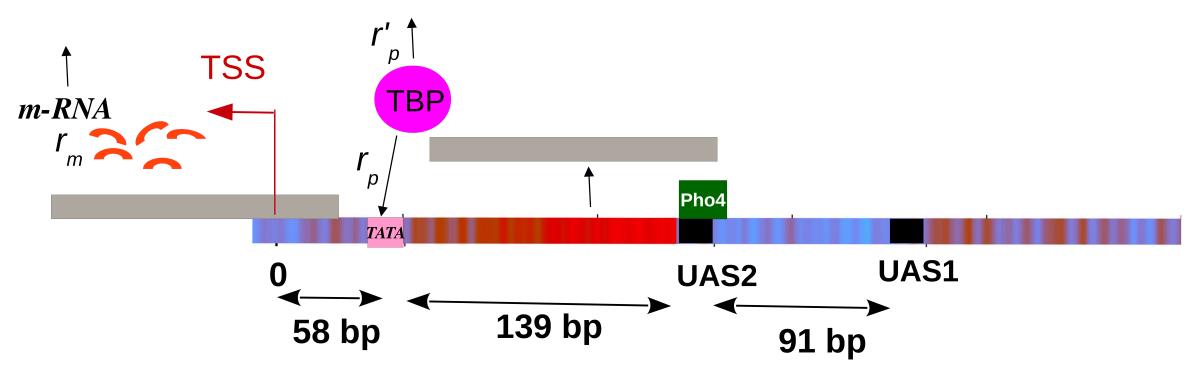
Some works on First Passage from our group

Biswas, Cruz, Parmanada & Das, Soft Matter (2020)

Active Camphor boat through passive floating crowders



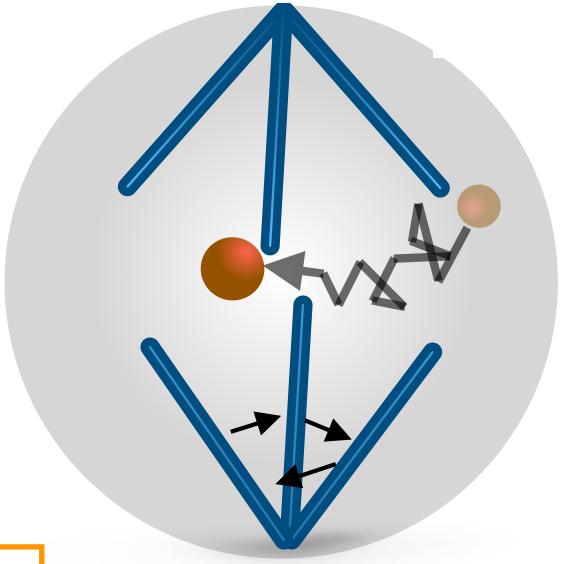
First binding of TBP on TATA box in promoter



Parmar, Das & Padinhateeri, NAR (2016)

Kinetochore Capture by multiple microtubules

Nayak, Das & Nandi, PRR (2020)



A lamb chased by many lions in confined volume

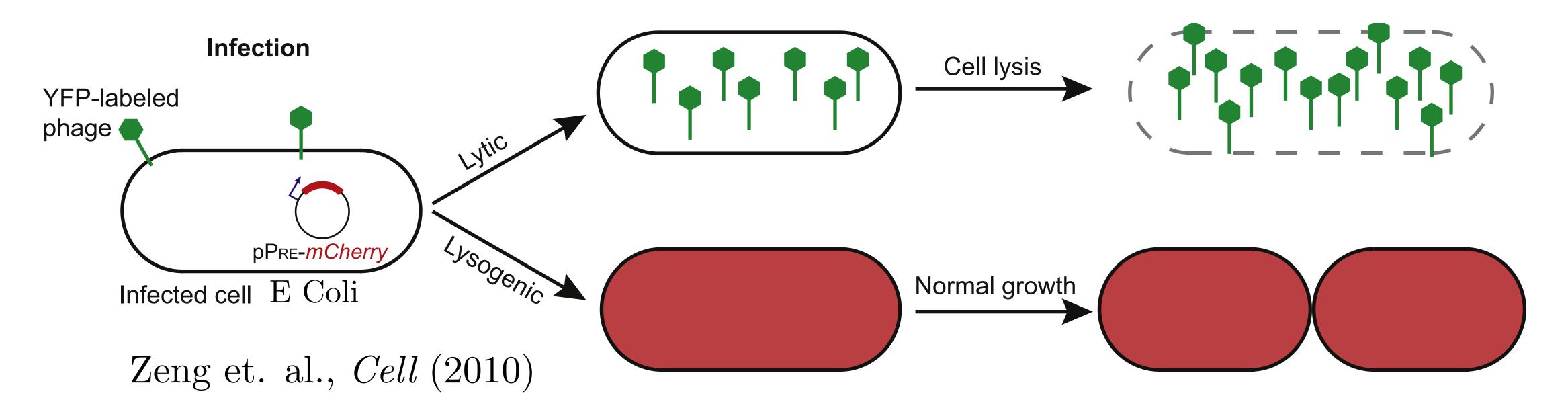
Nayak, Nandi & Das, PRE (2020)

Story of Phage-Lambda infecting E coli

Lysis caused by Holin accumulation

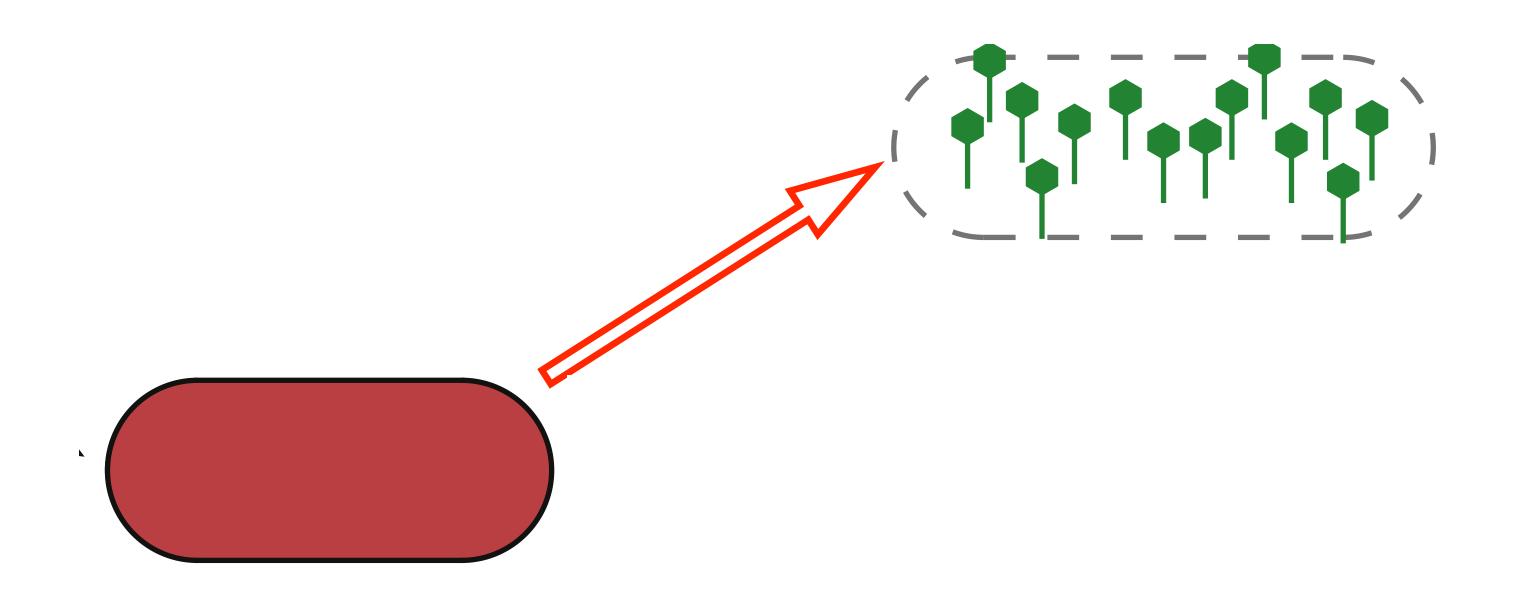
Virus infecting bacteria — Bacteriophages

 $Virulent \text{ (Only lysis, e.g. T4)} \qquad \qquad Temperate \\ \lambda \text{ phage: Lysis-Lysogeny switch}$



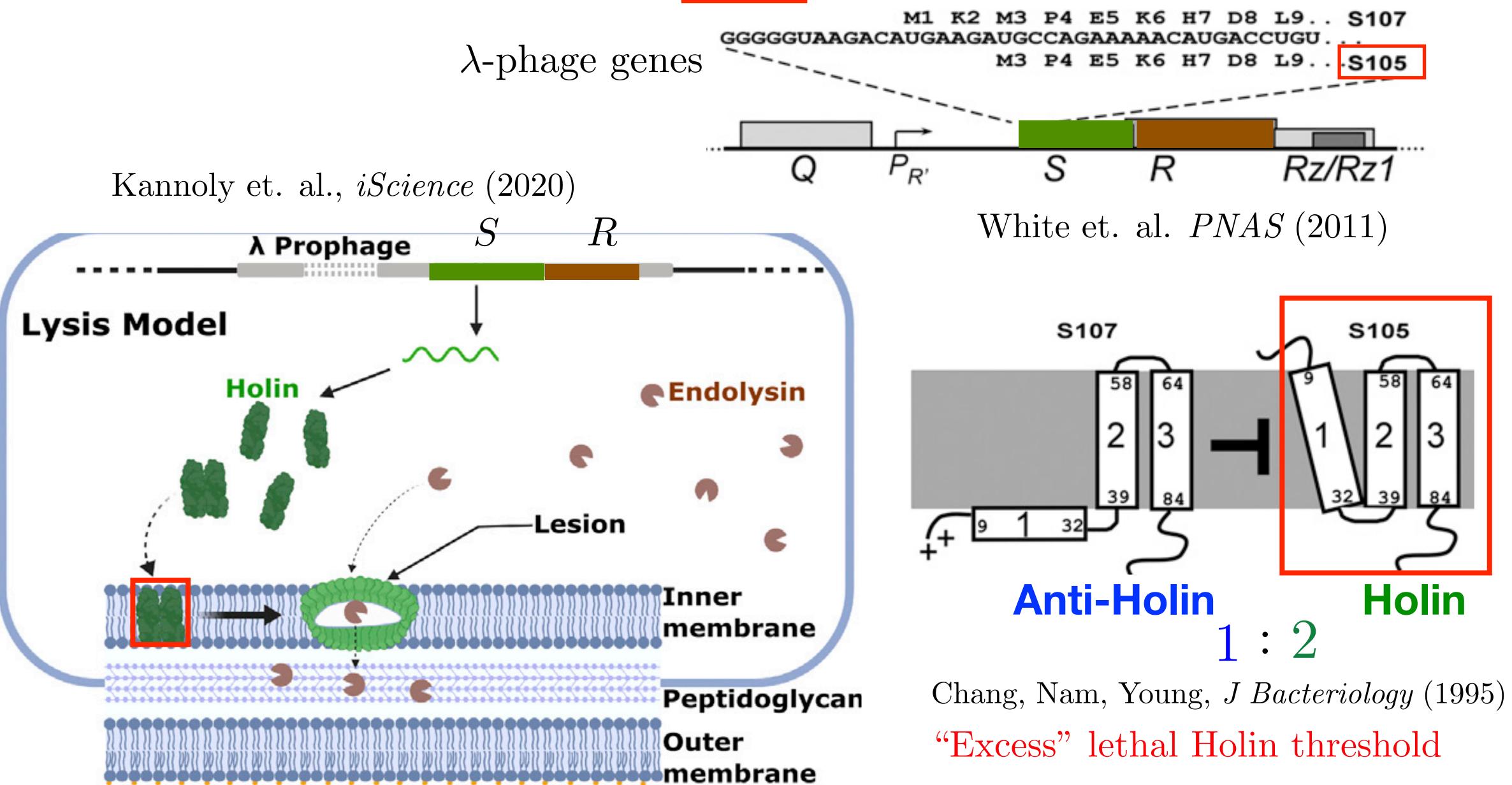
Many controlling factors: M.O.I, E coli cell volume, Stochastic expression of CI, Cro, CII

Virus infecting bacteria — Bacteriophages Virulent (Only lysis, e.g. T4) Temperate λ phage: Lysis-Lysogeny switch

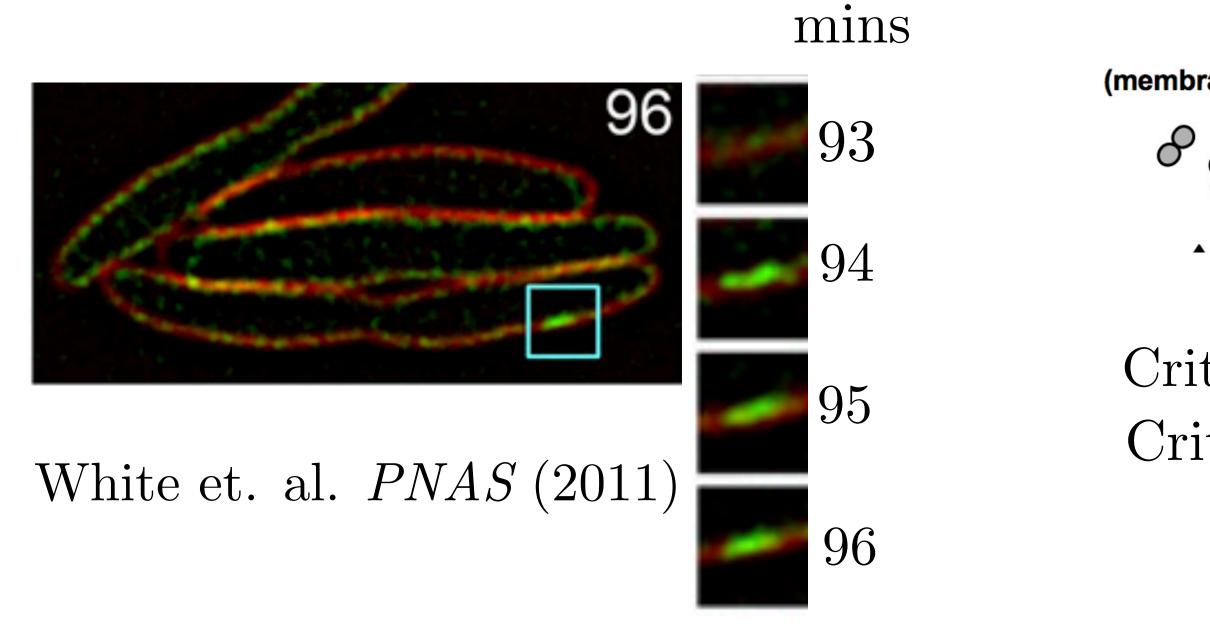


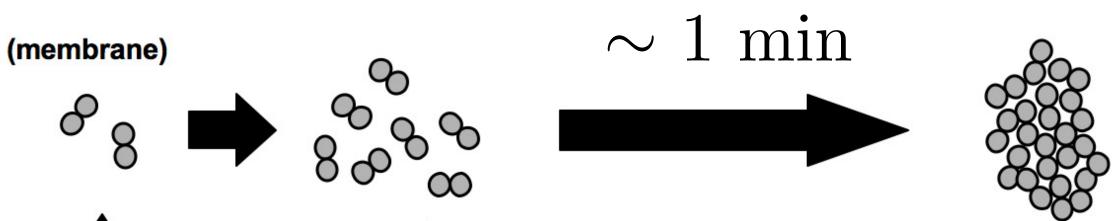
- Focus on Lysis Thermal induction of Lysogenic Bacteria (t = 0 set precisely)
- What role does Holin proteins play?

What controls lysis? Holin & Endolysin



What is important for Lysis? — Holin threshold and Time to reach it





Dennehy & Wang, BMC Microbiology (2011)

Critical Raft size - No

Critical number on membrane – YES

Holin Theshold: $X \approx 1500$

Chang, Nam, Young, J Bacteriology (1995)

In WT
$$\lambda$$
, Lysis time $\approx 65 \pm 3.2$ mins Dennehy & Wang, BMC Microbiology (2011)
$$\langle t_L \rangle = \langle t_{pR'} \rangle + \langle FPT \rangle$$
 ≈ 15 ≈ 50

As $n_{\text{Holin}}: 0 \to X$

Holin threshold & First Passage Time

It is very precise ...

HOLINS: The Protein Clocks of Bacteriophage Infections

Wang, Smith & Young, Annu Rev Microbiol (2000)

Holin triggering in real time

White et. al. PNAS (2011)

It is imprecise too (has stochasticity) ...

Factors influencing lysis time stochasticity in bacteriophage λ

Dennehy & Wang, BMC Microbiology (2011)

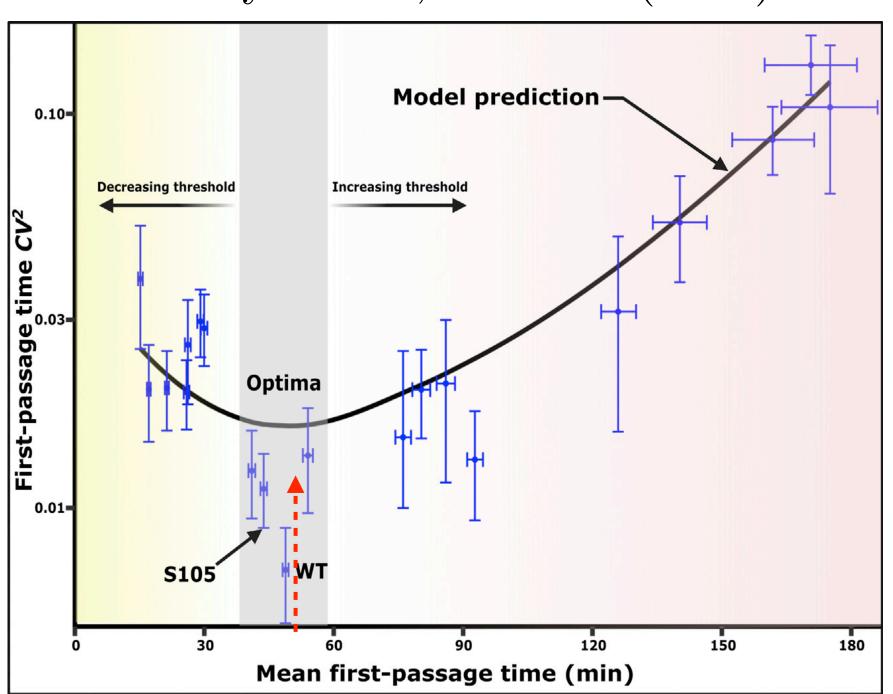
Stochastic holin expression can account for lysis time variation in the bacteriophage λ Singh & Dennehy, J.R. Soc Interface (2014)

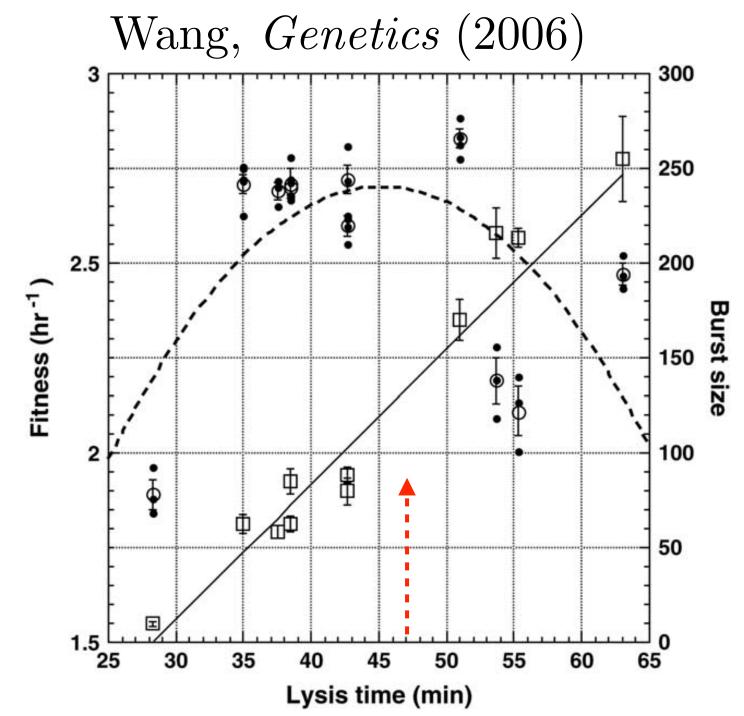
Can one "Control / Engineer "Mean First Passage Time and fluctuations?

- Bacteria growth rate
- Control late promoter pR' activity
- Holin mutants alleles, altered amino acid sequences (affect holin number on membrane)

Dennehy & Wang, BMC Microbiology (2011)

Kannoly et. al., iScience (2020)





• Different lysis times \Rightarrow different fitness

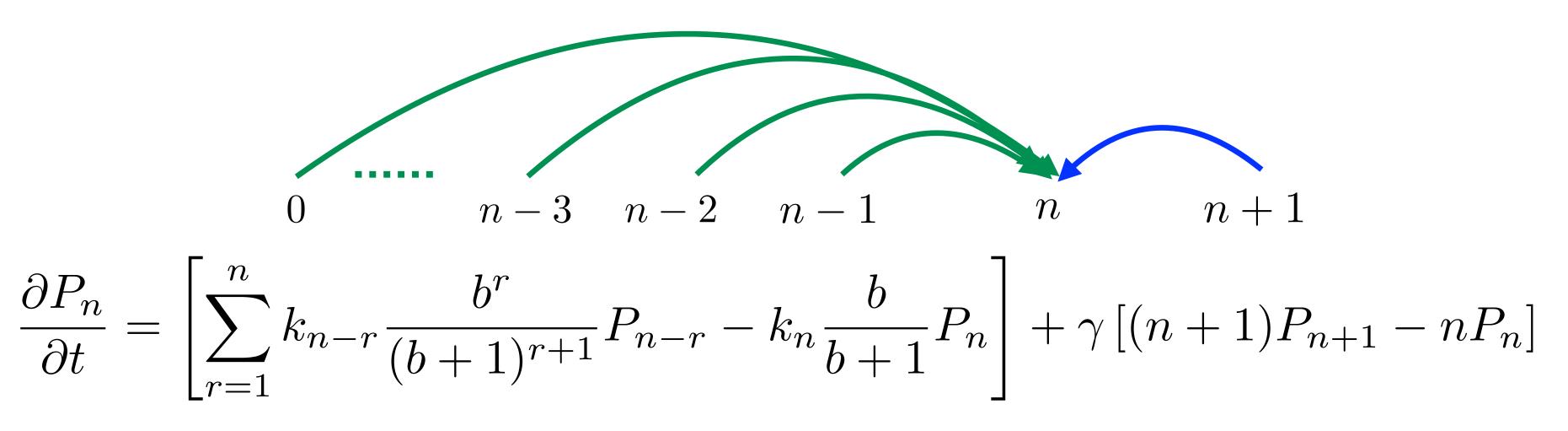
Different lysis time \Rightarrow different CV^2 Is lysis time $\langle t_L \rangle$ "Optimal" for WT λ ?

• Gene expression (noisy)— intrinsic Full Statistics?

Mathematical formulation of the problem

Calculation of the First Passage distribution

Protein bursts and degradation: Forward Equations



(for $k_n = k = \text{constant}$) Shahrezaei & Swain, PNAS (2008)

For
$$x = n/V$$
, $\frac{\partial p(x,t)}{\partial t} = k \int_0^x dx' \left[\nu(x-x') - \delta(x-x') \right] p(x',t) + \gamma \frac{\partial}{\partial x} [xp(x,t)]$
with $\nu(x) = (1/b) \exp(-x/b)$ Experiments, Nature (2006)
Friedman, Cai, & Sunney Xie, PRL (2006)

Steady State:
$$p(x, t \to \infty) = \frac{1}{b^a \Gamma(a)} x^{a-1} e^{-x/b}$$
 (here $a = k/\gamma$)

Gamma distribution

Moments of the First Passage time to reach the Holin threshold

Ghusinga, Dennehy, Singh, PNAS (2017)

Stochastic First Passage time = $min\{t: n(t) \ge X | n(0) = 0\}$

Solved: Using Forward Master Equation formalism, obtained Mean and Variance for any $k_n \neq \text{constant } k$ (with feedback), and $\gamma \neq 0$ (with degradation)

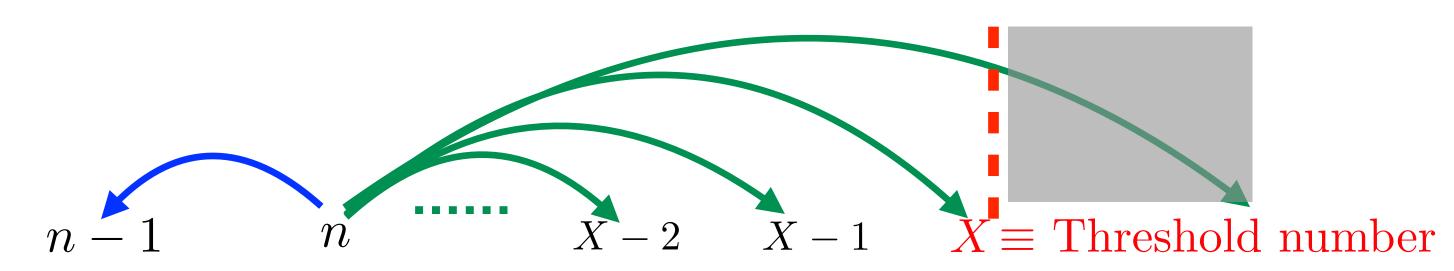
$$CV^2 = \sigma^2/\langle t \rangle^2$$

- For Holin ($\gamma \approx 0$), fluctuations (CV^2) minimum for no feedback ($k_n = k$)
- For other proteins which degrade fast $(\gamma > 0)$, CV^2 minimised for positive feedback (e.g. $k_n = c_1 + c_2 n$, or Hill form $k_n = k_{max} \frac{r + (cn)^H}{1 + (cn)^H}$)

Full distribution?

Full First Passage distribution: Backward Formalism

Rijal, Prasad & Das, PRE (2020)



Survival probability $S_n(t)$: Protein number stays below threshold X at time t, starting with n at t=0

$$\frac{\partial S_n}{\partial t} = k_n \left[\sum_{r=1}^{X-1-n} \frac{b^r}{(b+1)^{r+1}} S_{n+r} - \frac{b}{b+1} S_n \right] + \gamma \left[nS_{n-1} - nS_n \right]$$

$$S_{n \ge X}(t) = 0 \text{ and } S_{n < X}(0) = 1$$

Compact form: $\frac{\partial}{\partial t}\vec{S}(t) = \mathbf{A}\vec{S}(t)$

First Passage time distribution with n(0) = 0: $f_{0,X}(t) = -\frac{\partial}{\partial t} S_{0,X}(t)$

Technical details

Laplace Transforms: $\tilde{f}_{0,X}(s) = 1 - s\tilde{S}_{0,X}(s)$ where $\tilde{S}_{0,X}(s) = \sum_{j=1}^{X} (s\mathbf{I} - \mathbf{A})_{1j}^{-1}$

Challenges: (1) To find the matrix elements $(s\mathbf{I} - \mathbf{A})_{ij}^{-1}$ for any X

(2) To find the Laplace Inverse: $f_{0,X}(t) = \mathcal{L}^{-1}[\tilde{f}_{0,X}(s)]$

Our results suitable for Holin ($\gamma = 0$, i.e no degradation)

For equal $k_n = k$ (no feedback):

$$f_{0,X}(t) = \frac{kb^X}{(b+1)^X} \left[\sum_{n=0}^{X-1} \frac{1}{n!} {X-1 \choose n} \left(\frac{kt}{b+1} \right)^n \right] e^{-\frac{kbt}{b+1}}$$

Note: Exponential tail (not Gaussian)

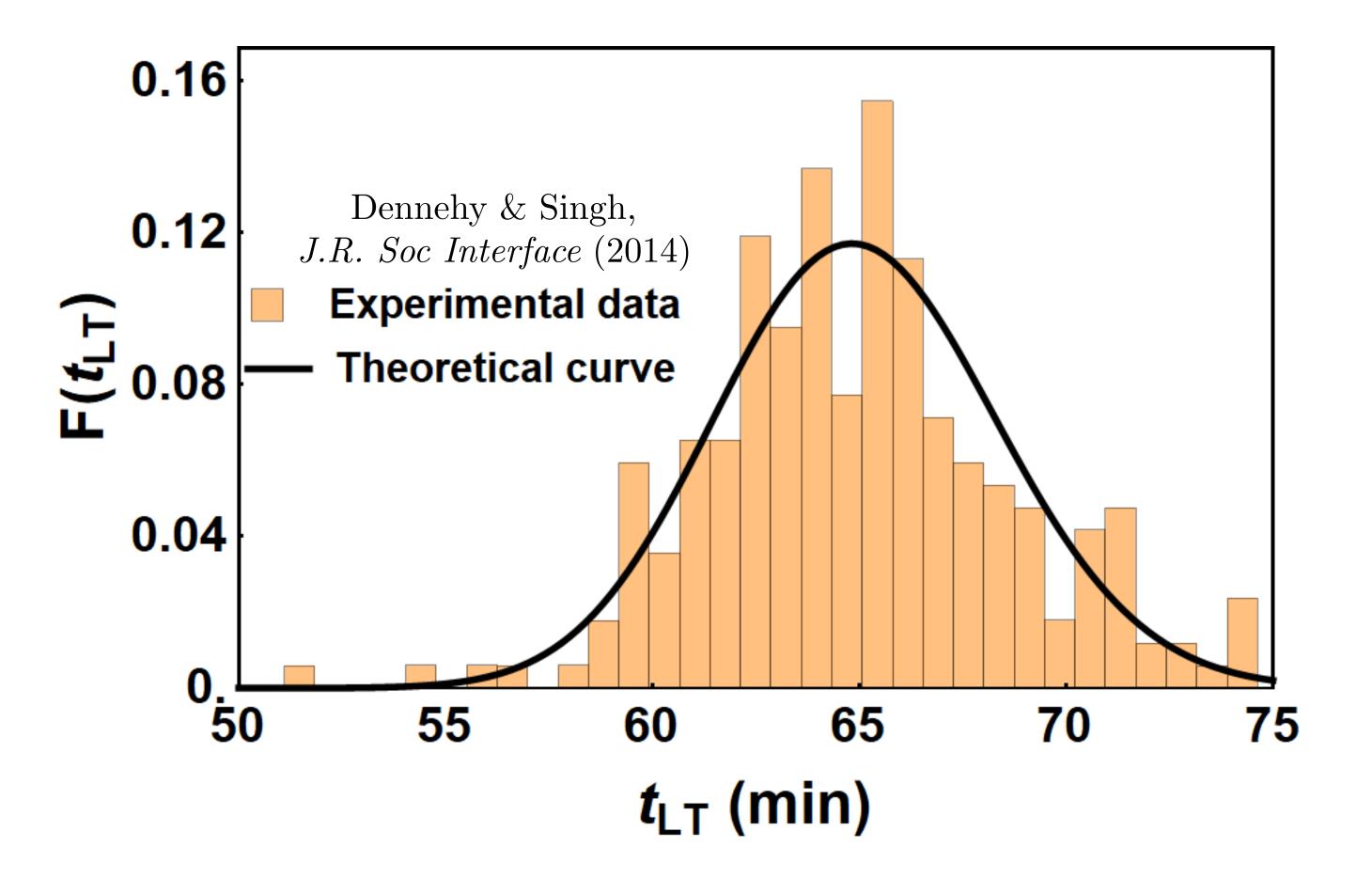
For unequal k_n (with positive or negative feedback):

$$f_{0,X}(t) = \frac{bk_0}{(b+1)^2} \left[\left(\frac{k_1}{k_1 - k_0} + b \right) e^{-\frac{bk_0t}{b+1}} + \left(\frac{k_1}{k_0 - k_1} \right) e^{-\frac{bk_1t}{b+1}} \right.$$
$$\left. + \sum_{j=3}^{X} \sum_{n=0}^{j-1} \frac{k_n}{(b+1)^{j-2}} \frac{\prod_{q=1}^{j-2} ((b+1)k_q - bk_n)}{\prod_{p=0, p \neq n}^{j-1} (k_p - k_n)} e^{-\frac{bk_nt}{b+1}} \right]$$

Comparison with experimental WT Holin, Lysis time distribution

Use Experimental values of: X = 1500, $\langle t_{LT} \rangle \approx 65$ mins & $\sigma_{LT} \approx 3.5$ mins

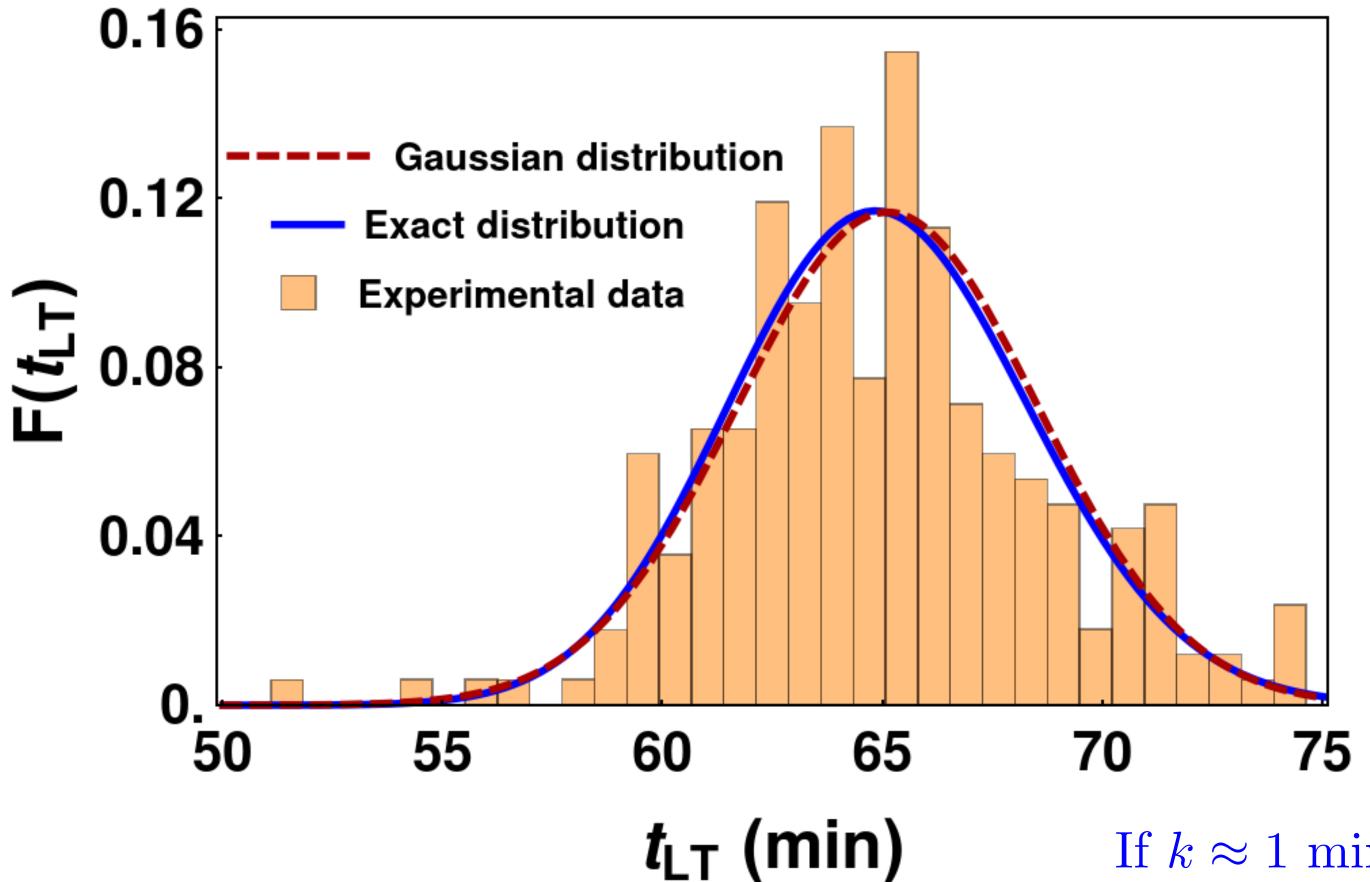
Theory:
$$\langle t \rangle = \frac{b+X}{bk}$$
 and $CV^2 = \frac{\sigma^2}{\langle t \rangle^2} = \frac{b^2 + X + 2bX}{(b+X)^2}$
 $\langle t_{LT} \rangle \approx 15 + \langle t \rangle \qquad \Rightarrow b = 3 \text{ and } k = 10 \text{ min}^{-1}$



Comparison with experimental (WT lambda) Lysis time distribution

Use Experimental values of: X = 1500, $\langle t_{LT} \rangle \approx 65$ mins & $\sigma_{LT} \approx 3.5$ mins

Theory:
$$\langle t \rangle = \frac{b+X}{bk}$$
 and $CV^2 = \frac{\sigma^2}{\langle t \rangle^2} = \frac{b^2 + X + 2bX}{(b+X)^2}$, all cumulants κ_n analytically $\langle t_{LT} \rangle \approx 15 + \langle t \rangle$ $\Rightarrow b = 3 \text{ and } k = 10 \text{ min}^{-1}$



Skewness =
$$\frac{\kappa_3}{(\kappa_2)^{\frac{3}{2}}}$$

Gaussian: 0

Theory: 0.10 , Exp: 0.06

Kurtosis =
$$3 + \frac{\kappa_4}{\kappa_2^2}$$

Gaussian: 3

Theory: 3.02 , Exp: 3.54

If $k \approx 1 \text{ min}^{-1}$ and $b \approx 30$, then skewness ≈ 0.30

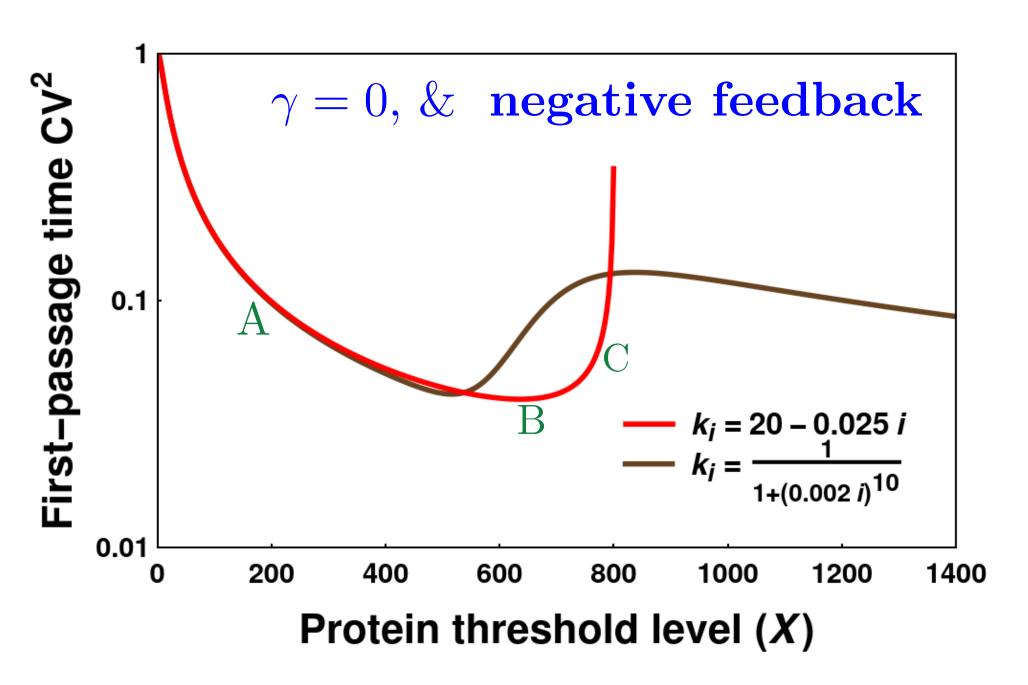
Short lived proteins: with degradation rate $\gamma \neq 0$

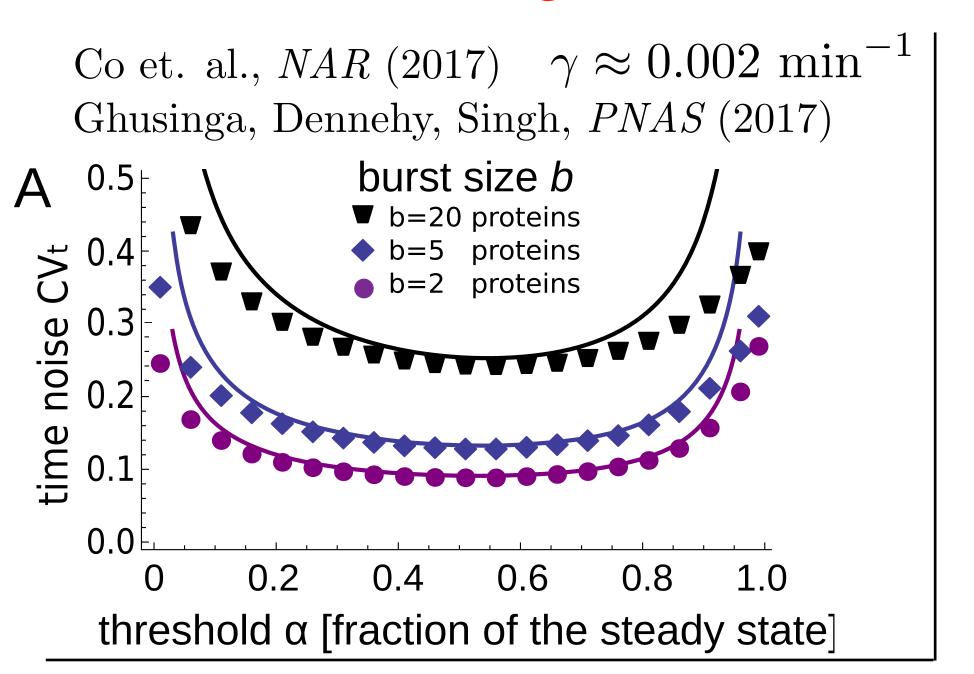
Laplace Transform:

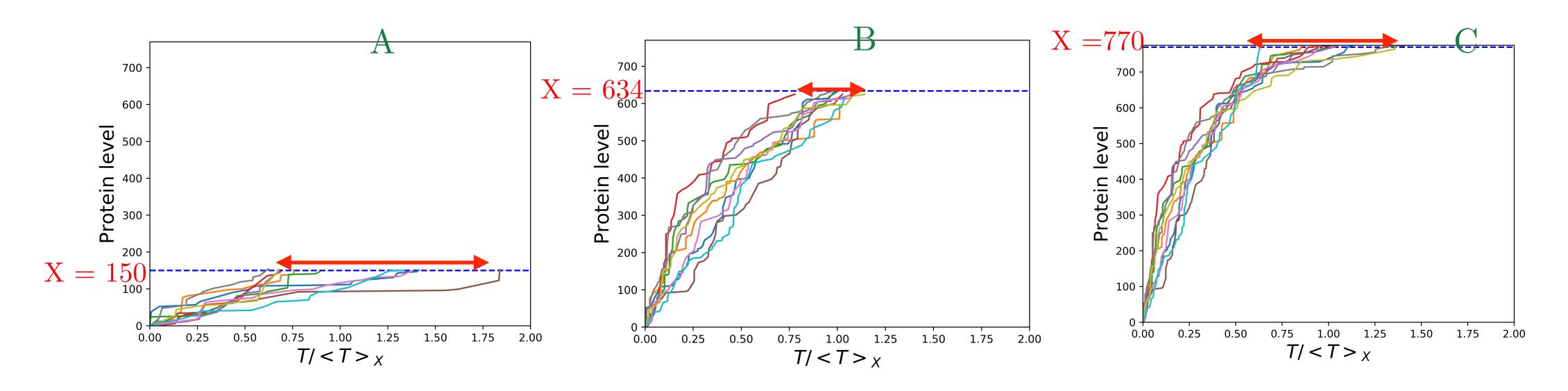
$$\tilde{f}_{0,X}(s) = \frac{b^X k \prod_{l=1}^{X-1} (\gamma l + k + s)}{\prod_{i=0}^{X-1} (\gamma i + s) + b^X (k + s) \prod_{j=1}^{X-1} (\gamma j + k + s) + \sum_{n=1}^{X-1} \left[\binom{X}{n} b^n \prod_{i=0}^{X-1} (\gamma i + k \Theta(i - X + n) + s) \right]}$$

- Finding the poles, for general X, and hence finding $f_{0,X}(t)$ remains difficult.
- We obtained the third moment (not known before), & can predict Skewness.

Noise versus X, non-monotonic behaviour for negative feedback







Concluding comments

- 1) We obtained the full First Passage time distribution, for long lived proteins (like Holin in Lambda phage). [For short lived proteins, we have the exact expression of the Laplace transform of the distribution.]
- 2) The skewness and departure from Gaussian may be more clearly discernible if transcription & translation rates and burst size may be engineered.
- 3) For long live proteins too, a non-monotonic noise (CV) is possible for "Negative feedback"
- 4) Results for continuous protein concentration x = n/V, have been derived, and those are consistent with the results for discrete protein numbers n.

Thank You