

Regulation of Motor Traffic and Control of Noisy Gene Expression: Roles of Signals, Sequence, Structure and Suppressors in Decoding and Recoding

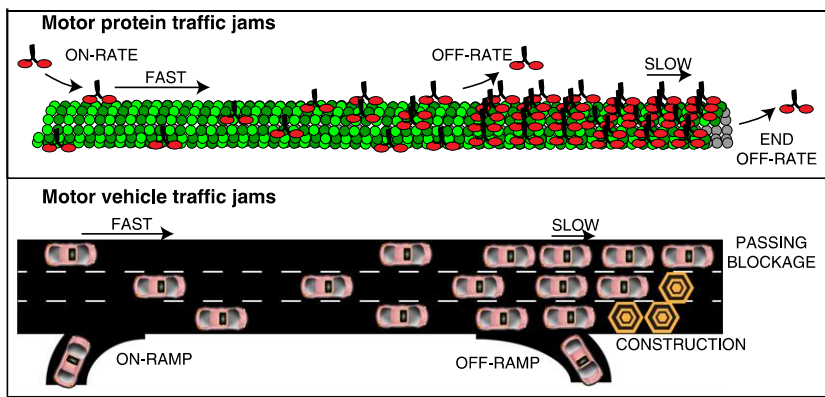
Debashish Chowdhury

Physics Department, I.I.T., Kanpur



Home page: http://home.iitk.ac.in/~debch/profile_DC.html

Alternative Home page: <https://sites.google.com/site/debchphy/>



J.L. Ross, PNAS (2012)

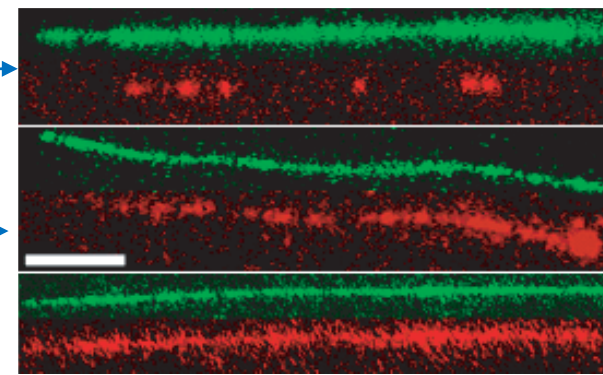
**NO underlying HAMILTONIAN,
NO equilibrium exists!!**

Traffic of molecular
motors on MT track

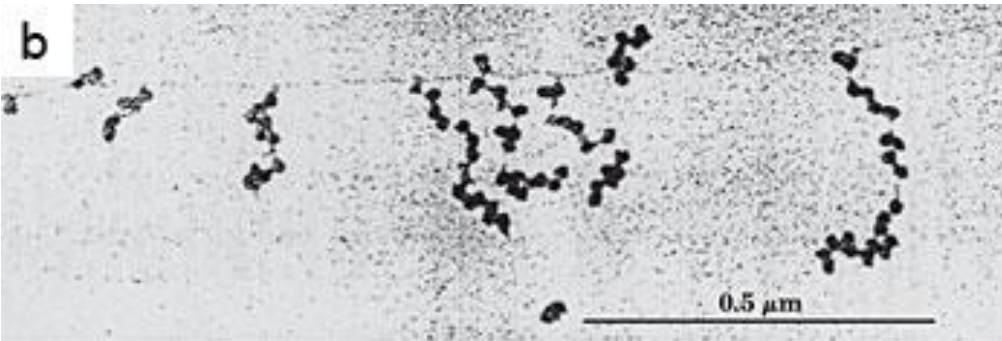
← MCAK

KIF1A →

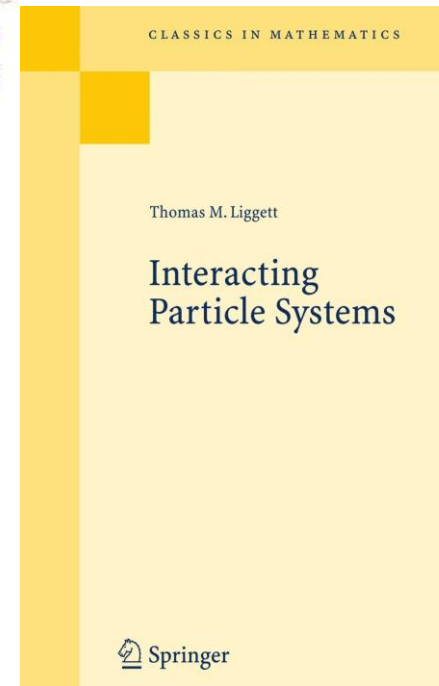
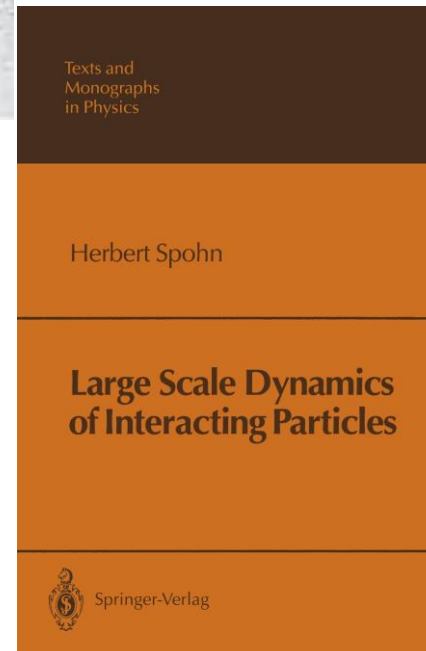
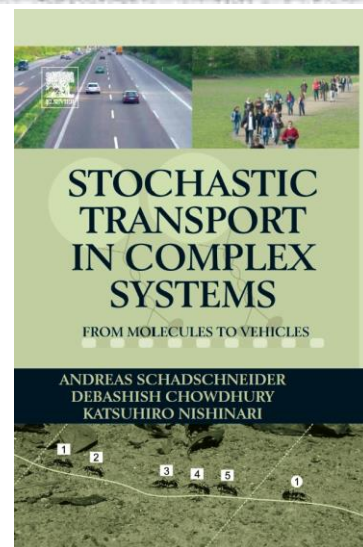
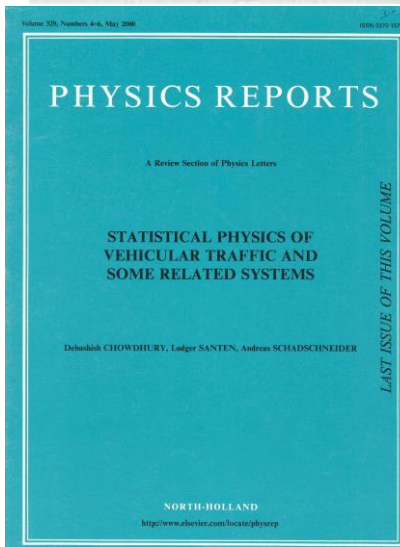
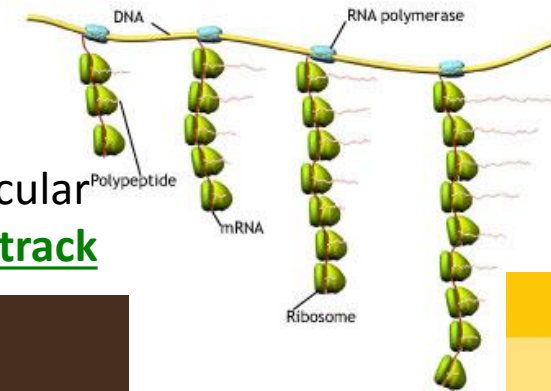
**Non-equilibrium
STEADY-STATE**



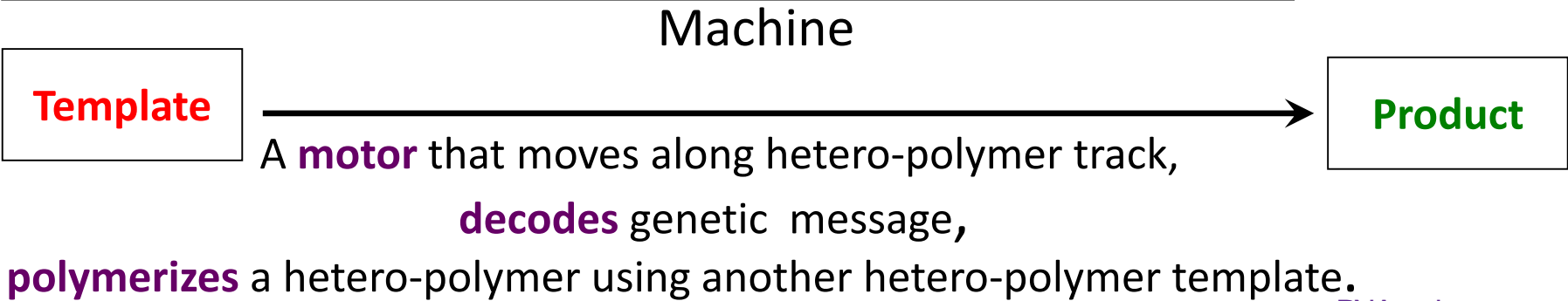
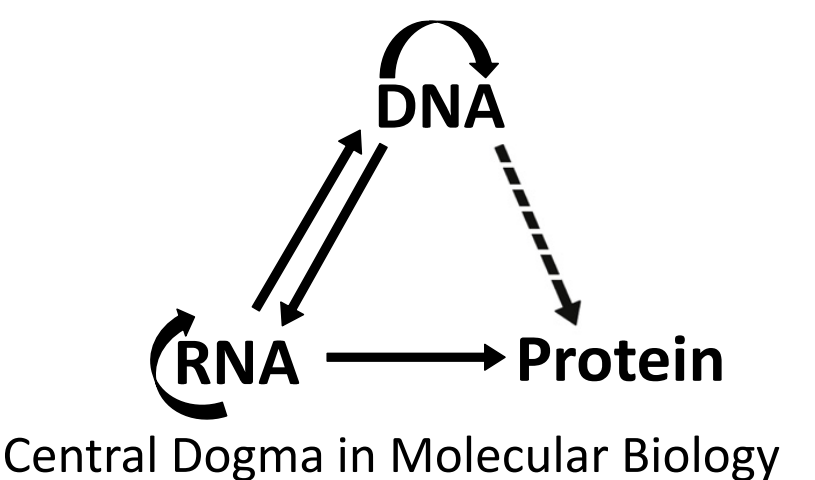
Nishinari, Okada, Schadschneider,
Chowdhury. Phys. Rev. Lett (2005)



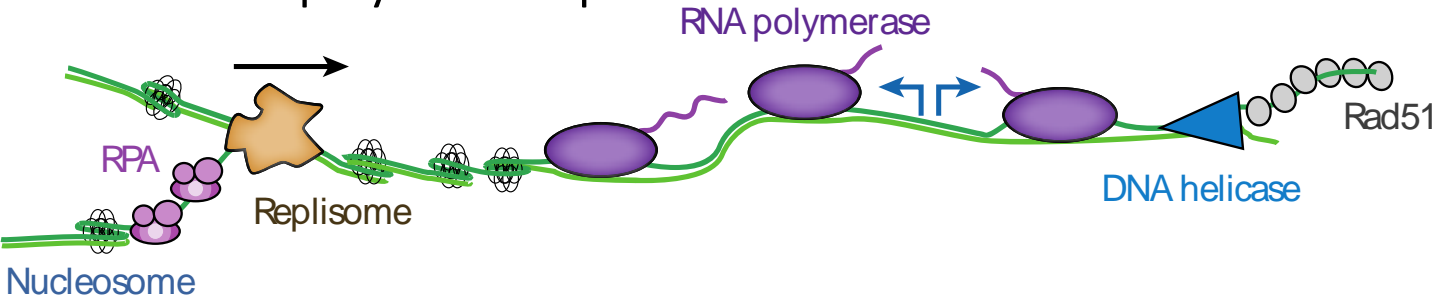
Traffic of molecular
motors on NA track



Machine	Template	Product	Function
DdRP	DNA	RNA	Transcription
DdDP	DNA	DNA	DNA replication
RdRP	RNA	RNA	RNA replication
RdDP	RNA	DNA	Reverse transcription
Ribosome	mRNA	Protein	Translation



Molecular Traffic Jams on DNA

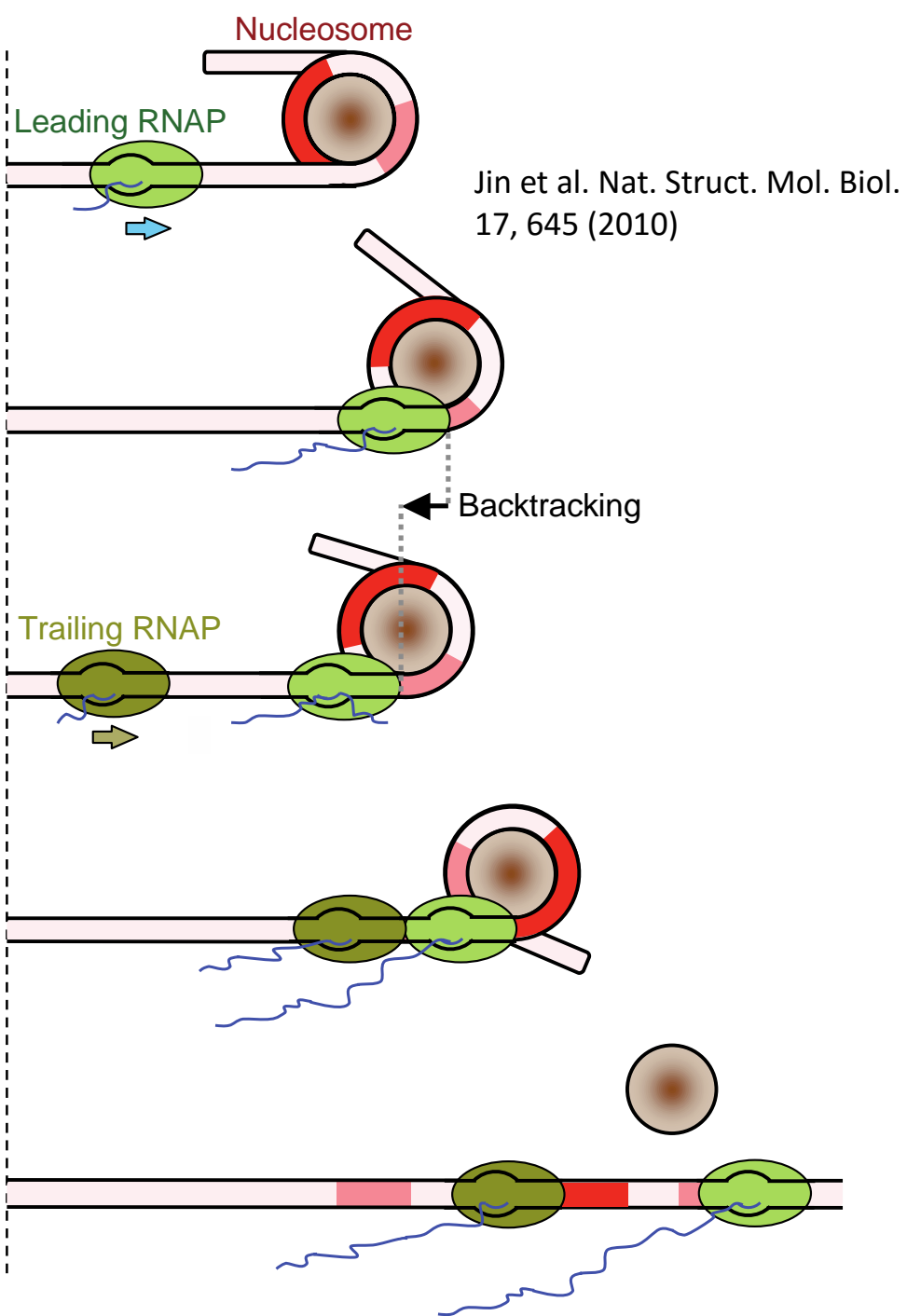


Ilya J. Finkelstein^{1,2} and Eric C. Greene^{3,4}

Figure 1

DNA replication, transcription, and repair must occur simultaneously on a crowded nucleic acid substrate. Conflicts between these molecular machines must be resolved rapidly to maintain cell viability and to avoid genomic instability.

Beneficial effects of Traffic Congestion



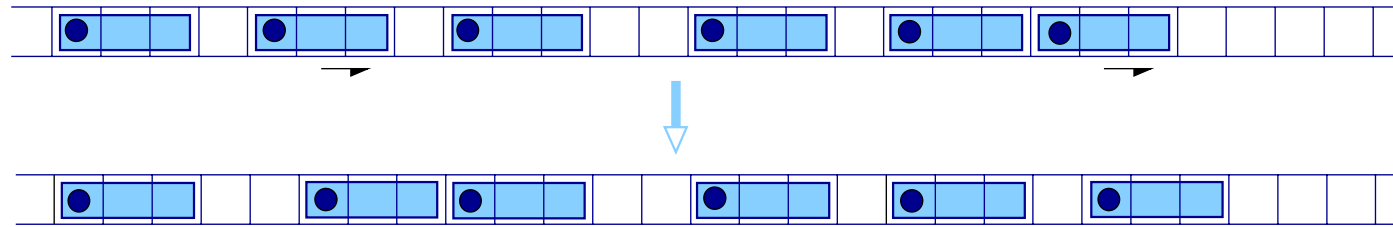
Following RNAP can push the leading stalled RNAP assisting it in dislodging the nucleosome proteins thereby resuming Transcription.

Arabidops

Ribosome Profiling

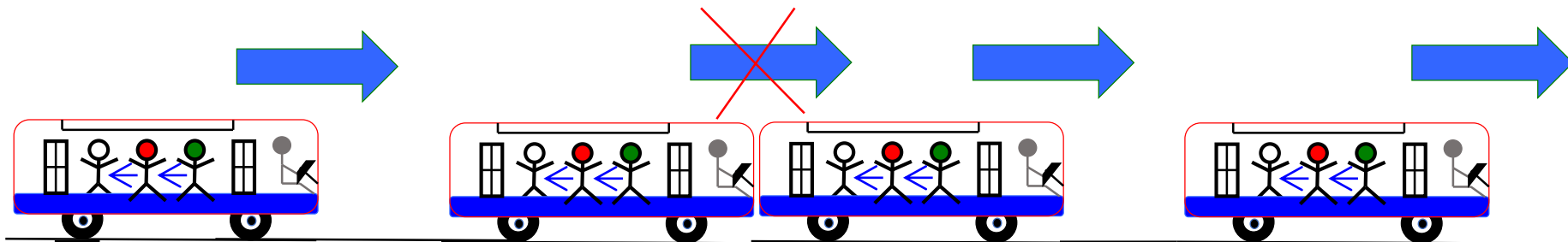
somes Polysomes
Juntawong et al. PNAS (2014)

$$\rightarrow | \ell = 3 | \leftarrow$$



Kinetics of biopolymerization on nucleic acid templates

Carolyn T. MacDonald, Julian H. Gibbs, Allen C. Pipkin
Biopolymers, Volume 6, Issue 1 January 1968 Pages 1–25



Polysome: Ribosome Traffic on a single mRNA

Basu & Chowdhury, Phys. Rev. E (2007)

Garai, Chowdhury, Chowdhury, Ramakrishnan, Phys. Rev. E (2009)

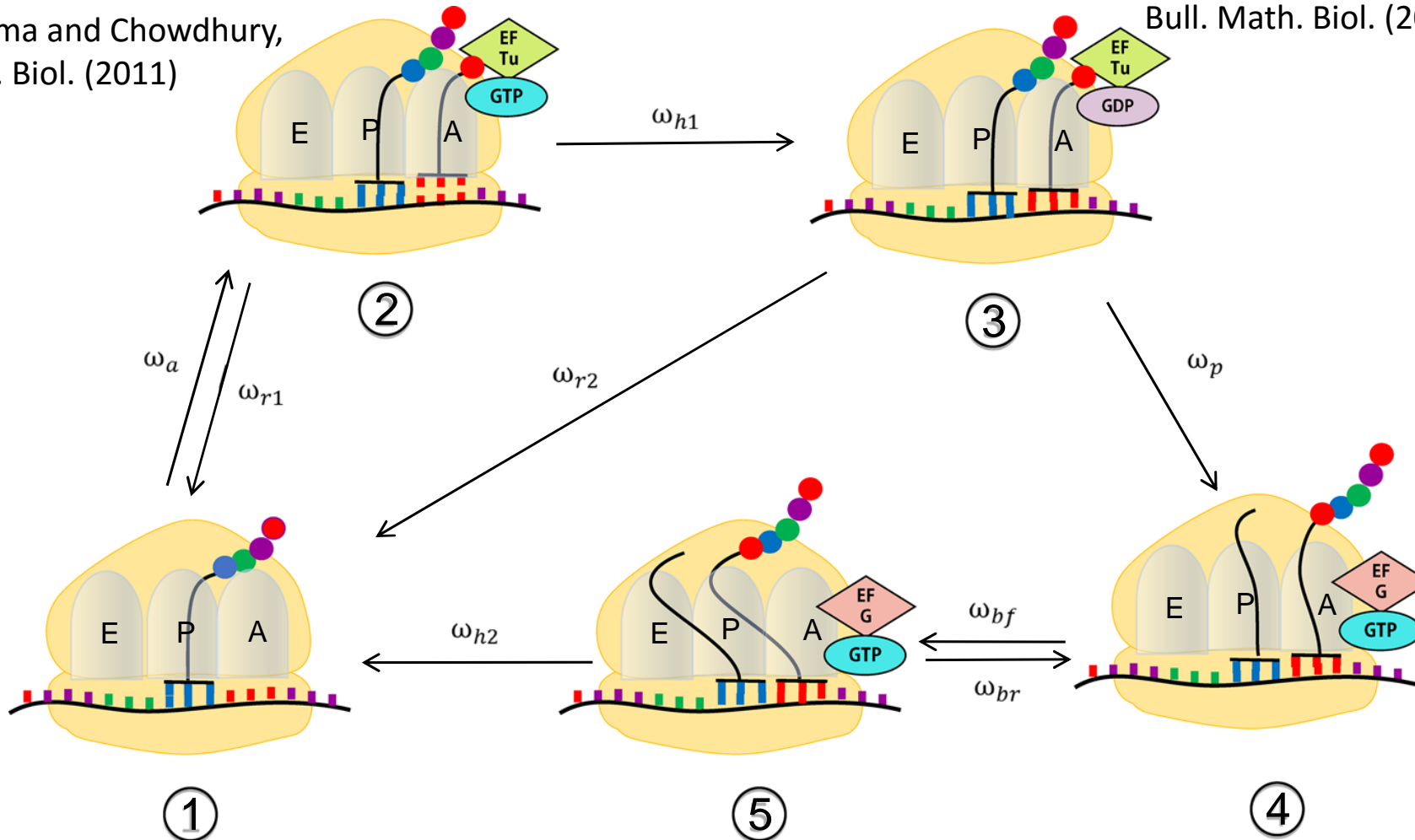
Sharma & Chowdhury, J. Theor. Biol. (2011)

The discrete mechano-chemical states of the machine form the vertices of a **network (graph)** while the directed edges denote the allowed transitions.

Machine operation, i.e., its stochastic kinetics, is modeled as a **Markov process** in a heat bath at a constant temperature and it is formulated in terms of **master equations**.

Sharma and Chowdhury,
Phys. Biol. (2011)

Dutta and Chowdhury,
Bull. Math. Biol. (2017)



Master equation: general form for “*States and Rates*”

$P_n(t)$ = Probability of finding the “particle” in the discrete state n at time t .

$$dP_n(t)/dt = \underbrace{\sum_m P_m(t) W(m \rightarrow n)}_{\text{GAIN terms}} - \underbrace{\sum_{m'} P_n(t) W(n \rightarrow m')}_{\text{Loss terms}}$$

GAIN terms

Loss terms

$W(n \rightarrow m)$ = Probability of transition $n \rightarrow m$ per unit time
 (“rate constant”).



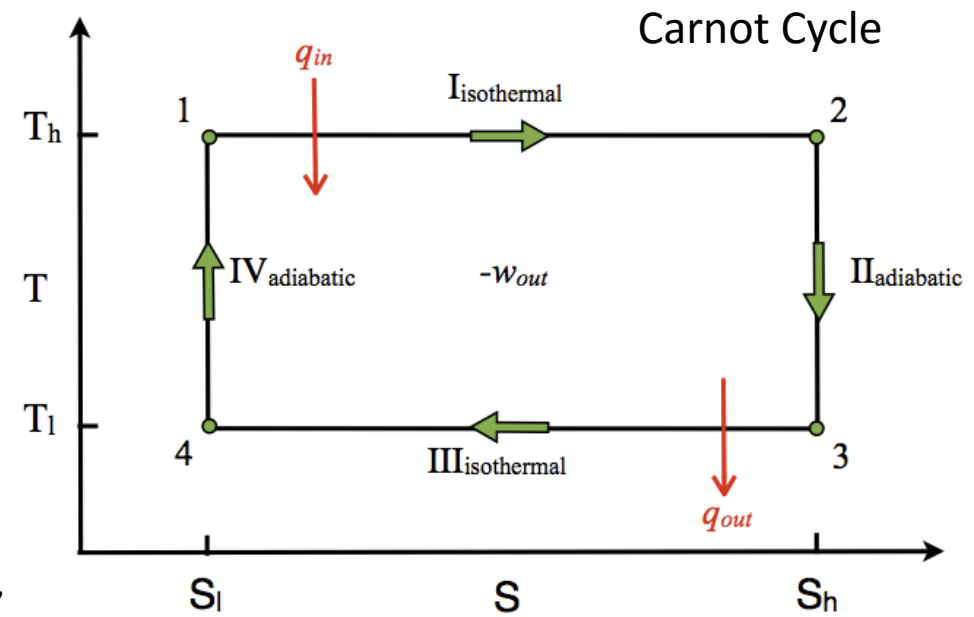
**Markov Process:
Frog in a Lily Pond**



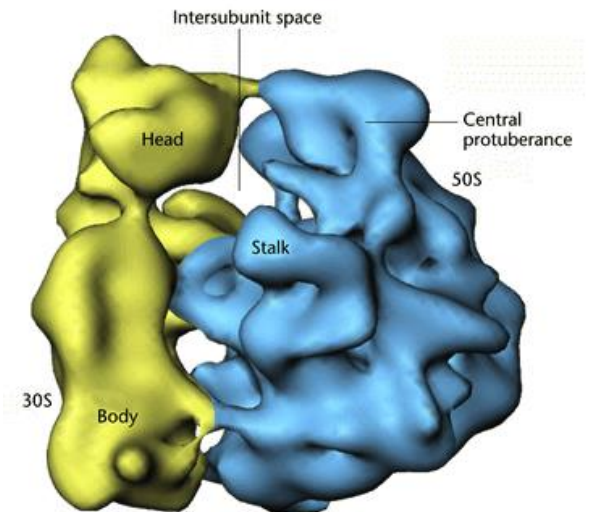
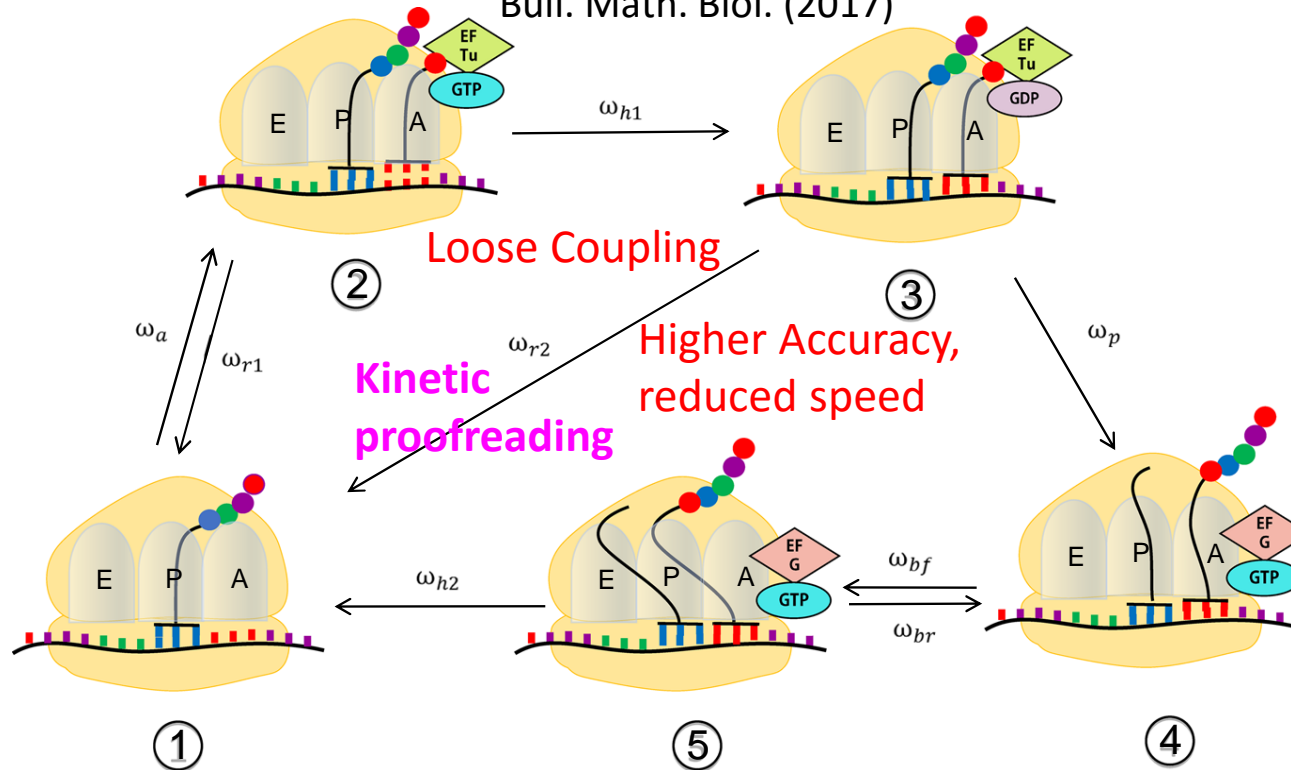
A. A. Markov



Sadi Carnot (1796-1832)
Source: wikipedia



Dutta and Chowdhury,
Bull. Math. Biol. (2017)



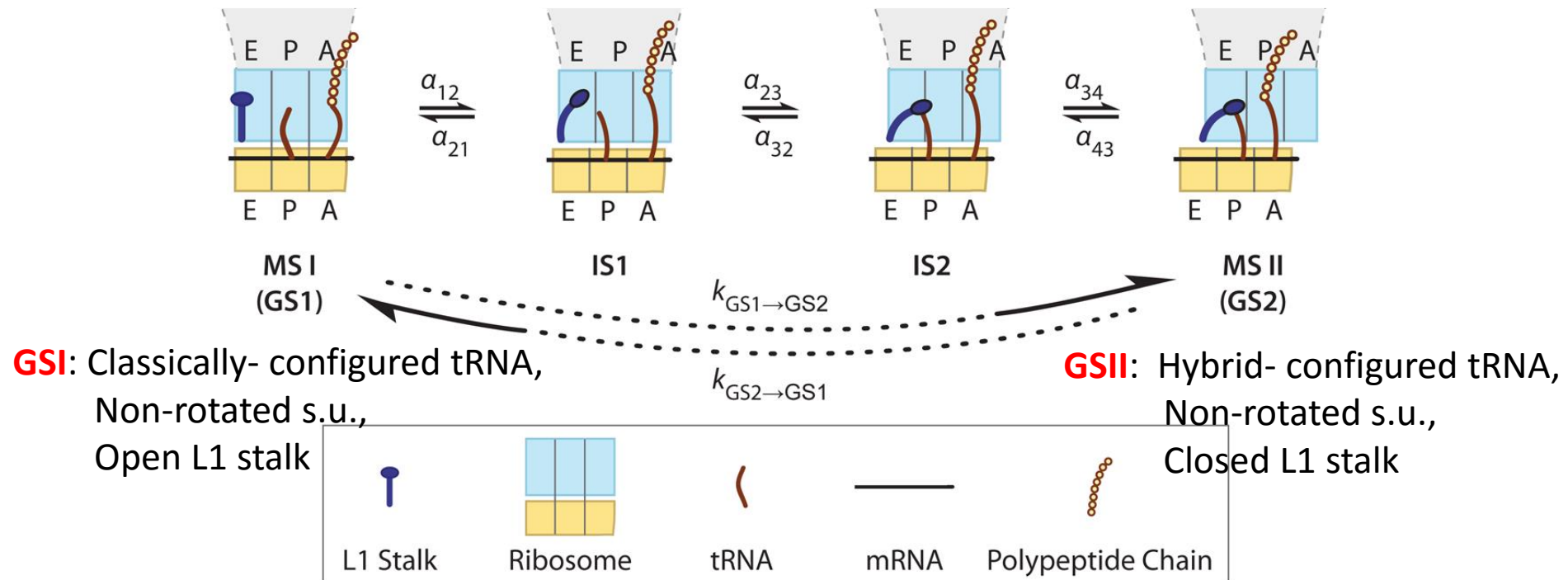
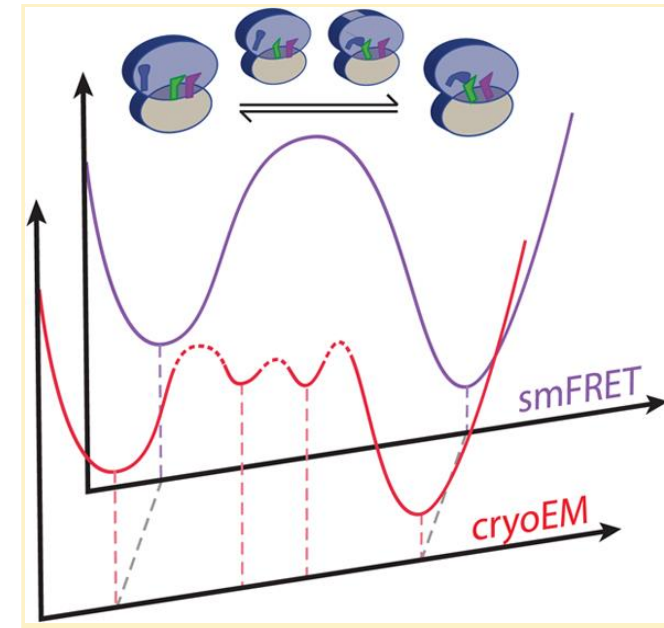
Ribosome

Quantitative Connection between Ensemble Thermodynamics and Single-Molecule Kinetics: A Case Study Using Cryogenic Electron Microscopy and Single-Molecule Fluorescence Resonance Energy Transfer Investigations of the Ribosome

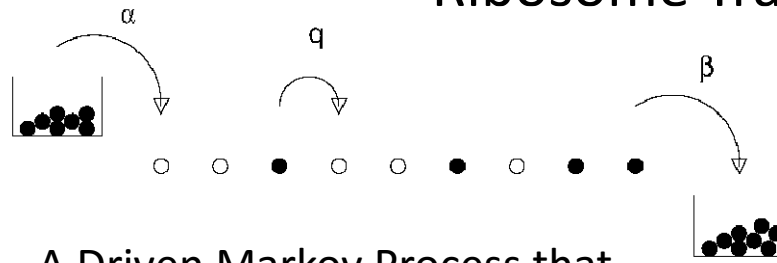
Colin D. Kinz-Thompson,^{†,||} Ajeet K. Sharma,^{‡,||,¶} Joachim Frank,^{⊥,§} Ruben L. Gonzalez, Jr.,[†] and Debashish Chowdhury^{*,‡}

THE JOURNAL OF
PHYSICAL CHEMISTRY B

Fluctuating kinetics of PRE until
EF-G-catalyzed Translocation



Ribosome Traffic on a single mRNA



A Driven Markov Process that reaches NON-Eq Steady State (NESS).

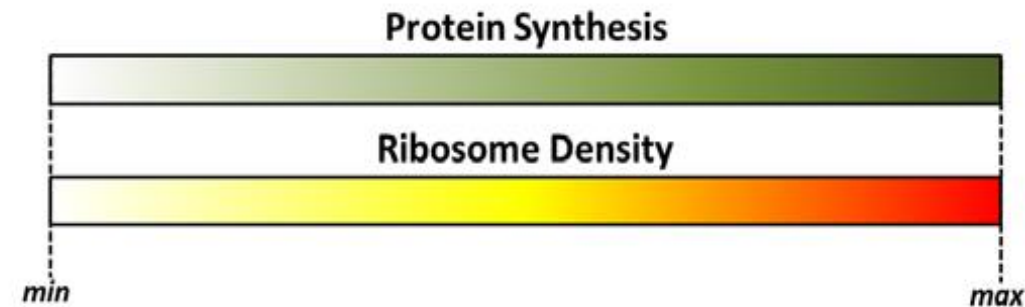
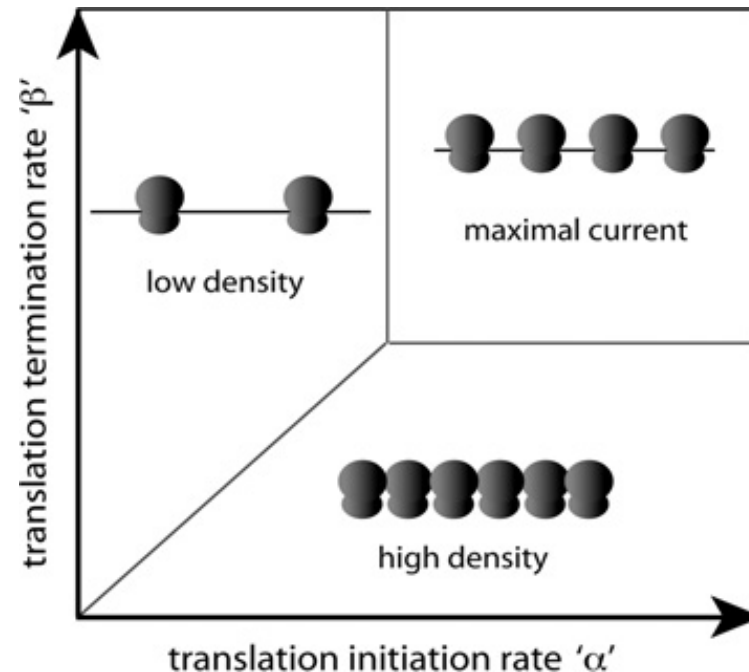
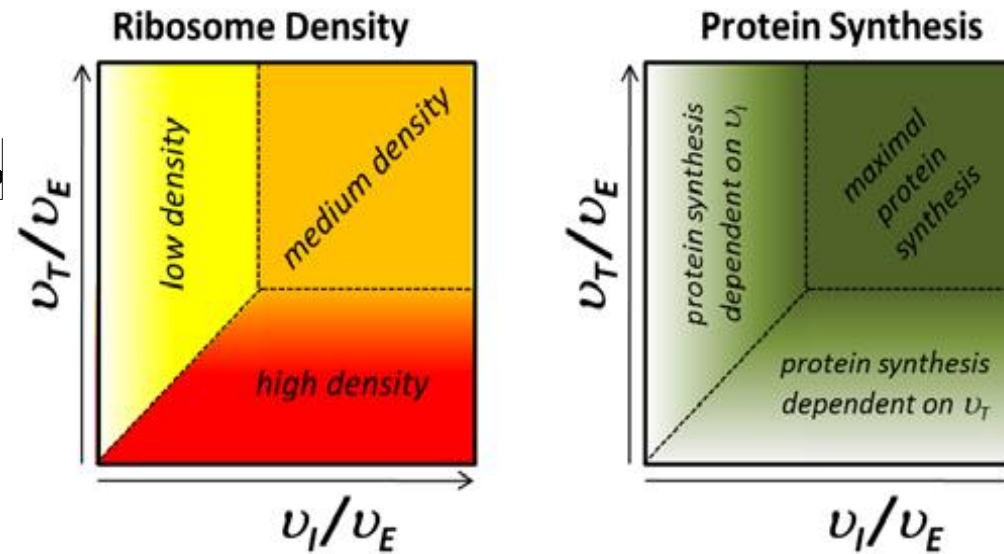
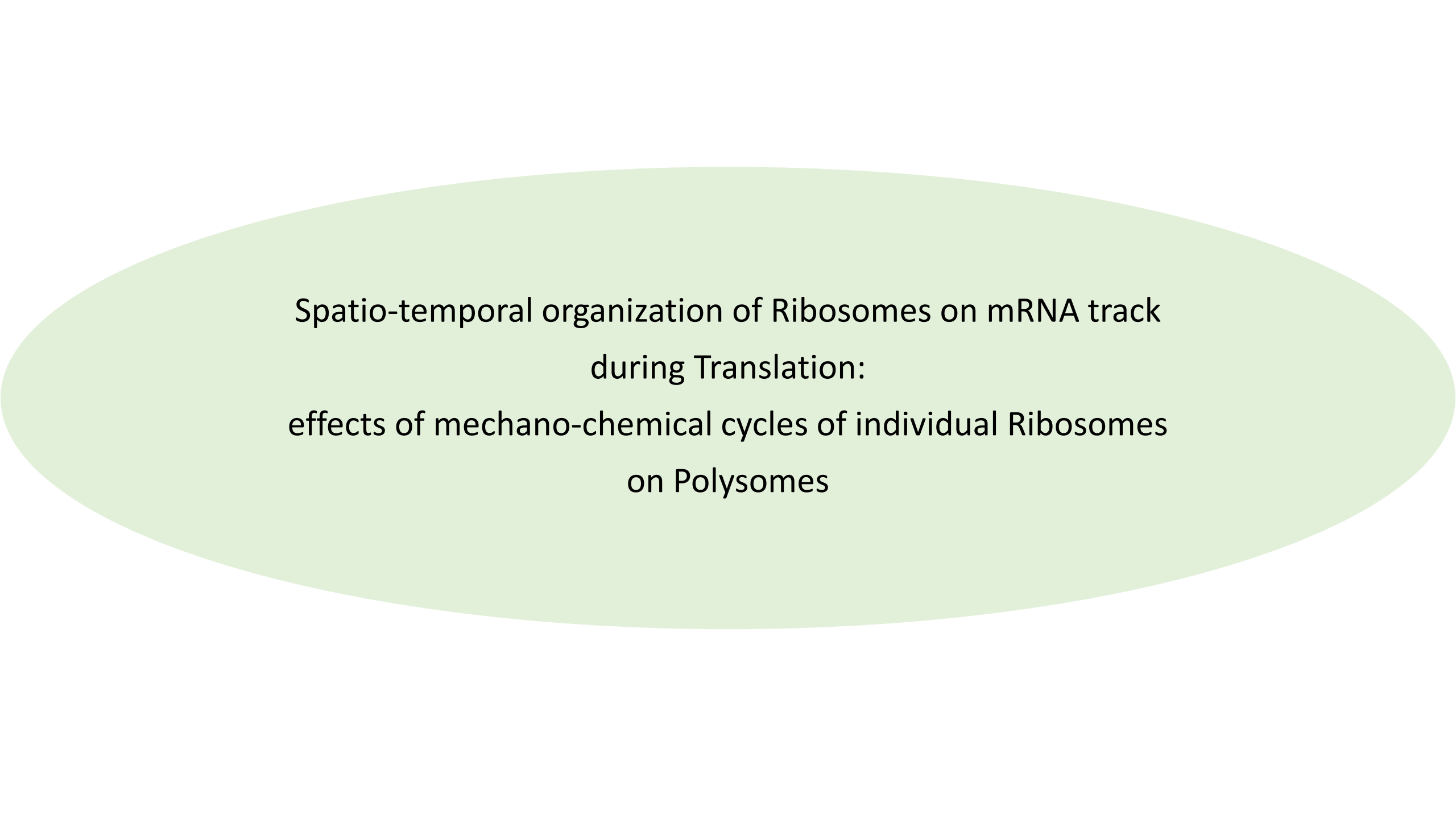


Figure 4. The relationship between ribosome density and protein synthesis, as predicted by TASEP models. The left panel shows ribosome densities as a function of different ratios of v_I, v_E and v_T (initiation, elongation and termination rates) under the standard TASEP assumptions of a uniform elongation rate and of unlimited ribosome supply. The right panel shows the corresponding protein synthesis rates.

Von der Haar, Comp. & Struct. Biotech. J. (2012)



Spatio-temporal organization of Ribosomes on mRNA track
during Translation:
effects of mechano-chemical cycles of individual Ribosomes
on Polysomes

A. Basu and D. Chowdhury, Phys. Rev. E 75, 021902 (2007)

Garai, Chowdhury and Ramakrishnan, *Phys. Rev. E*, 79, 011916 (2009)

Garai, Chowdhury, Chowdhury and Ramakrishnan, Phys. Rev. E 80, 011908 (2009)

Sharma and Chowdhury, Phys. Rev. E, 82, 031912 (2010)

Sharma and Chowdhury, Physical Biology, 8, 026005 (2011).

Sharma and Chowdhury, Journal of Theoretical Biology, 289, 36 (2011).

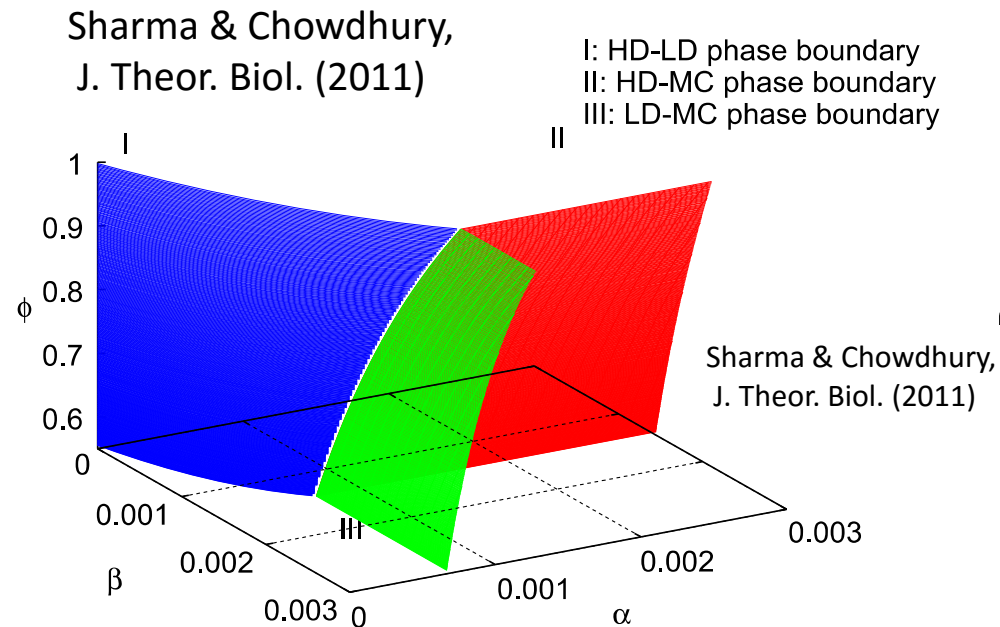


Fig. 2. Phase diagram of ribosome traffic model in the 3-dimensional space spanned by α, β and ϕ .

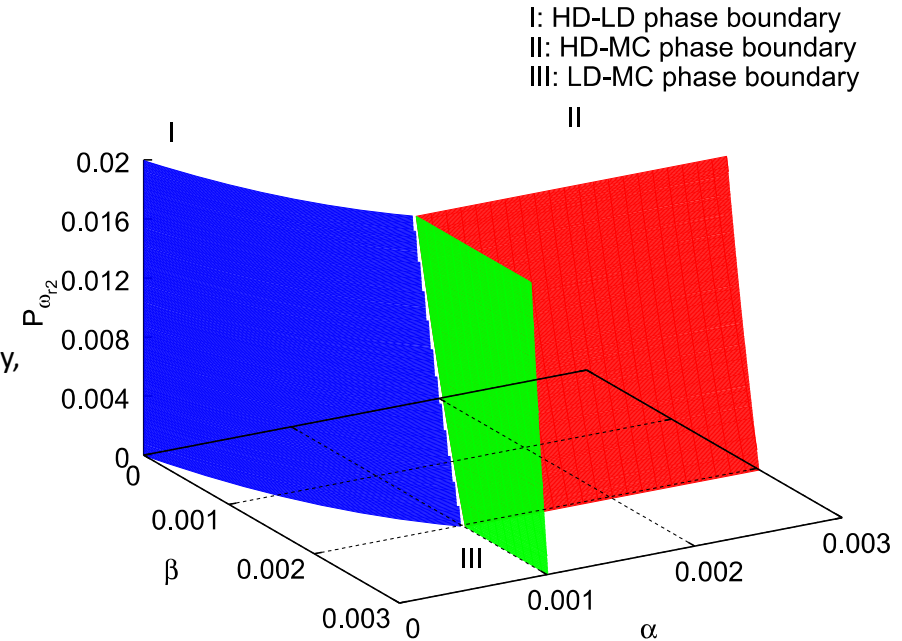



Fig. 4. Phase diagram of ribosome traffic model in the 3-dimensional space spanned by α, β and P_{ω_2} .



Regulation of Ribosome Traffic on mRNA template and
Control of Unconventional Translation:
Consequences of Programmed Error and Recoding

“Unlike DNA, which universally adopts a double helical conformation, RNA has extensive intramolecular interactions that cause it to fold into an array of complex structures. RNA structure is highly dynamic and is governed by factors such as temperature, cellular energy state, RNA structures enable a myriad of functions, which include encoding genetic information.....”

Lewis et al., Nat. Rev. Mol. Cell Biol. (2017).

Review

CellPress

Gene regulation by structured mRNA elements

Andreas Wachter

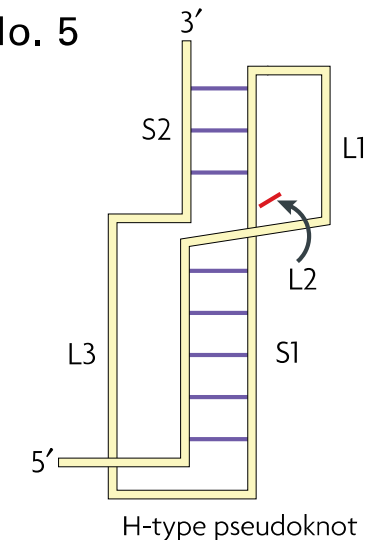
Trends in Genetics, May 2014, Vol. 30, No. 5

Viral RNA pseudoknots: versatile motifs in gene expression and replication

Ian Brierley*, Simon Pennell† and Robert J. C. Gilbert§

www.nature.com/reviews/micro

AUGUST 2007 | VOLUME 5



Next we explore the regulatory roles of the secondary structures of mRNA on Ribosome Traffic

Accuracy

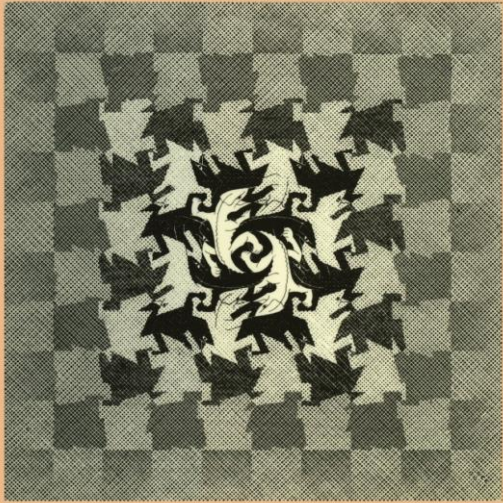
in

Molecular Processes

ITS CONTROL AND RELEVANCE TO LIVING SYSTEMS

Edited by

T.B.L. KIRKWOOD,
R.F. ROSENBERGER and D.J. GALAS

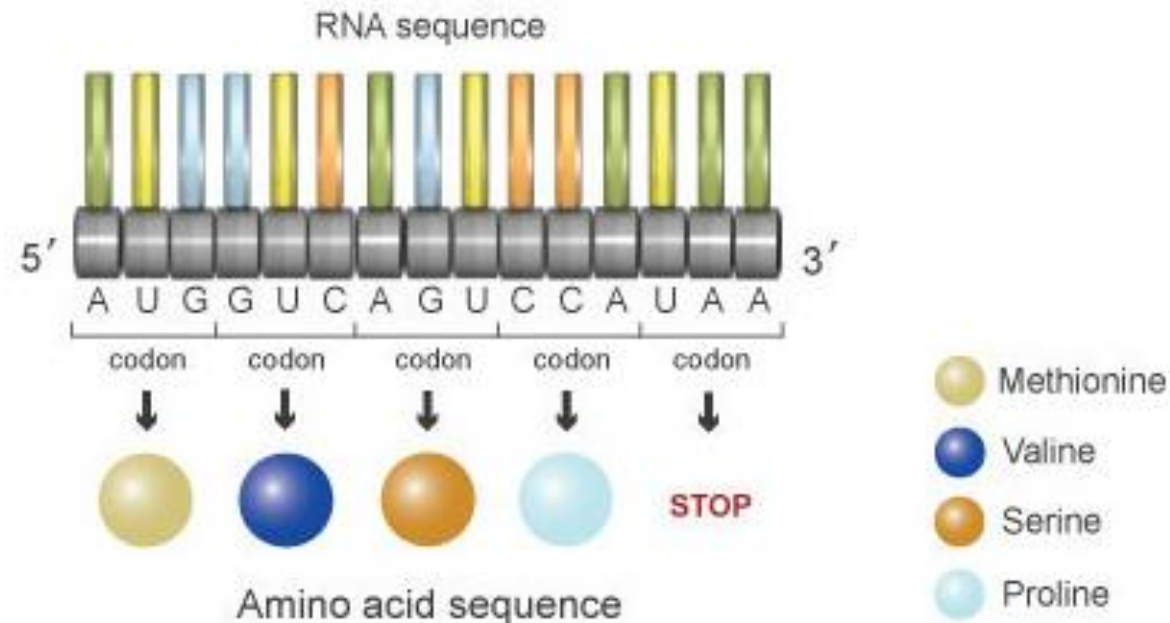


Accuracy

Fidelity and infidelity

“Nature does not exhaust itself for the sake of fidelity and perfectionism. Rather, errors are made, often repaired or discarded, but always tested as the source of blind innovation during the continuous adaptation to unpredictable environmental changes and challenges.”

For polymerase motors,
accuracy is as important,
if not more, as the **speed**.



Miroslav Radman

Nature, 413, 115 (2001)

Molecular evolution

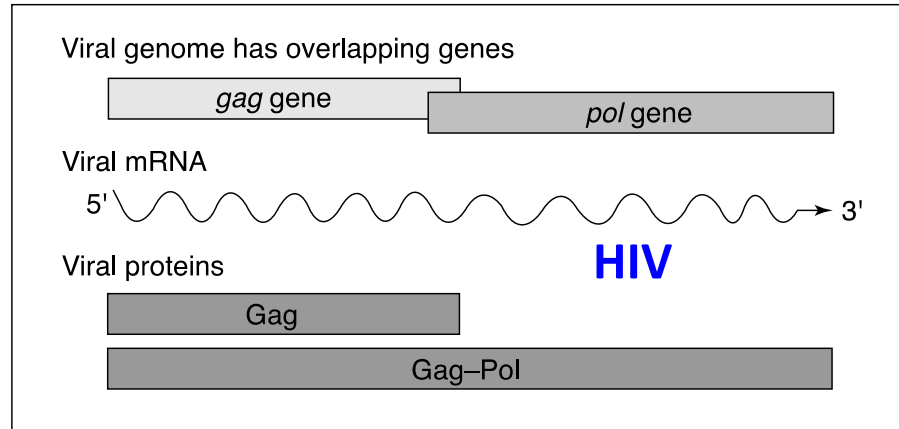
Errors and infidelity, even wastefulness, can cause individual failure, but they are also a source of innovation and robustness, ensuring the perpetuation of life.

Recoding

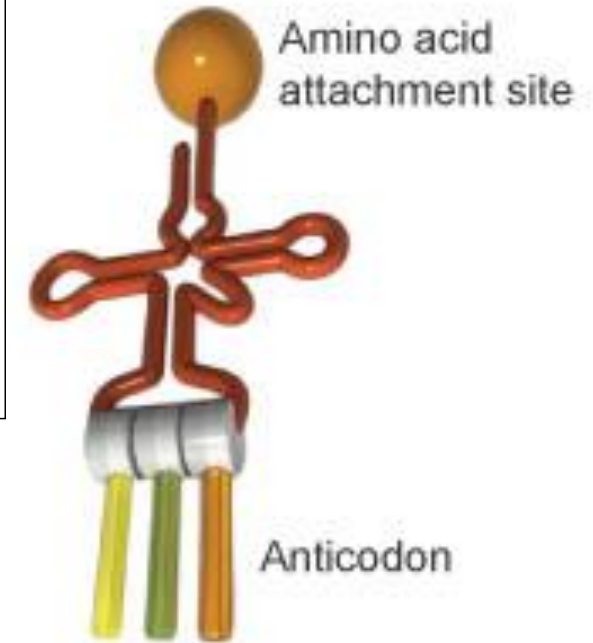
Expansion of Decoding Rules Enriches
Gene Expression

"Programmed" Errors: Unconventional Translation

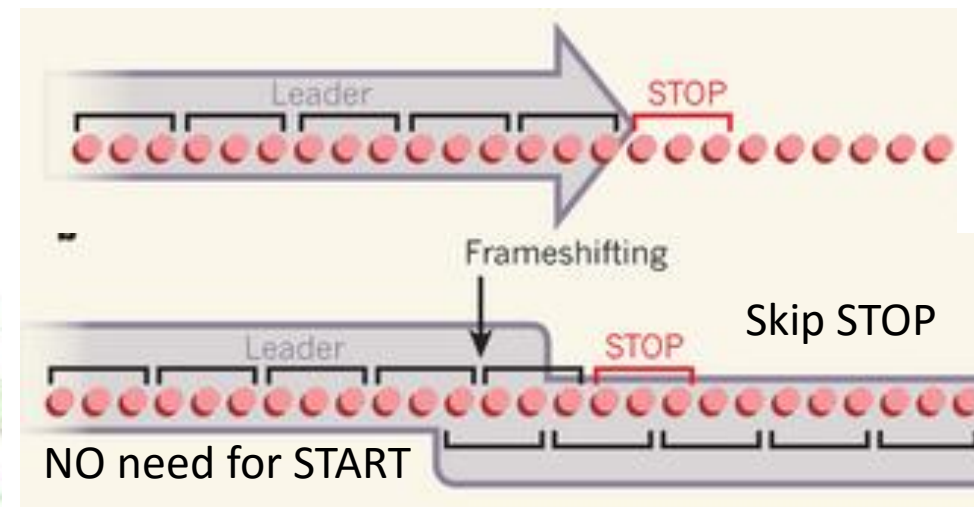
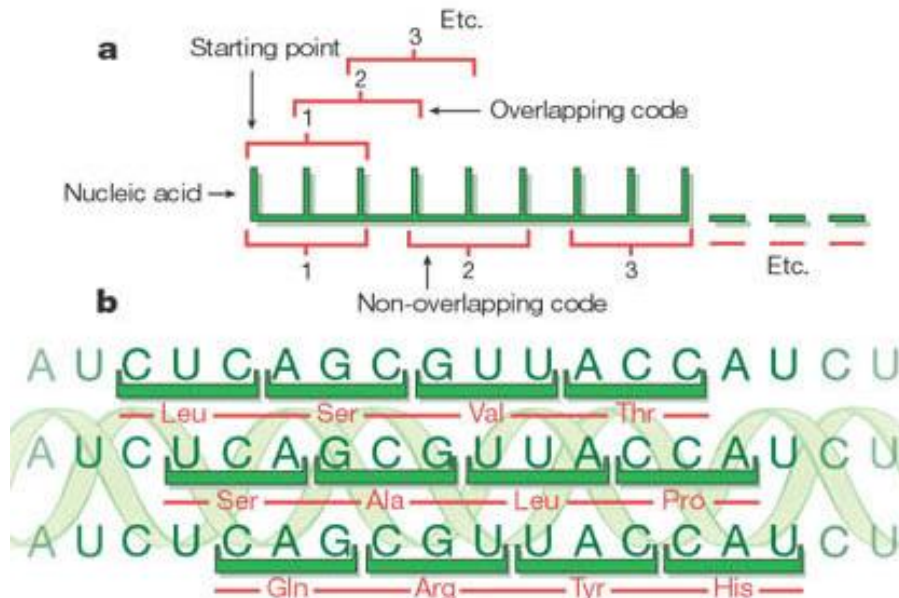
From: Nature Education



Dinman et al.
TIBSTECH (1998)

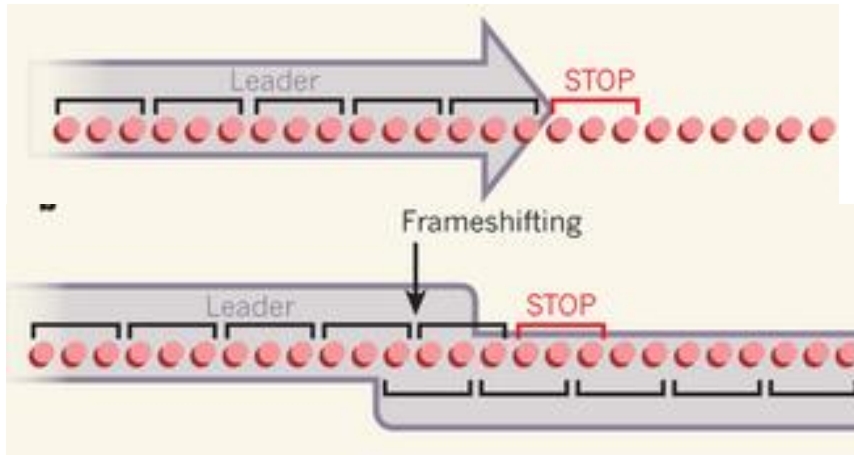


Atkins & Gesteland, Nature 414, 693 (2001)

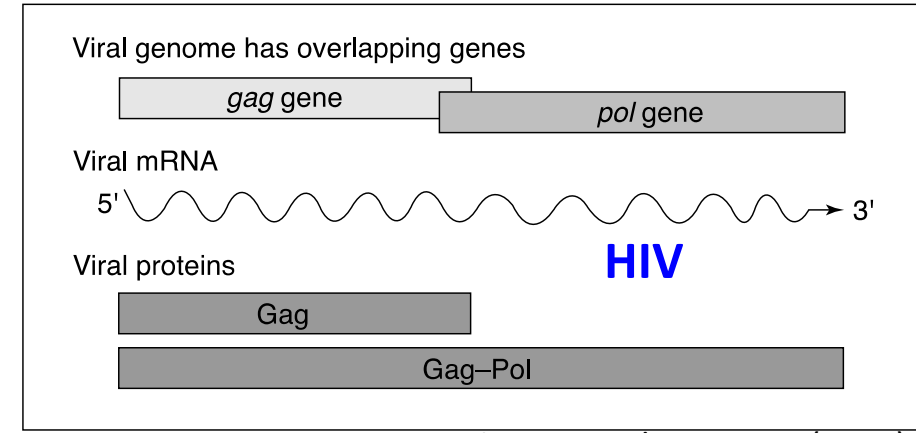


Atkins & Baranov, Nature 503, 478 (2013)

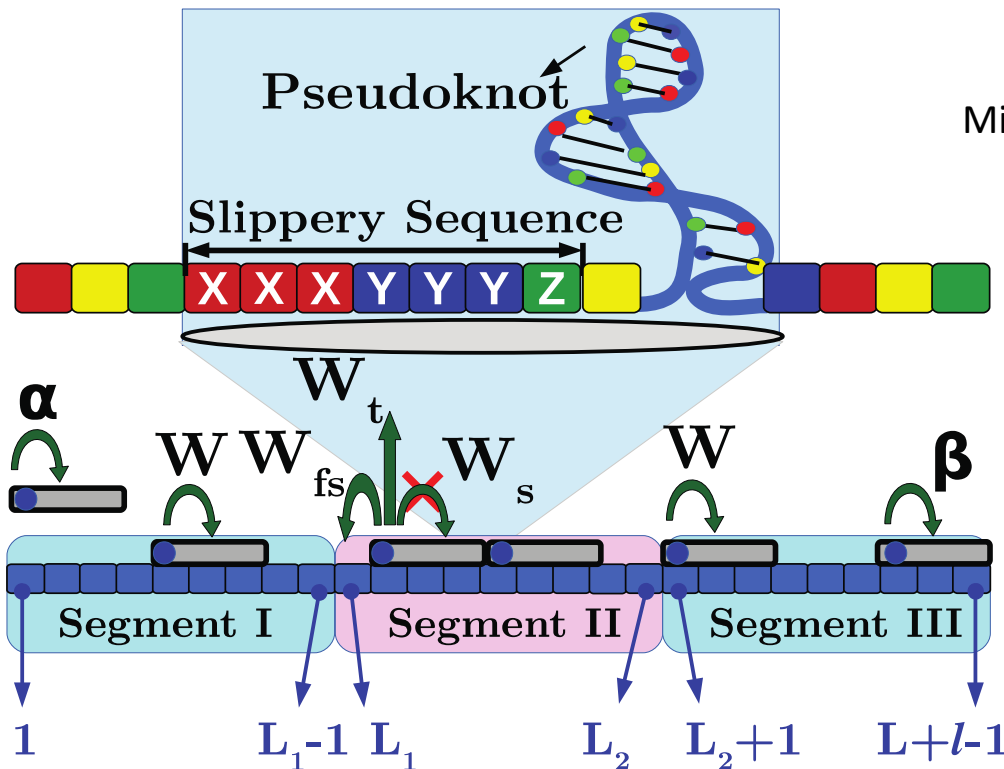
Non-canonical Elongation: Frameshifting



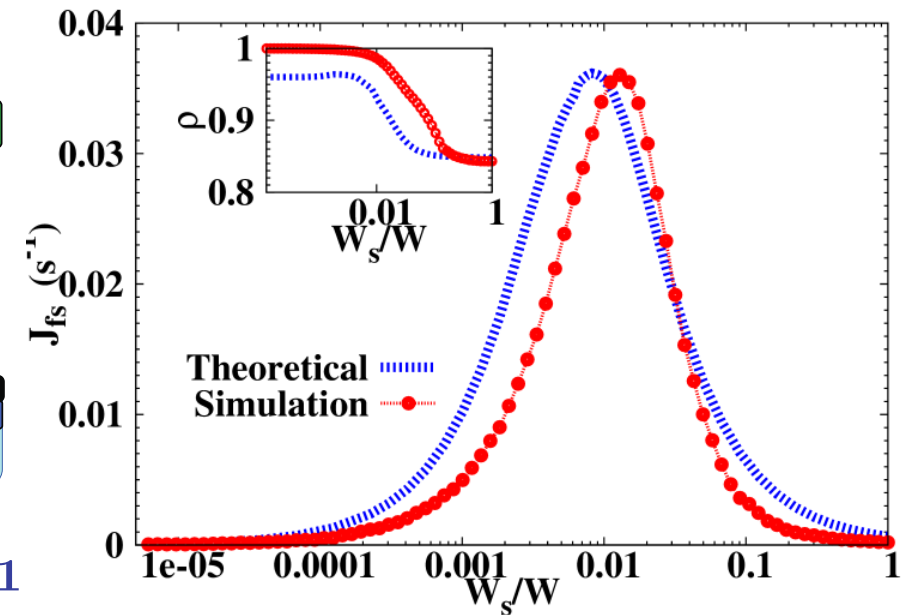
Atkins & Baranov, Nature 503, 478 (2013)



Dinman et al. TIBSTECH (1998)



Mishra, Schutz & Chowdhury EPL 114, 68005 (2016)



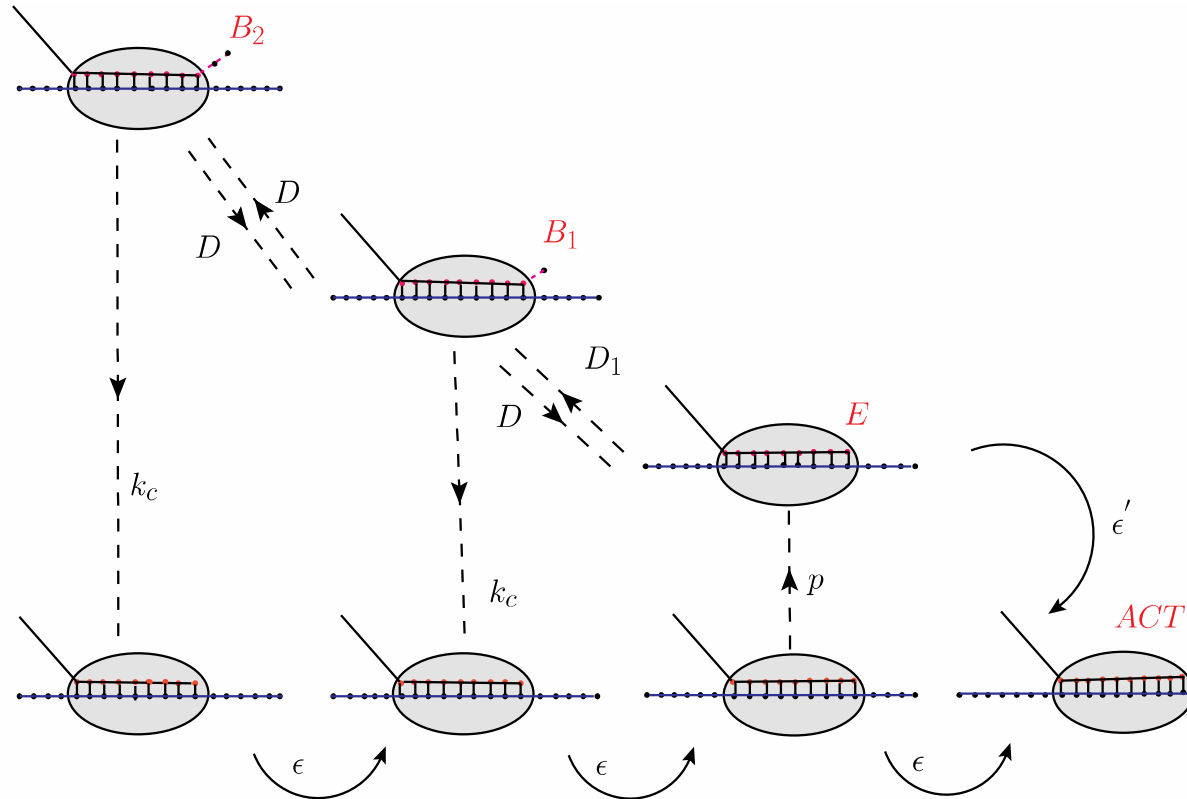
EPL, **96** (2011) 60004

doi: 10.1209/0295-5075/96/60004

Transcriptional proofreading in dense RNA polymerase traffic

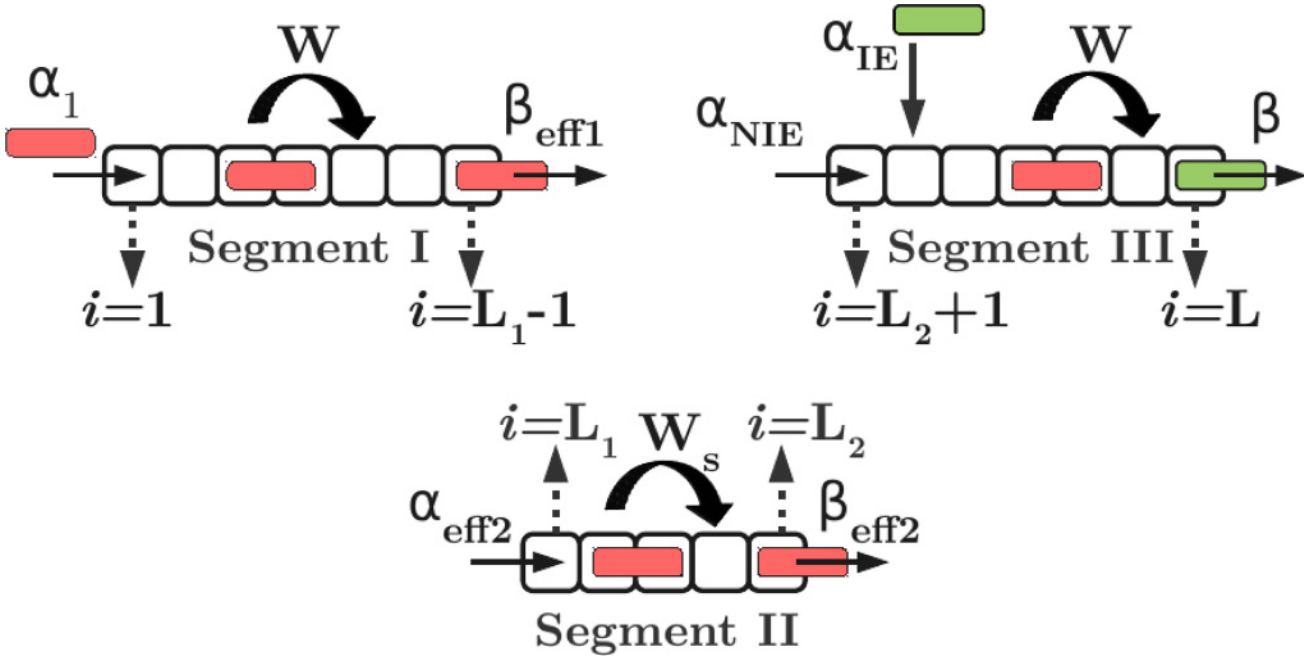
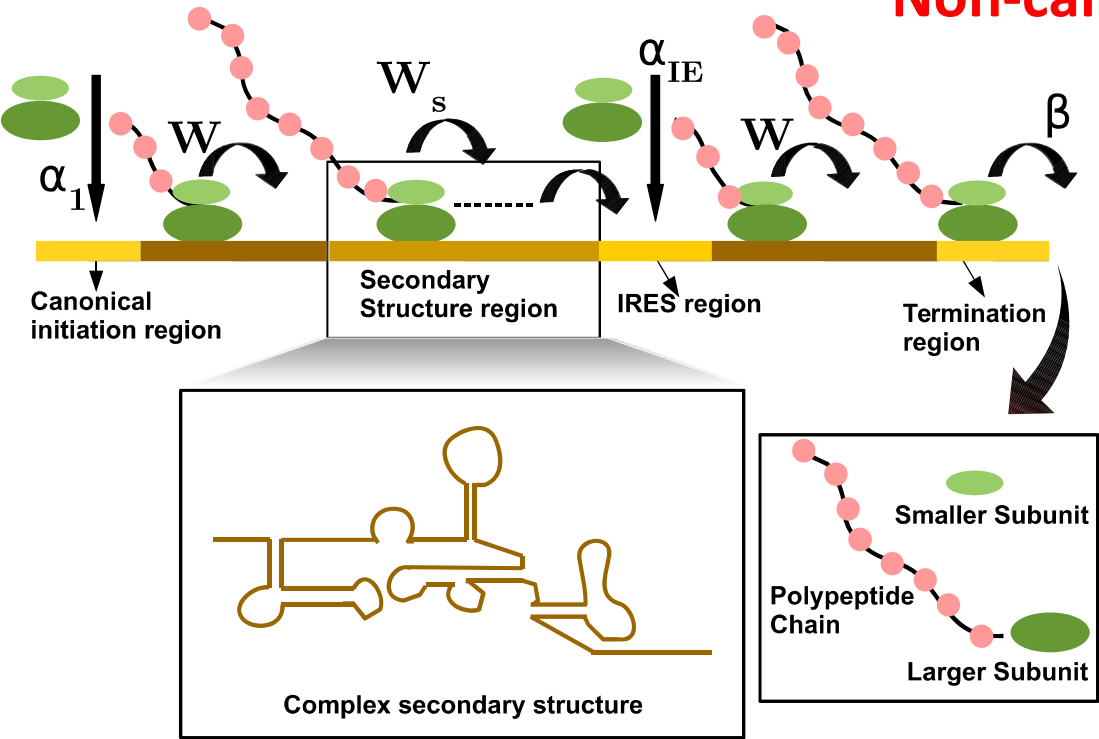
MAMATA SAHOO and STEFAN KLUMPP^(a)

Max Planck Institute of Colloids and Interfaces, Science Park Golm - 14424 Potsdam, Germany, EU



Abstract – The correction of errors during transcription involves the diffusive backward translocation (backtracking) of RNA polymerases (RNAPs) on the DNA. **A trailing RNAP on the same template can interfere with backtracking as it progressively restricts the space that is available for backward translocation and thereby ratchets the backtracked RNAP forward. We analyze the resulting negative impact on proofreading** theoretically using a driven lattice gas model of transcription under conditions of dense RNAP traffic.

Non-canonical Initiation: IRES



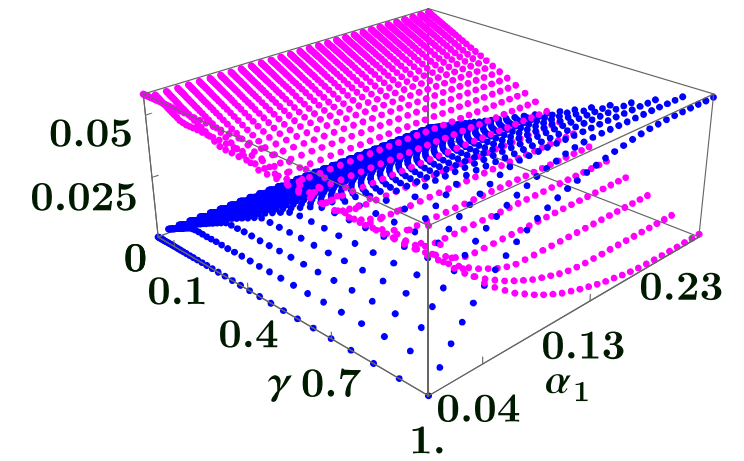
Mishra and Chowdhury, Phys. Rev. E (2017)

TABLE II. The effective rate constants and their defining expressions.

Rate constant	Expression
α_{eff2}	$\alpha_{\text{eff2}} = W\rho_{c1}/\ell$
α_{NIE}	$\alpha_{\text{NIE}} = W_s\rho_{c2}/\ell$
α_{eff3}	$\alpha_{\text{eff3}} = \alpha_{\text{NIE}} + \alpha_{\text{IE}}$
β_{eff1}	$\beta_{\text{eff1}} = W P_1(\underline{L_1 - 1} L_1 + \ell - 1)$
β_{eff2}	$\beta_{\text{eff2}} = W_s P_1(\underline{L_2} L_2 + \ell)$

Internal Ribosome Entry Site (IRES):
An Unconventional mode of Translation

Translational Interference?

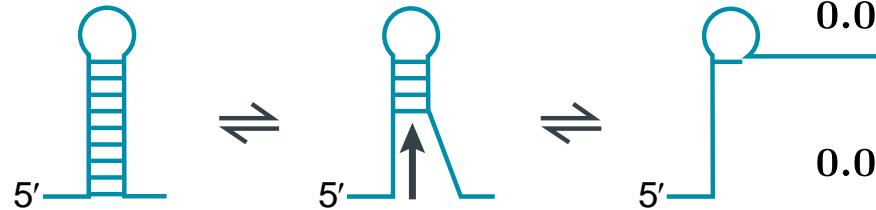


Bacterial RNA thermometers

(c)
 --- J_1 (MC Simulation)
 --- J_2 (MC Simulation)

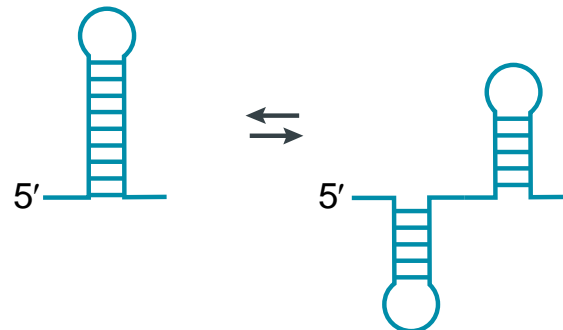
a Zipper

Low temperature High temperature Higher temperature

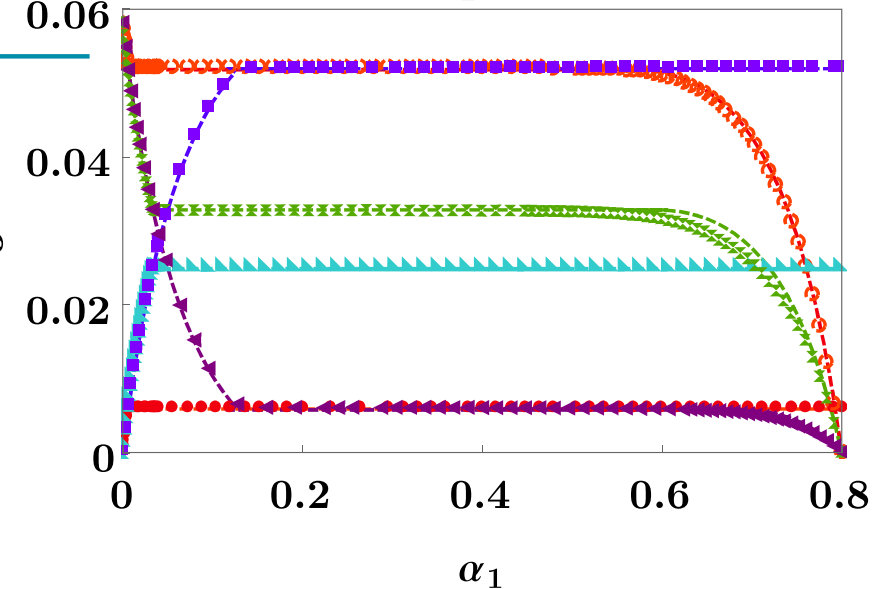


b Switch

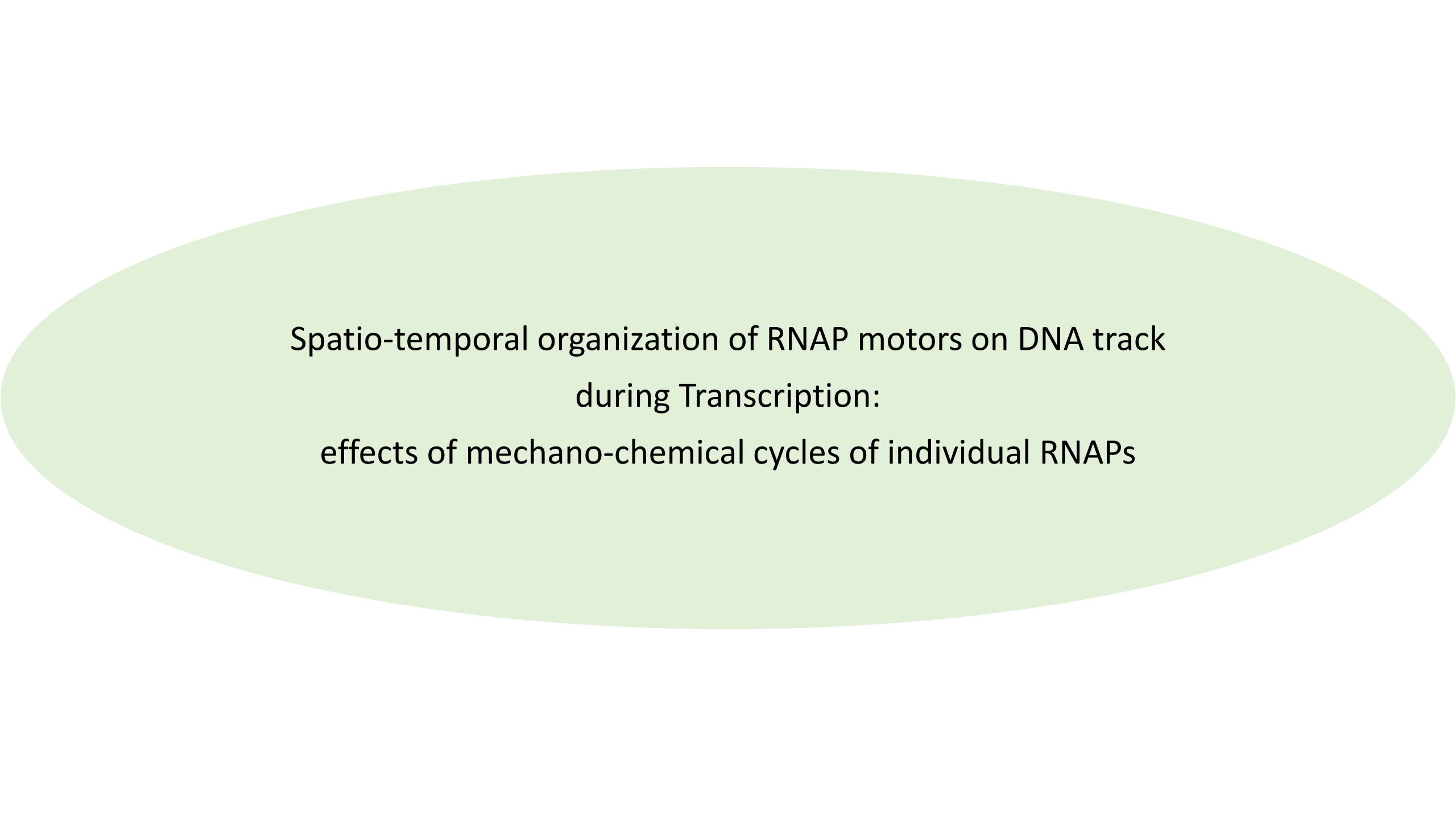
Temperature 1 Temperature 2



(a)
 --- J_1 (MC Simulation)
 --- J_2 (MC Simulation)



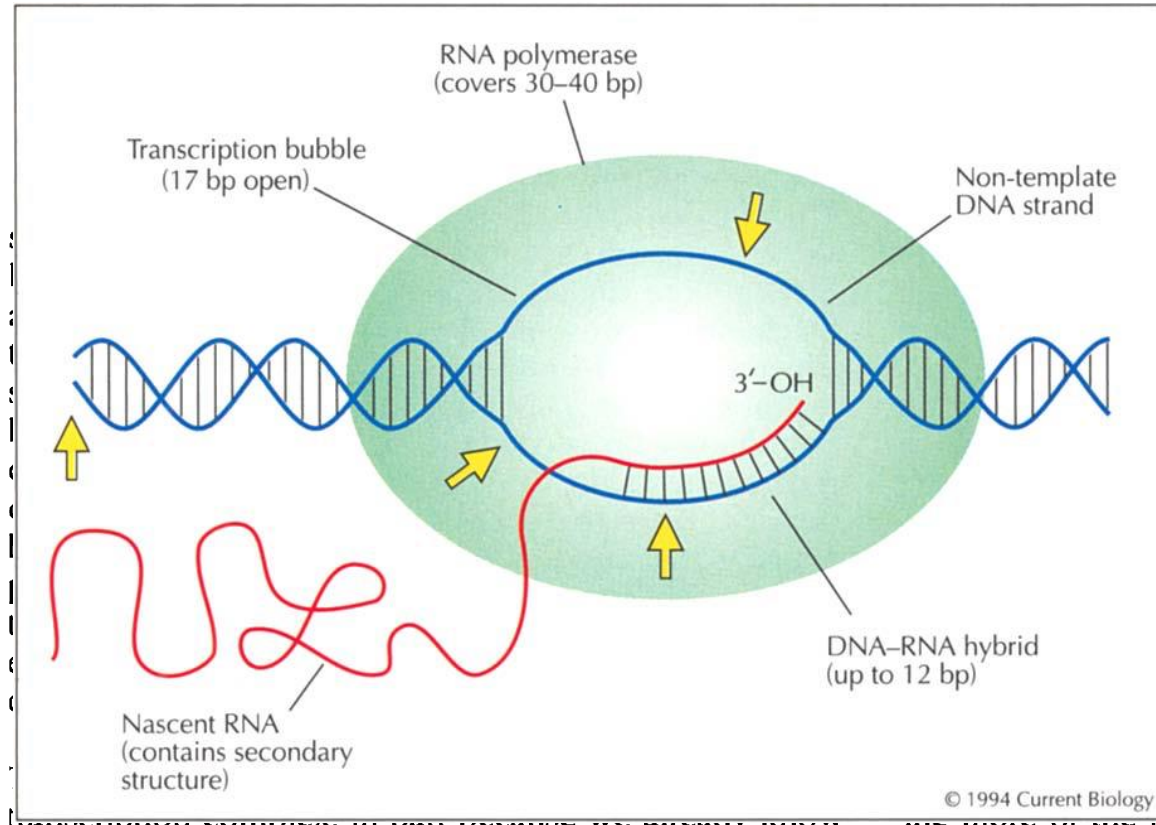
• $J_1(\gamma=0.1)$ ◊ $J_2(\gamma=0.1)$
 ▲ $J_1(\gamma=0.4)$ ▼ $J_2(\gamma=0.4)$
 ■ $J_1(\gamma=0.9)$ ◀ $J_2(\gamma=0.9)$



Spatio-temporal organization of RNAP motors on DNA track
during Transcription:
effects of mechano-chemical cycles of individual RNAPs

Theoretical model of RNAP and RNA synthesis

T. Tripathi and D. Chowdhury, Phys. Rev. E 77, 011921 (2008)

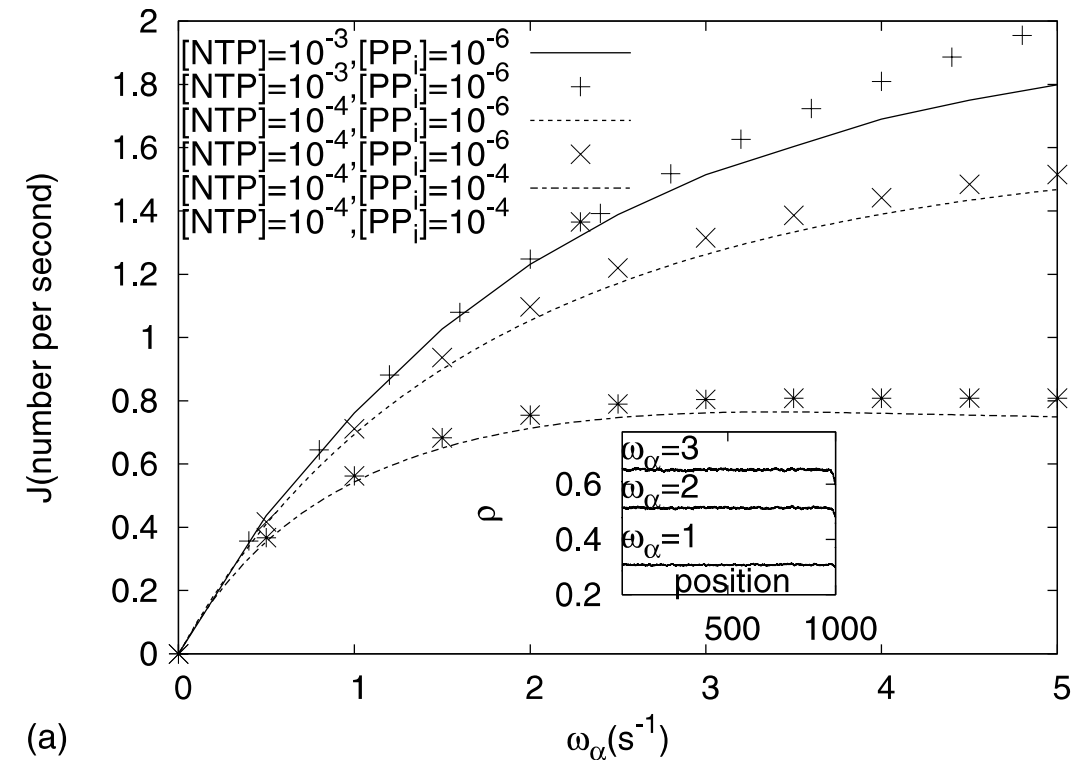
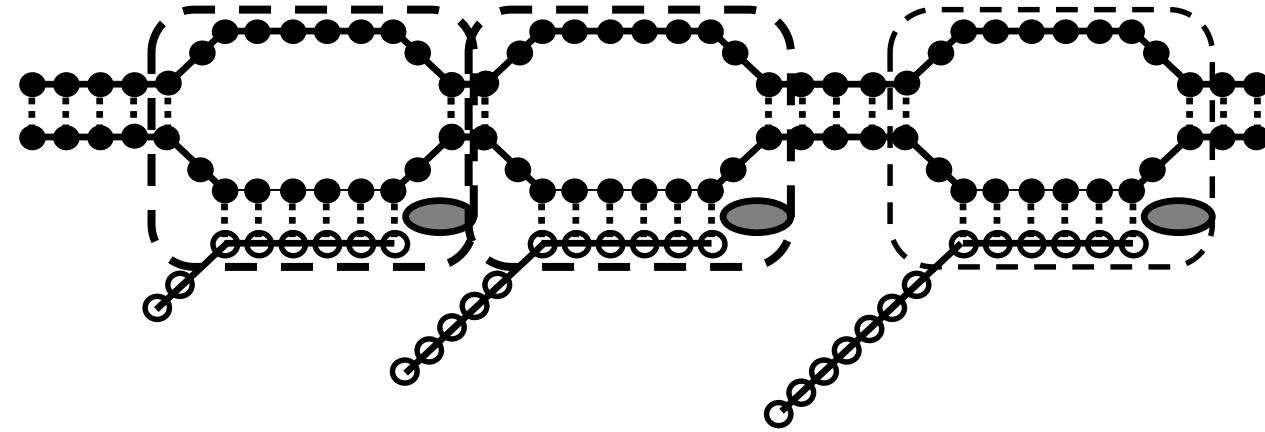


Transcription-elongation complex (TEC)

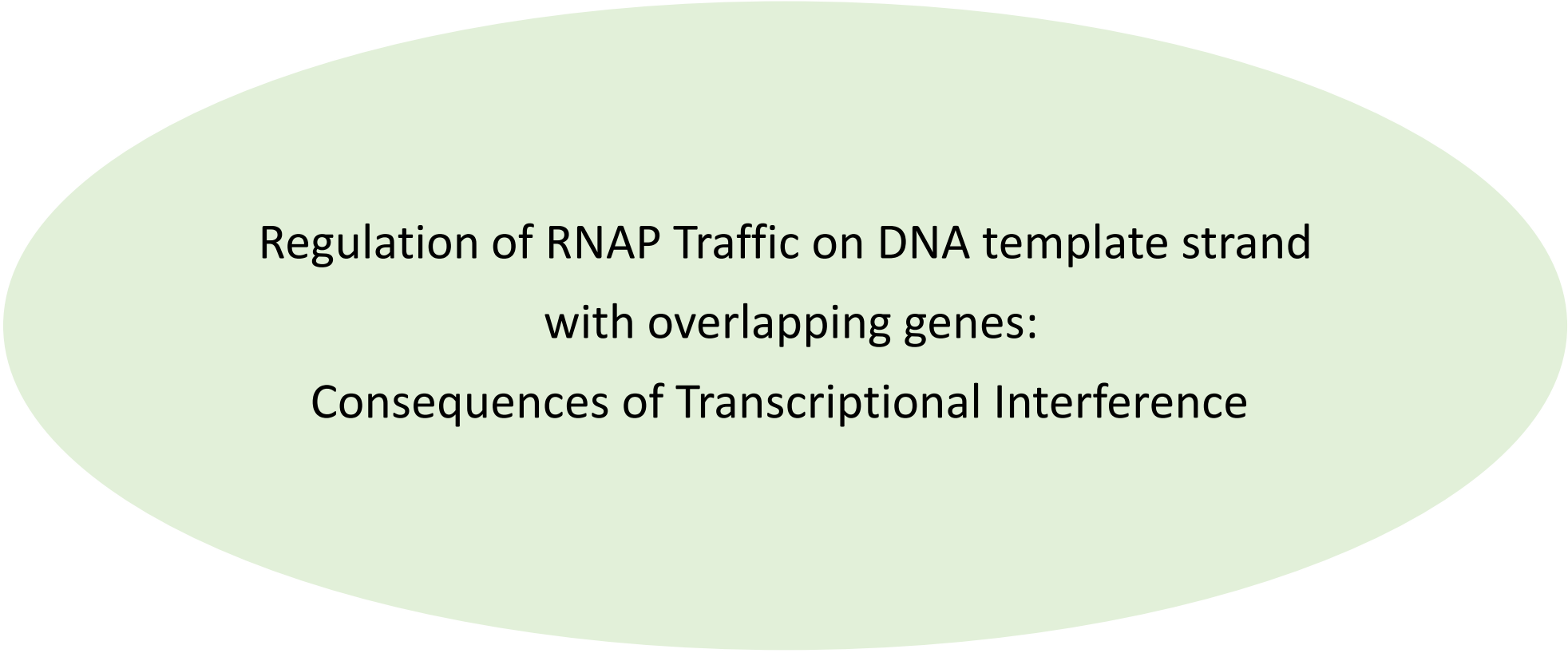
= RNAP + DNA template

+ mRNA transcript

Mechano-chemistry of each RNAP
+ Steric interactions



(a)

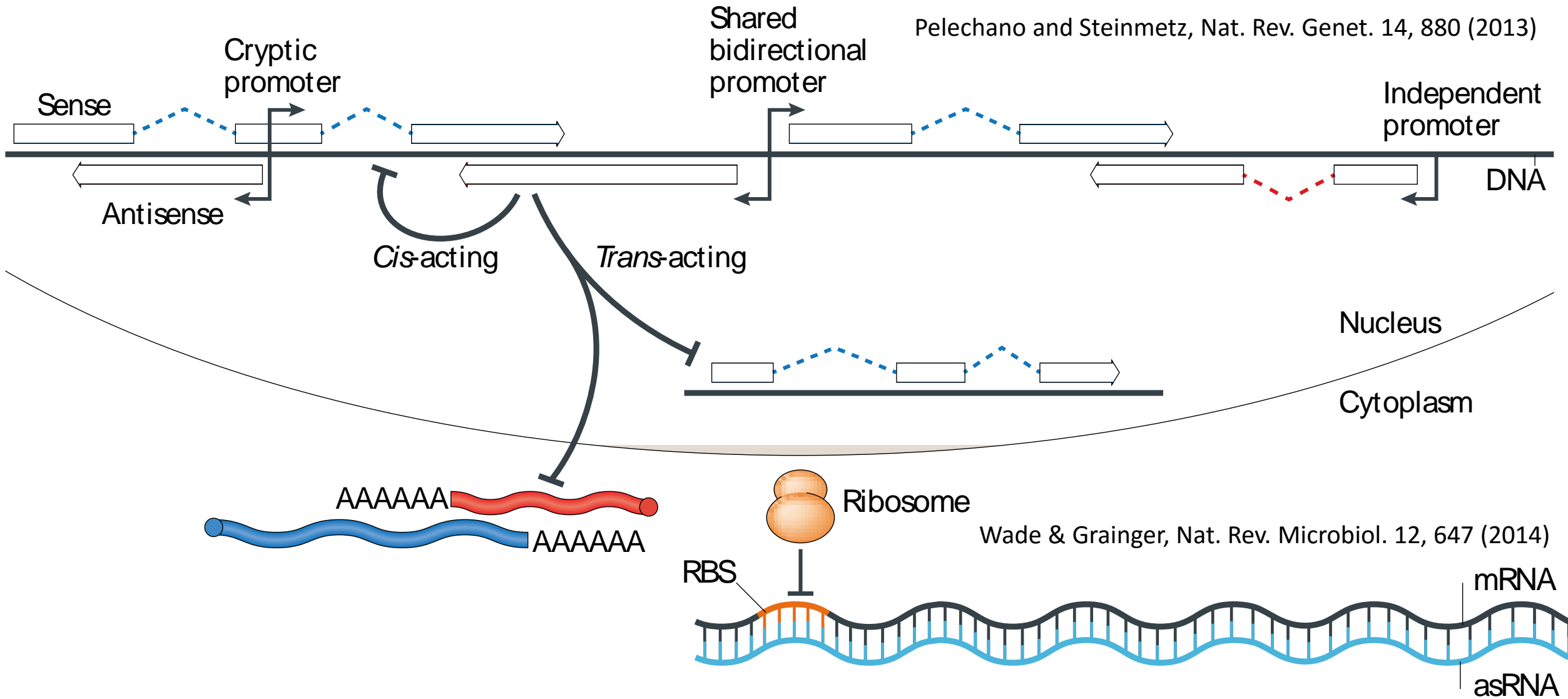


Regulation of RNAP Traffic on DNA template strand
with overlapping genes:

Consequences of Transcriptional Interference

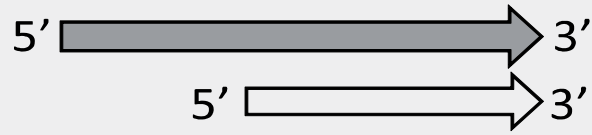
- Antisense transcripts are transcribed from the strand opposite to that of the sense transcript.
- Like transcription factors, these are also regulators of gene expression.
- These can establish on-off (bi-stable) switches.

Classification of Anti-Sense Transcripts



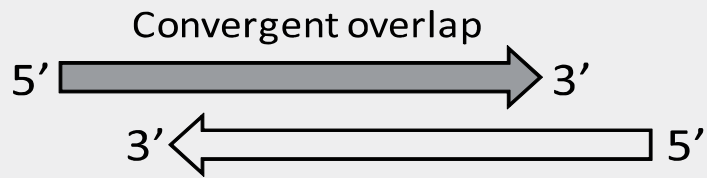
Overlapping/Nested Genes

A

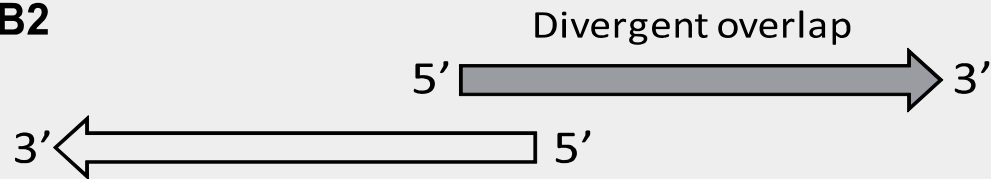


PARALLEL OVERLAP

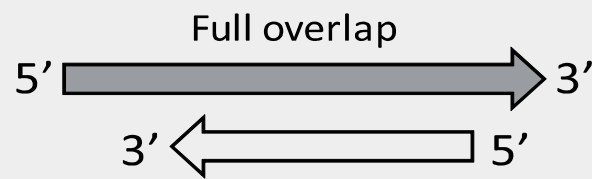
B1



B2



B3

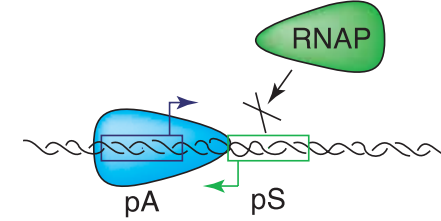


ANTIPARALLEL OVERLAPS

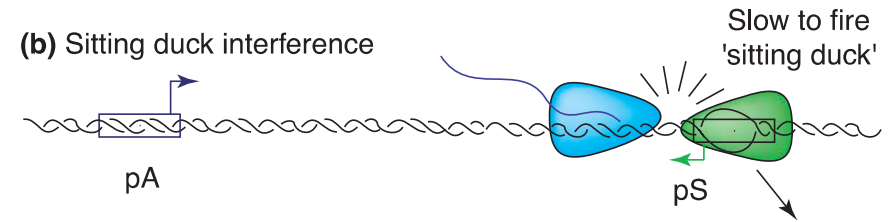
Z. Boldogkoi, Front. Genet. (2012)

Co-Transcriptional effects of antisense transcription: Transcriptional Interference

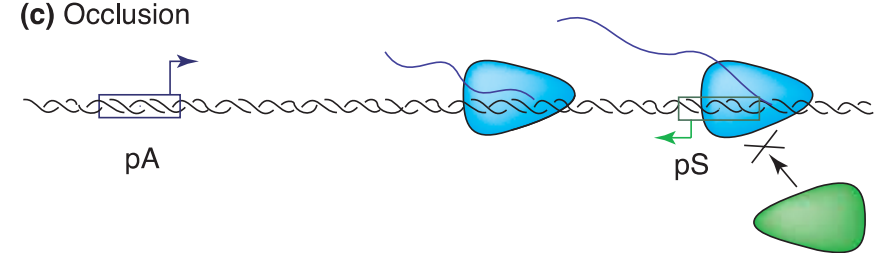
(a) Promoter competition



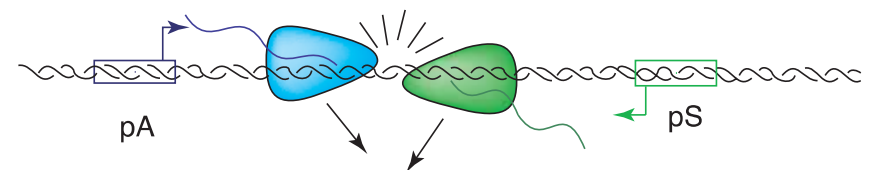
(b) Sitting duck interference



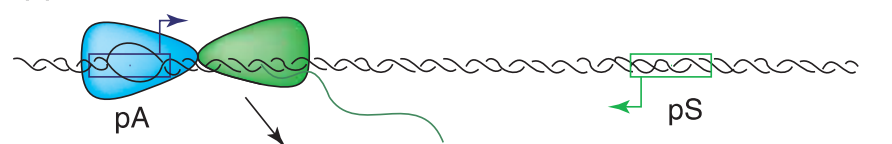
(c) Occlusion



(d) Collision



(e) Roadblock

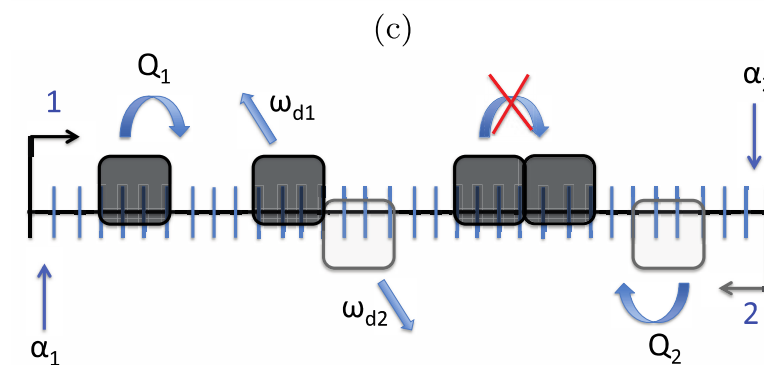
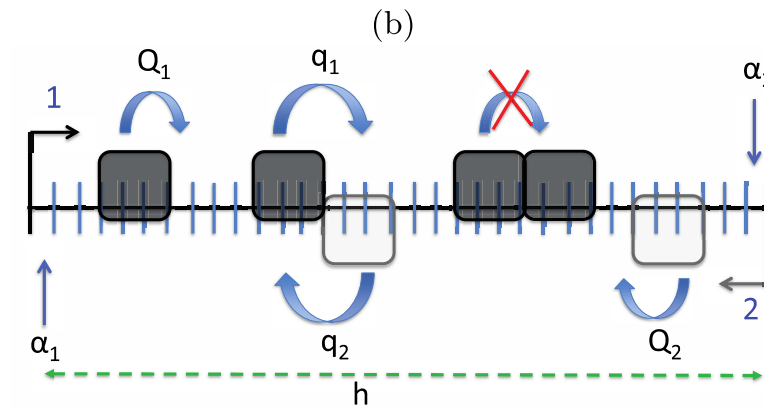
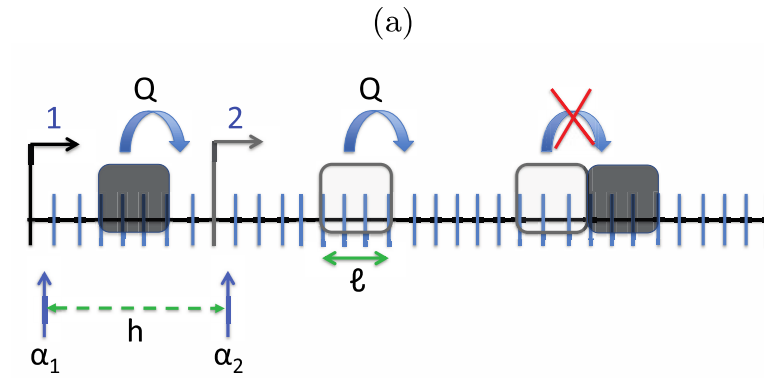
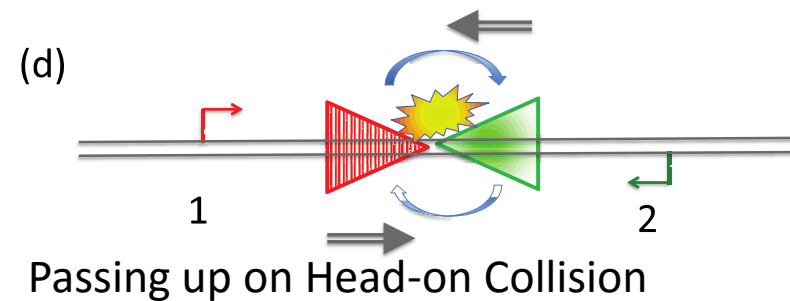
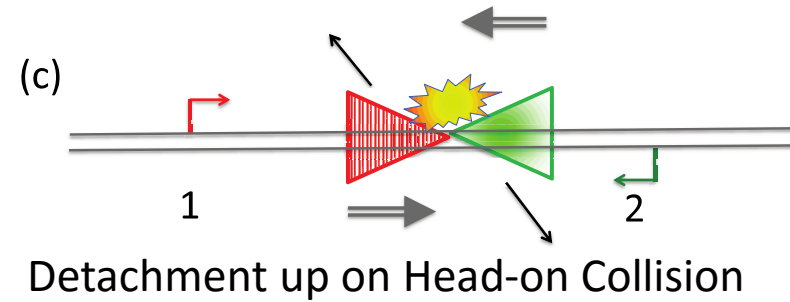
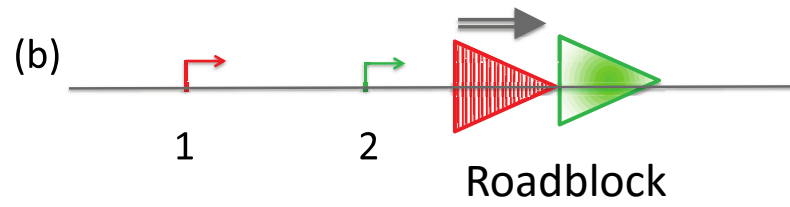
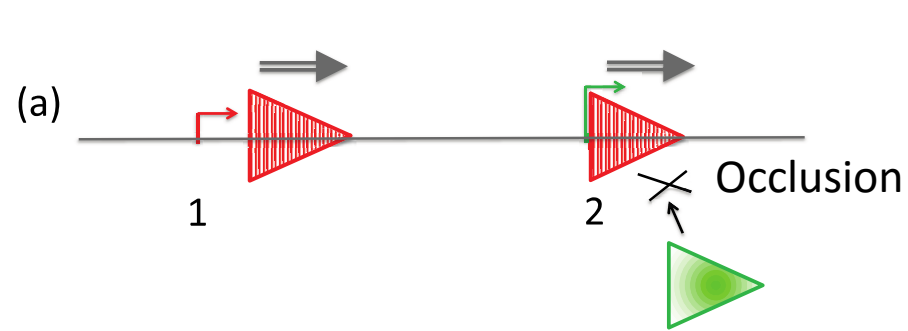


TRENDS in Genetics

Shearwin et al. Trends in Genet. (2005)

Transcriptional Interference

Ghosh, Bameta, Ghanti, Chowdhury, J. Stat. Mech.: Theor. & Expt. (2016)



$$\frac{dP_1(1, t)}{dt} = \underbrace{\alpha_1 \left(1 - \sum_{s=1}^{\ell} P(s) \right)}_{\text{Entry at ON-ramp}} - \underbrace{QP_1(1, t) \xi(\underline{1}|1 + \ell)}_{\text{Forward Hopping from } j=1},$$

$$\frac{dP_1(i, t)}{dt} = \underbrace{QP_1(i-1, t) \xi(\underline{i-1}|i-1 + \ell)}_{\text{Forward Hopping to } j=i} - \underbrace{QP_1(i, t) \xi(\underline{i}|i + \ell)}_{\text{Forward Hopping from } j=i} \quad \text{for, } (1 < i < L_1),$$

$$\frac{dP_1(L_1, t)}{dt} = \underbrace{QP_1(L_1-1, t) \xi(\underline{L_1-1}|L_1-1 + \ell)}_{\text{Forward Hopping to } j=L_1} - \underbrace{\beta P_1(L_1, t)}_{\text{Exit at OFF-ramp}}.$$

$$\frac{dP_2(1+h, t)}{dt} = \alpha_2 \xi(1+h) \left(1 - \sum_{s=1}^{\ell} P(s+h) \right) - QP_2(1+h, t) \xi(\underline{1+h}|1+h + \ell),$$

$$\frac{dP_2(i, t)}{dt} = QP_2(i-1, t) \xi(\underline{i-1}|i-1 + \ell) - QP_2(i, t) \xi(\underline{i}|i + \ell)$$

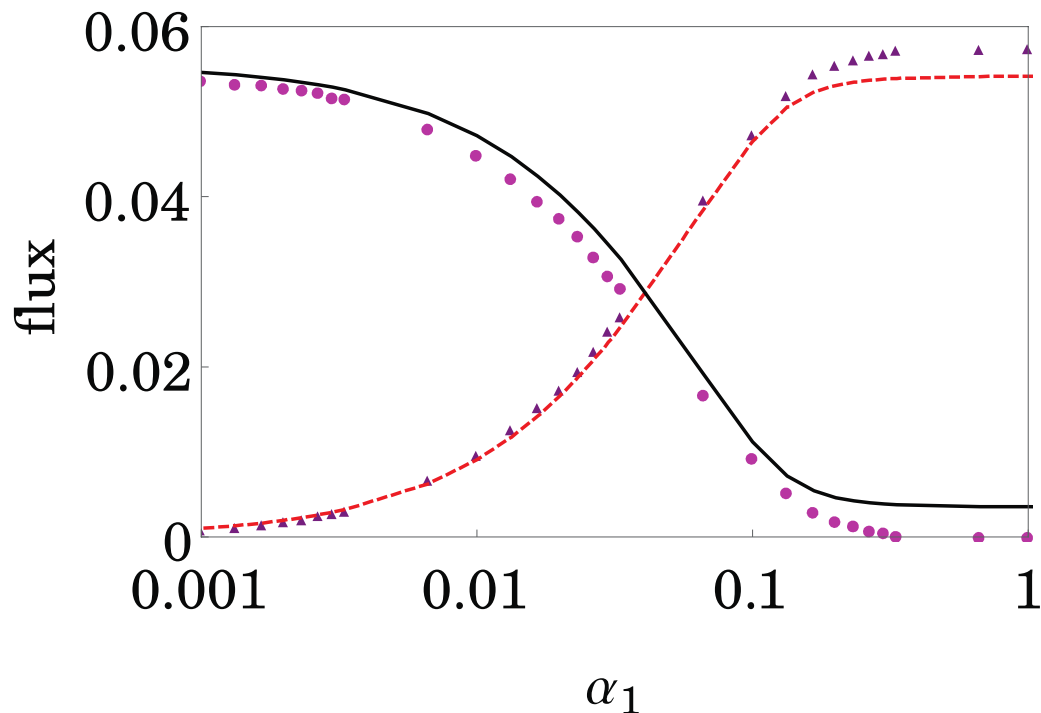
for , $(1+h < i < L_2+h)$,

$$\frac{dP_2(L_2+h, t)}{dt} = QP_2(L_2+h-1, t) \xi(\underline{L_2+h-1}|L_2+h-1 + \ell) - \beta P_2(L_2+h, t).$$

$$\xi(\underline{i}|i + \ell) = \frac{1 - \sum_{s=1}^{\ell} P(i+s)}{1 + P(i+\ell) - \sum_{s=1}^{\ell} P(i+s)}.$$

Co-directional TI

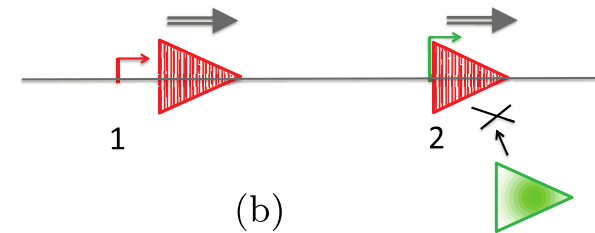
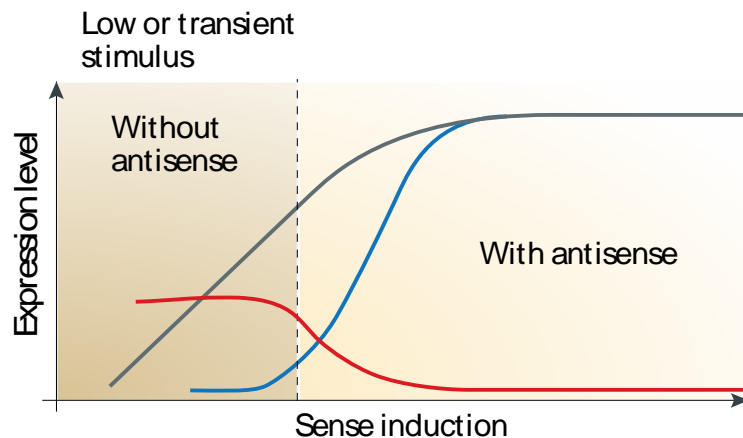
(a)



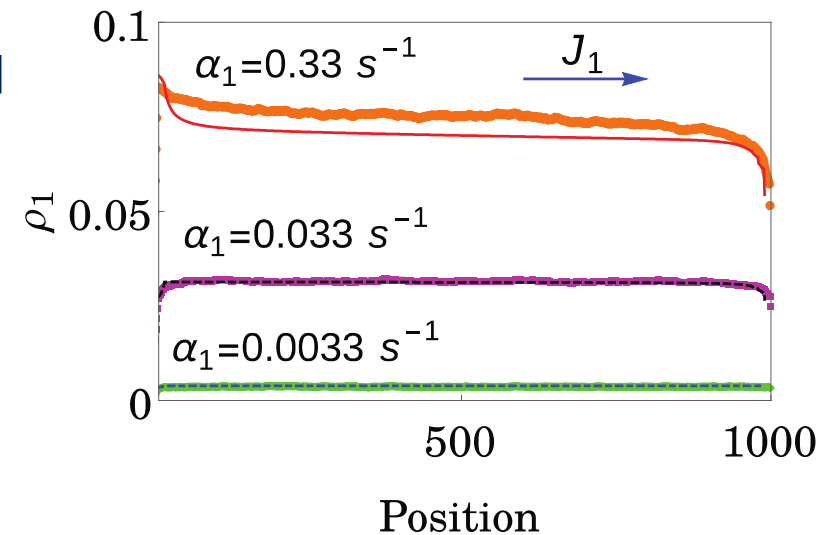
The pair can form a **bistable switch**: it can be in **either 'ON' state** (first gene is expressed and second gene is repressed) **or in an 'OFF' state** (first gene is repressed and the second gene is expressed)

■

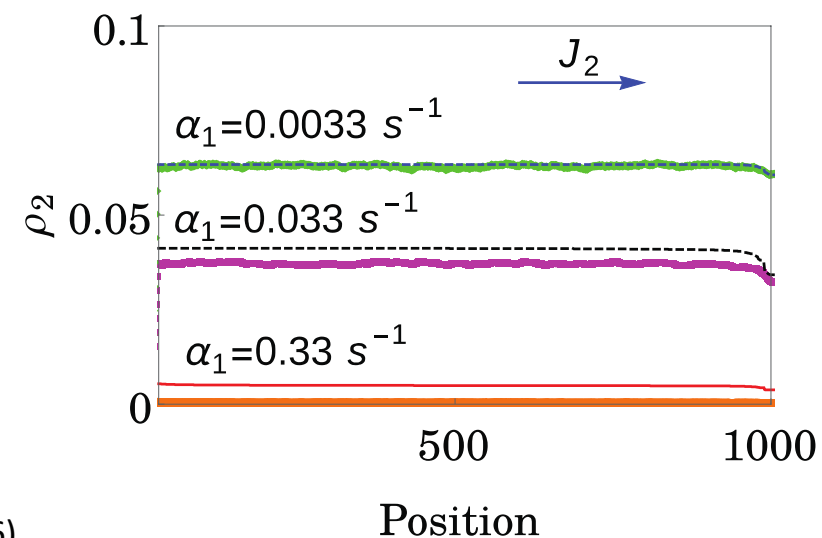
Pelechano and Steinmetz,
Nat. Rev. Genet. 14, 880 (2013).



(b)



(c)



Ghosh et al. J. Stat Mech. Theor. Expt (2016)

Contra-directional TI with Passing without detachment. (Phage RNAPs)

In a Head-on Collision, Two RNA Polymerases Approaching One Another on the Same DNA May Pass by One Another

Na Ma^{1,2,3} and William T. McAllister^{1,4} □

J. Mol. Biol. (2009) 391, 808–812

“To allow two polymerases to move toward one another in a controlled manner, a template was constructed that contains a promoter for T7 RNAP and a promoter for T3 RNAP arranged in opposite directions.”

(T)

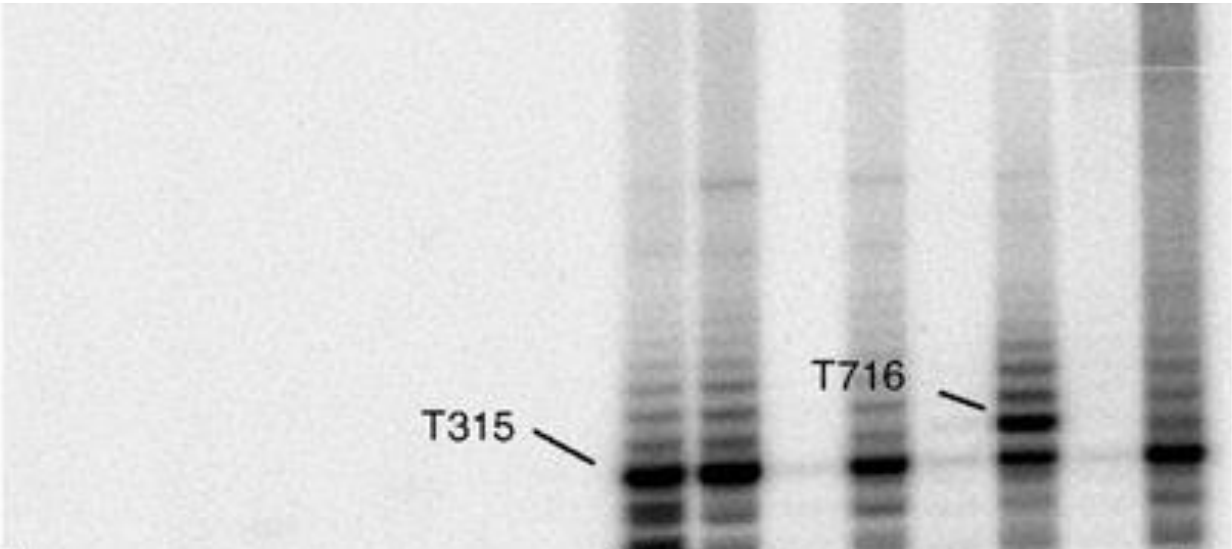
Step	NTP	T3 EC	H-T7 EC	Dis
1	G,A*	-	15*	NA
Immobilize			15*	NA
2	G,A*	13*	15*	16

T7 could advance past halted T3 EC and after collision halted T3 EC could resume transcription.



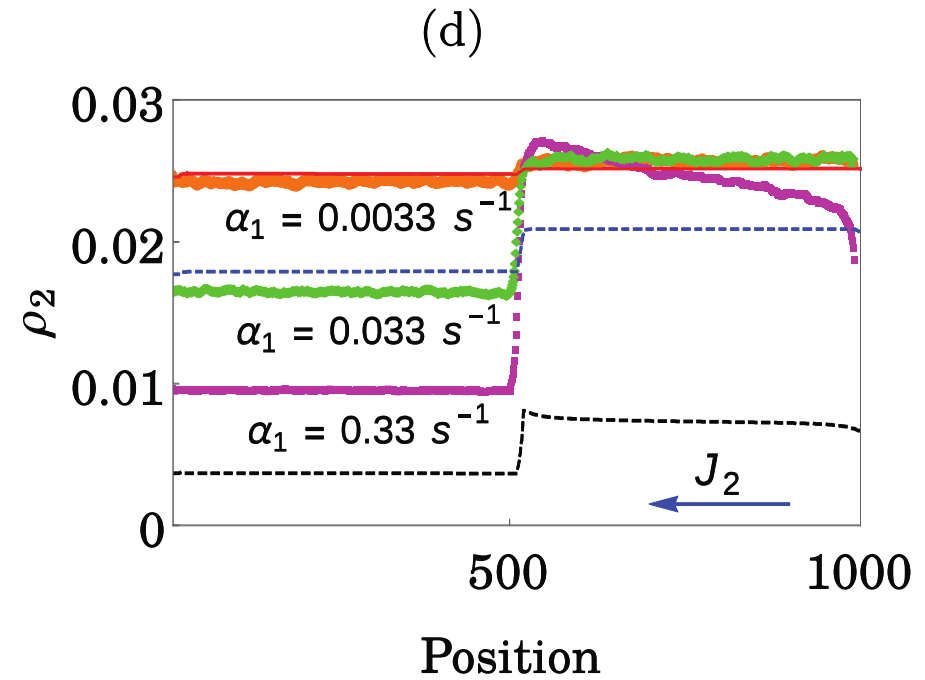
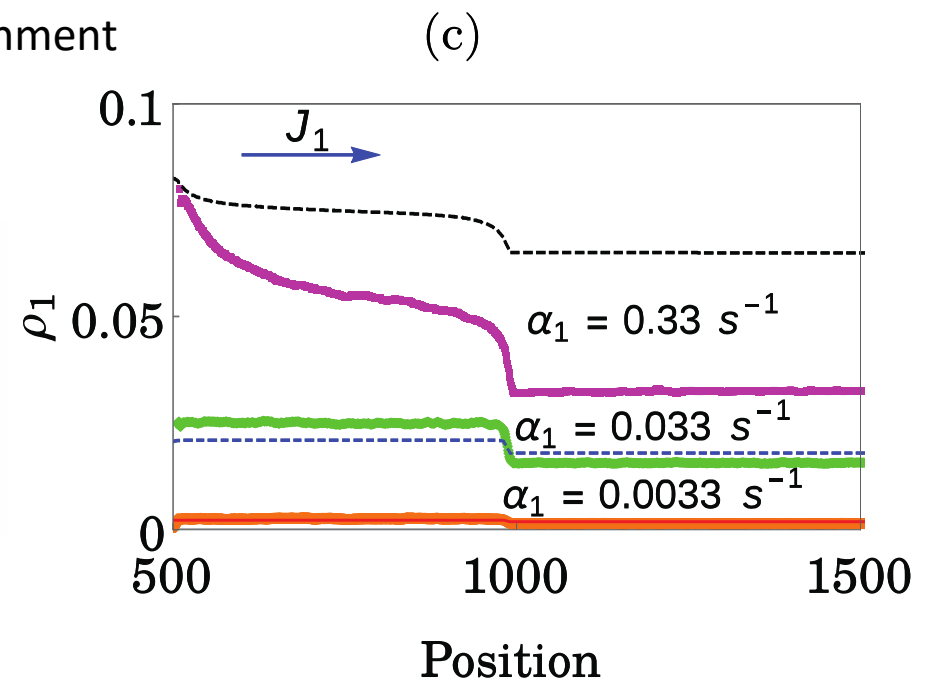
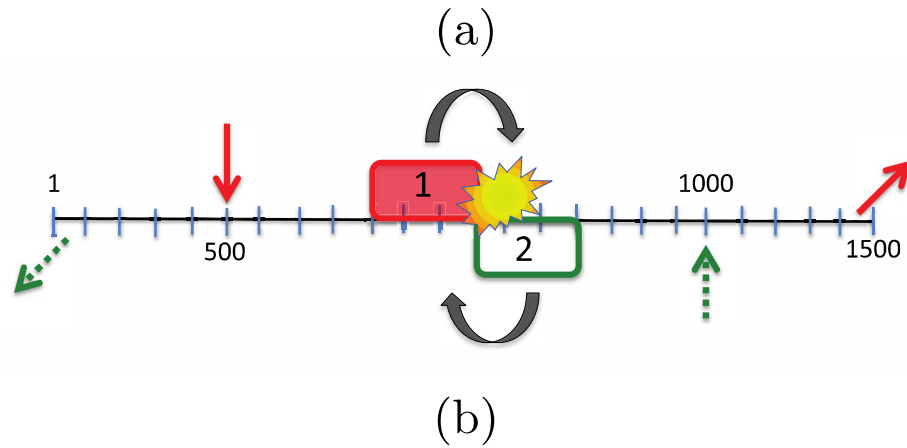
(9)

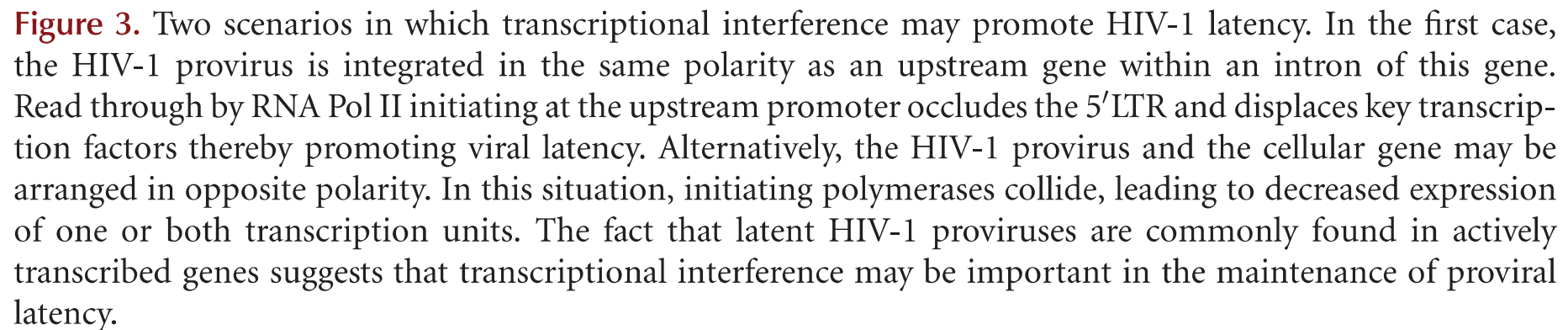
Step	1	2	3	4	5	6	7



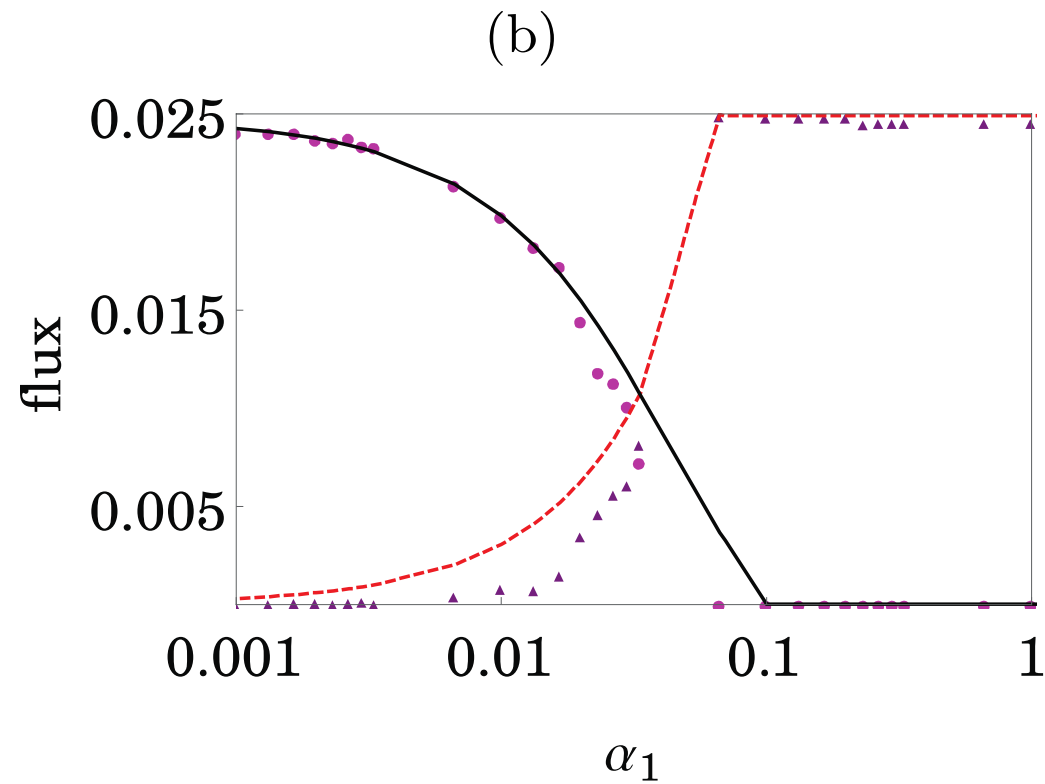
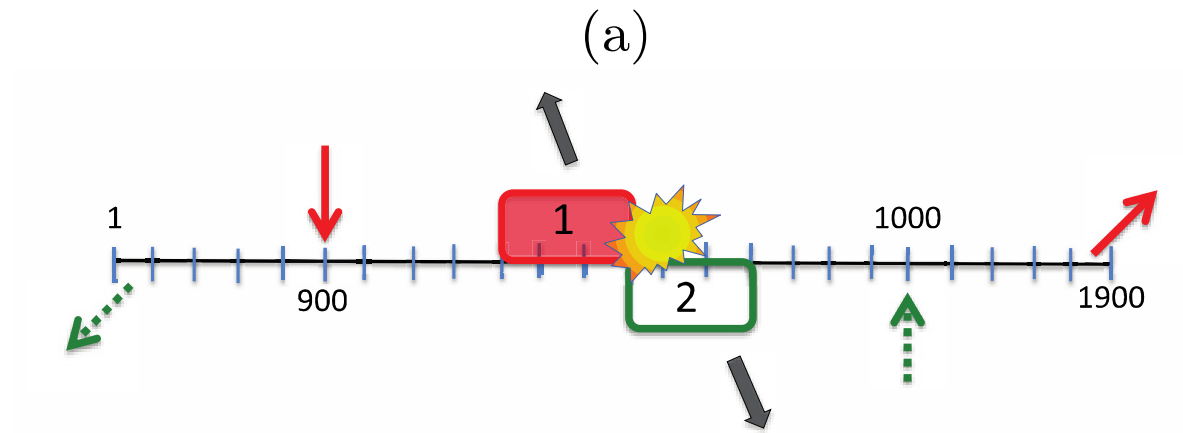
Since RNAPs moving in opposite directions use two different strands of the DNA as their templates, it seems likely that they manage to pass by one other by temporarily releasing their non-template strand while maintaining association with their template strand.

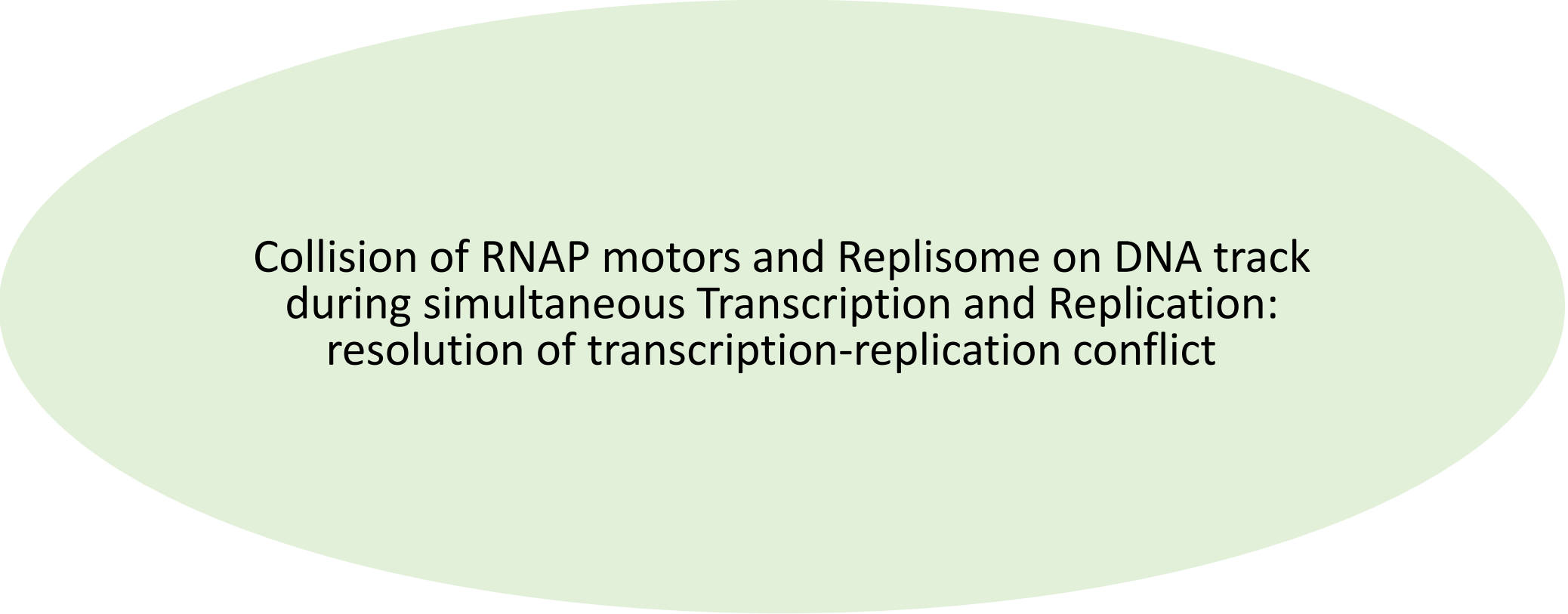
Contra-directional TI with Passing without detachment





Contra-directional TI with Detachment without Passing

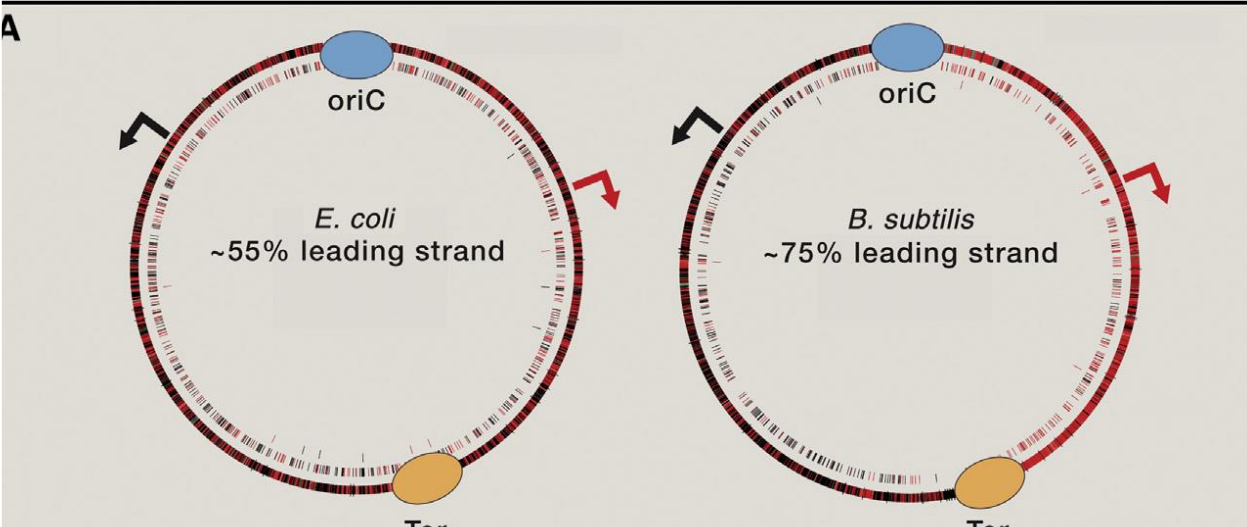




Collision of RNAP motors and Replisome on DNA track
during simultaneous Transcription and Replication:
resolution of transcription-replication conflict

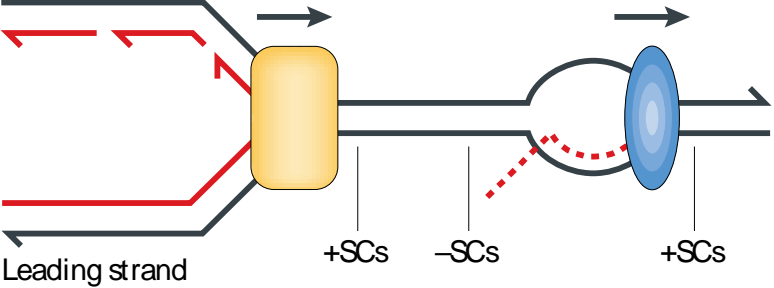
Genome wide Transcription and Replication:
Conflict Resolution

Hamperl and Cimprich, Cell 167, 1455 (2016)



a Co-directional replisome and RNAP

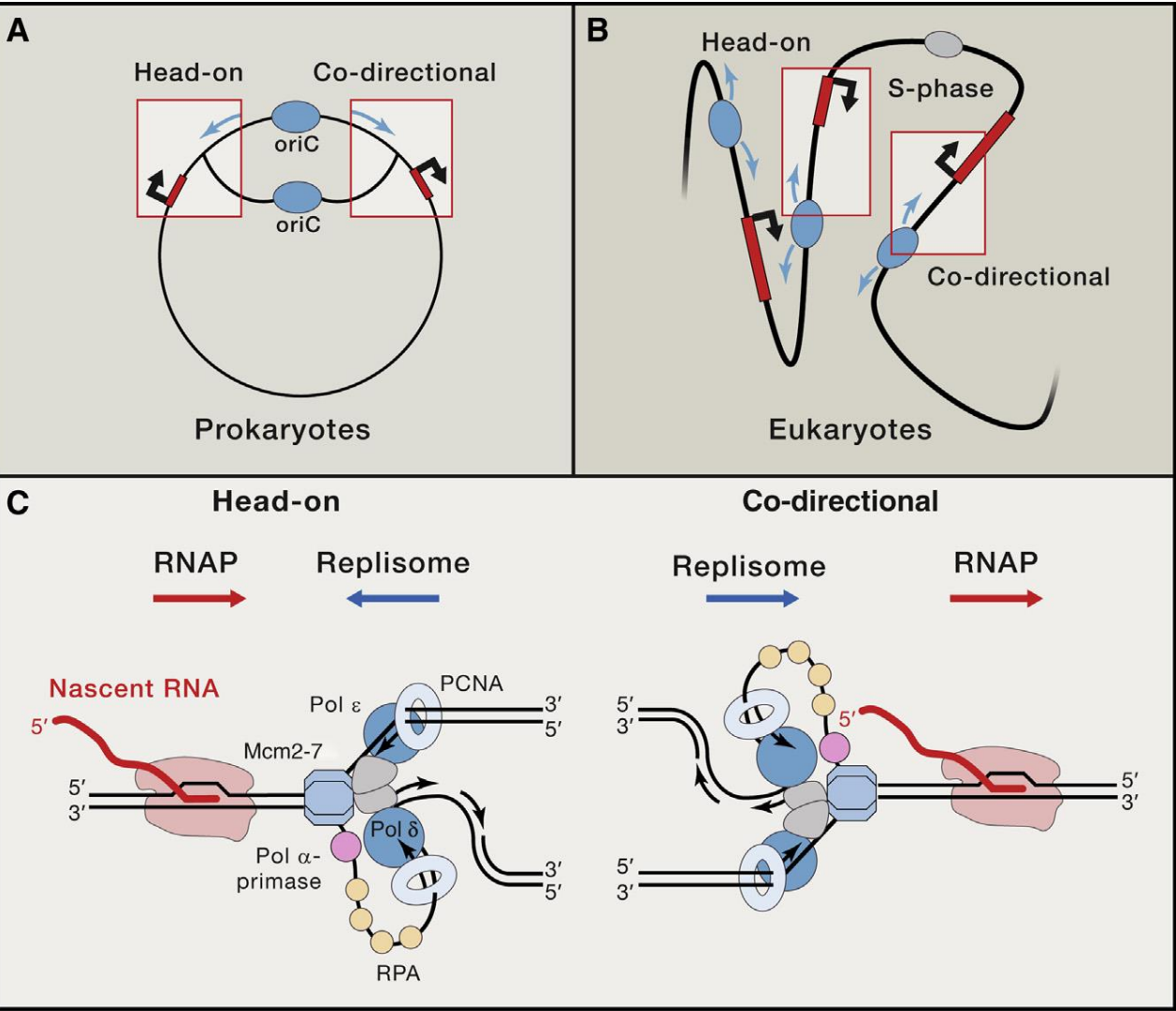
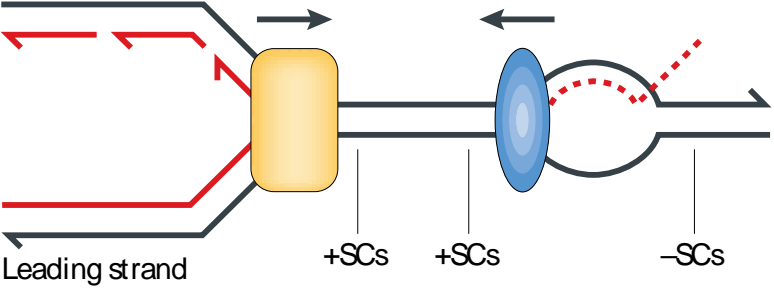
Lagging strand



Kim and Robertson, Nat. Rev. Genet. 13, 204 (2012)

b Head-on replisome and RNAP

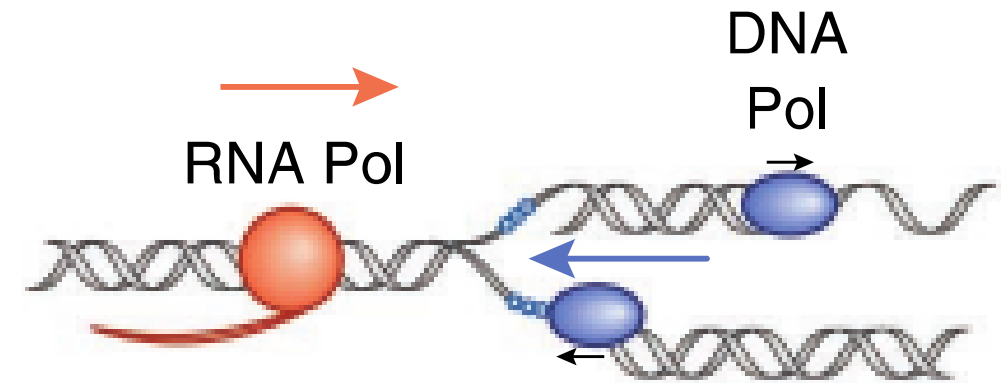
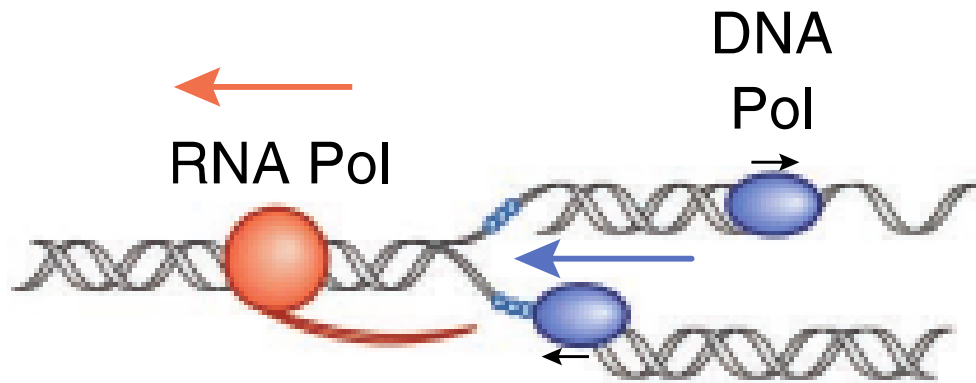
Lagging strand



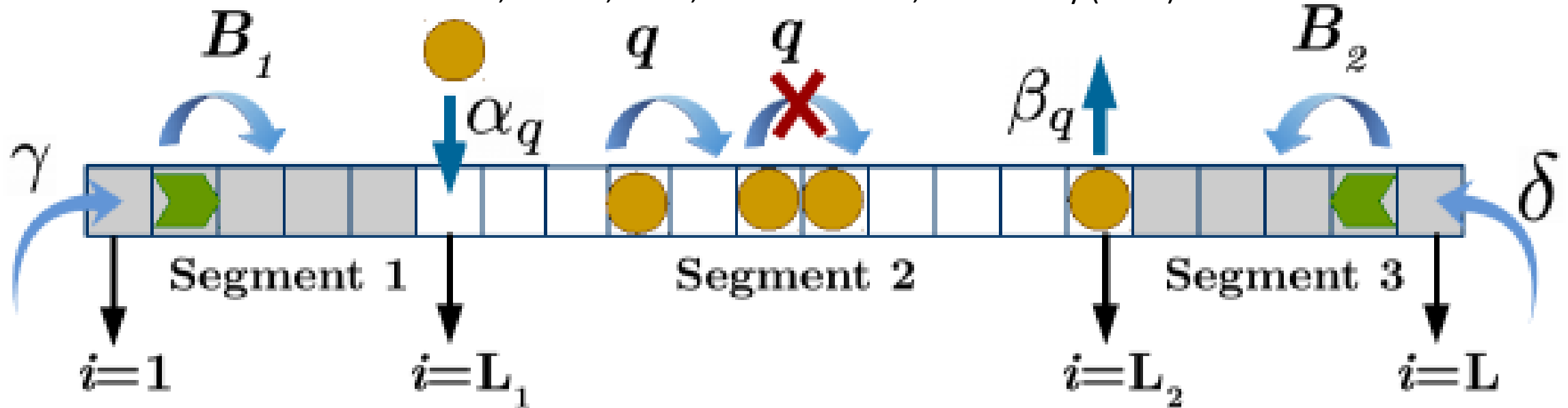
Codirectional encounters

Helmrich, Ballarino, Nudler, Tora,
Nat. Struct. Mol. Biol.20, 412 (2013)

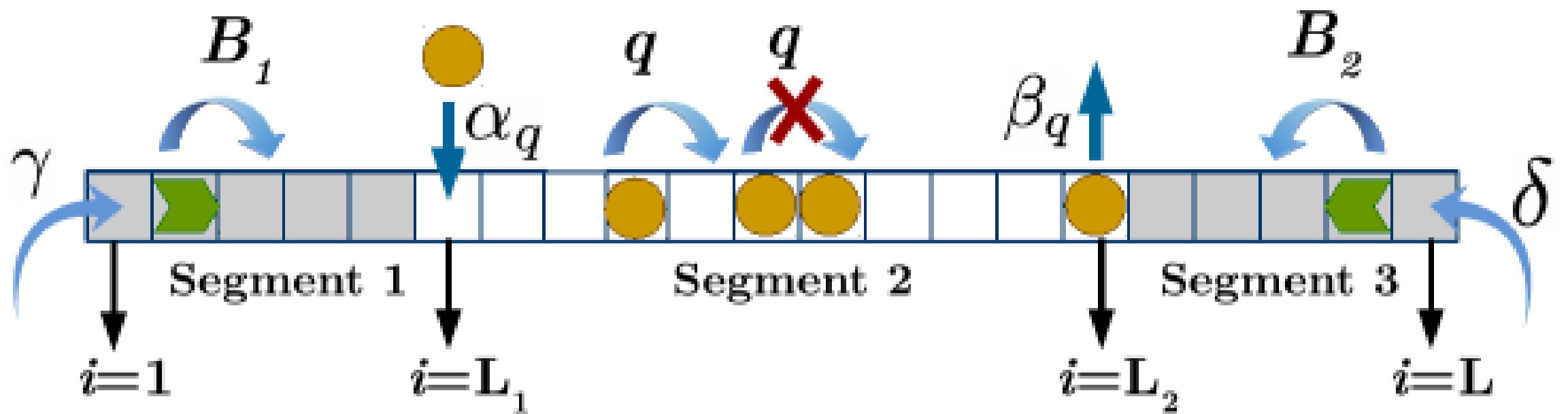
Head-on encounters



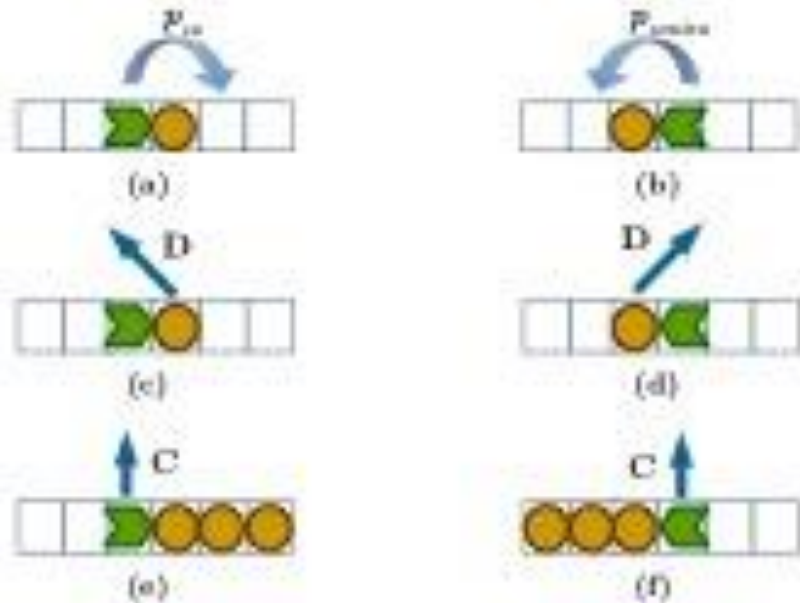
Ghosh, Mishra, Patra, Schadschneider, Chowdhury (2017)



Ghosh, Mishra, Patra, Schadschneider, Chowdhury (2017)



Ghosh, Mishra, Patra, Schadschneider, Chowdhury (2017)

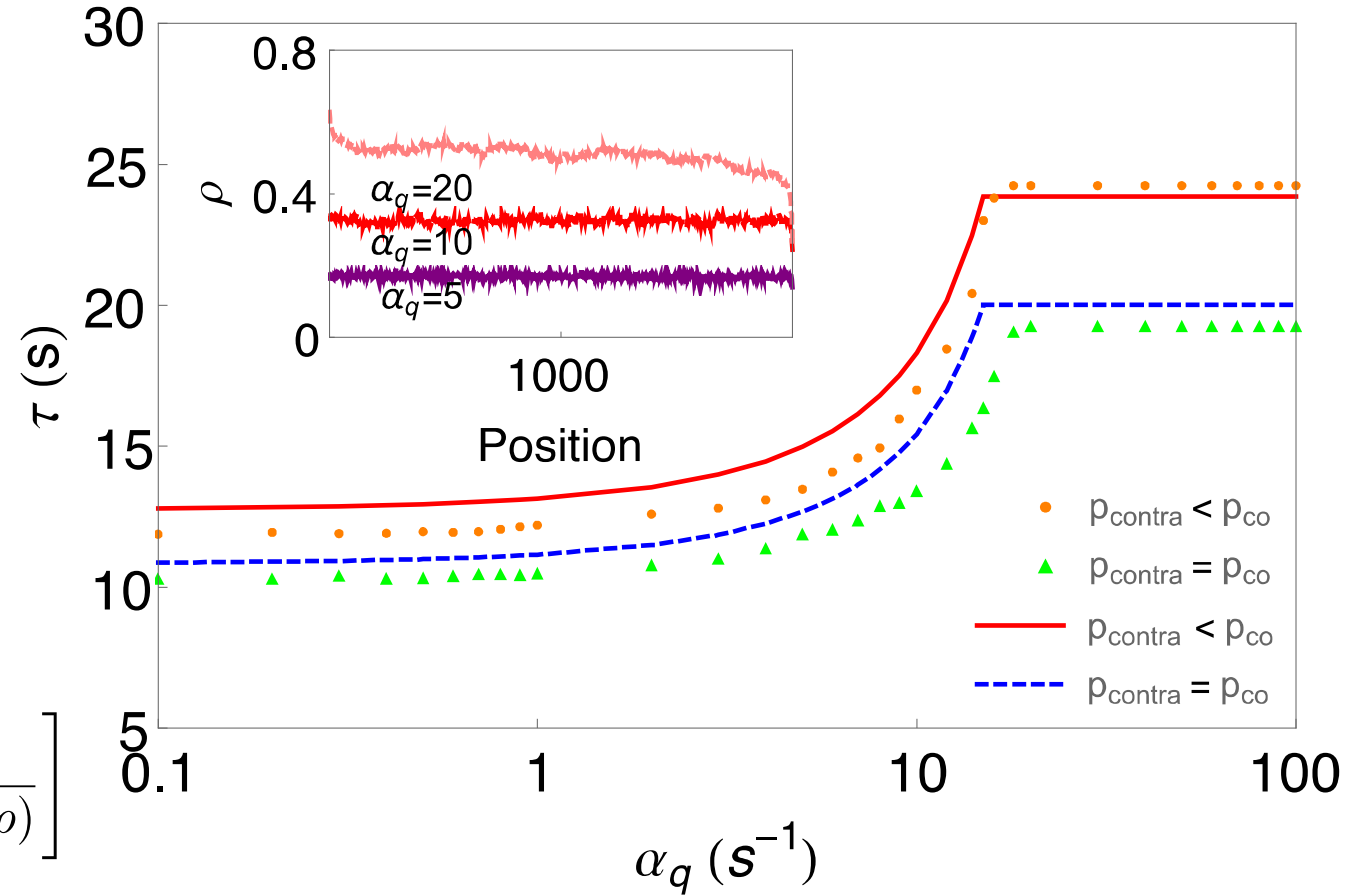


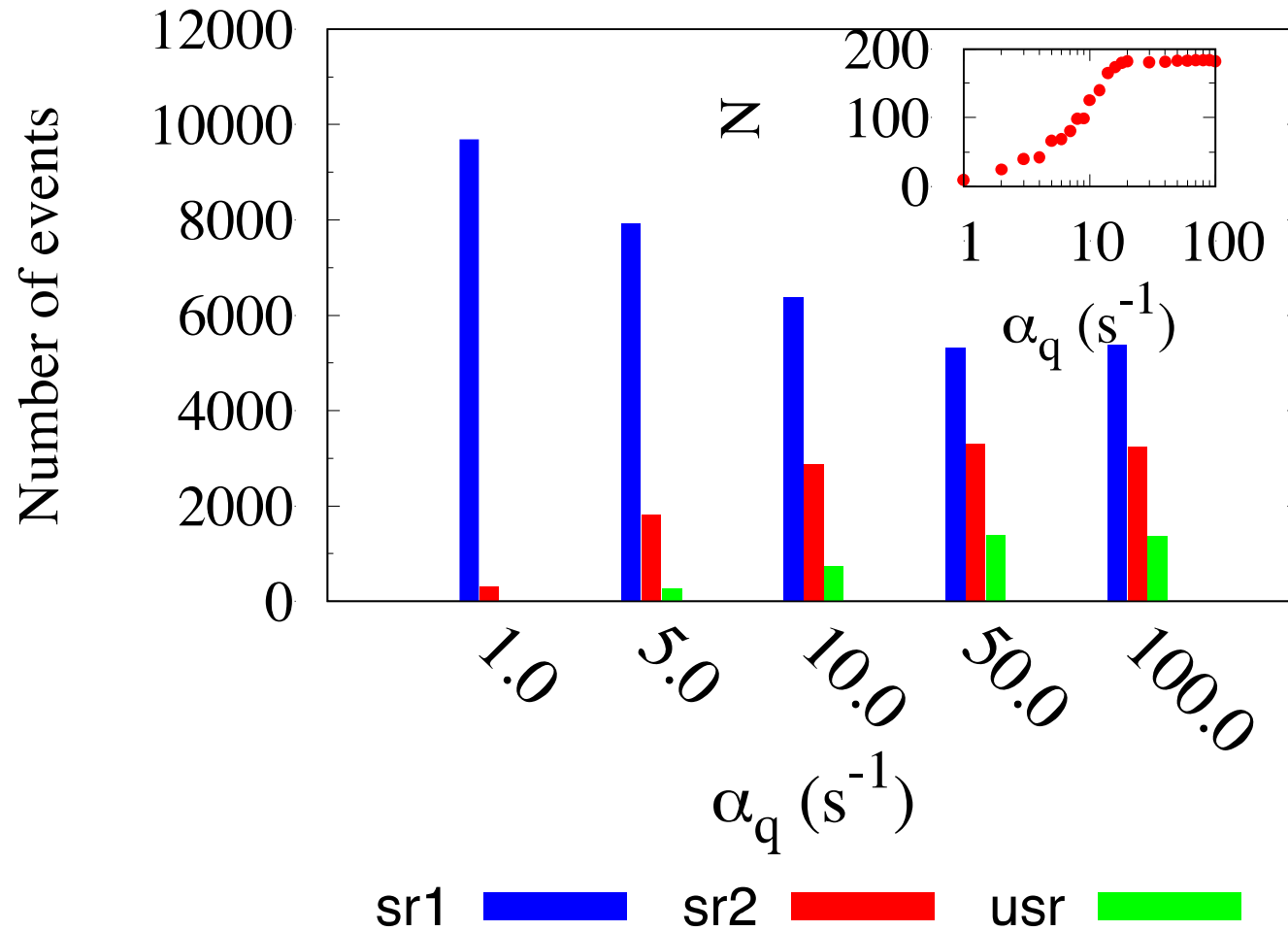
Effect of Transcription on Replication

$$\begin{aligned}\tau &= \frac{\tau_{int}}{2} + \frac{L}{4B_1} \\ &= \frac{\rho L}{8} (\tau_{co} + \tau_{contra}) + \frac{L}{4B_1},\end{aligned}$$

$$\begin{aligned}\tau_{co} &= \frac{d}{v_{R_\ell} - v_P} \\ &= \frac{1}{2\rho} \left[\frac{1}{B_1(1-\rho) - q(1-\rho)} + \frac{1}{p_{co}(1-\rho) - q(1-\rho)} \right]\end{aligned}$$

$$\begin{aligned}\tau_{contra} &= \frac{d}{v_{R_r} + v_P} \\ &= \frac{1}{2\rho} \left[\frac{1}{B_2(1-\rho) + q(1-\rho)} + \frac{1}{p_{co}(1-\rho) + q(1-\rho)} \right]\end{aligned}$$





Effect of Transcription on Replication

Ribosome Drop-off Model

Bonnin, Kern, Young, Stansfield, Romano, PLoS Comp. Biol. (2017)

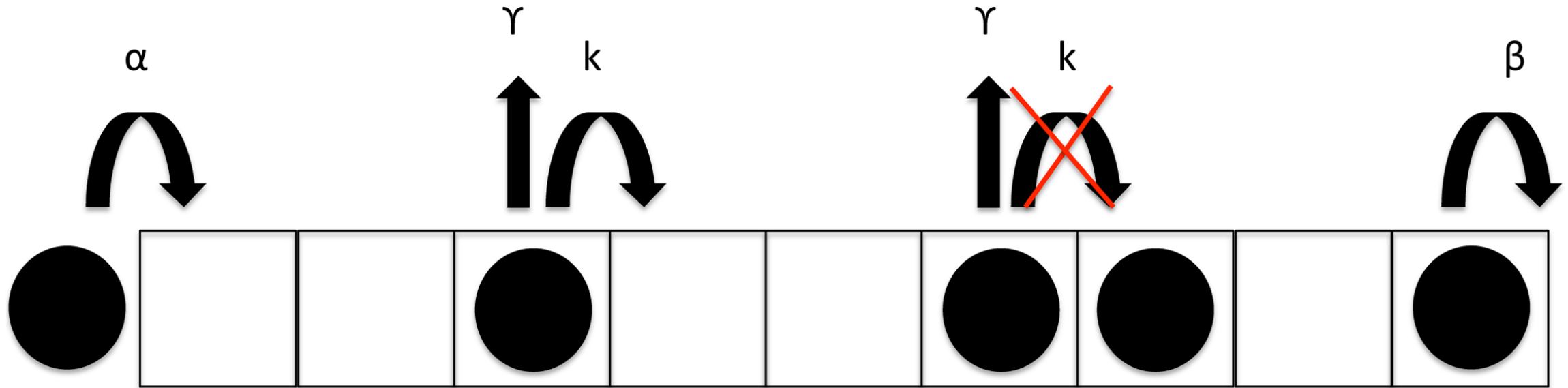
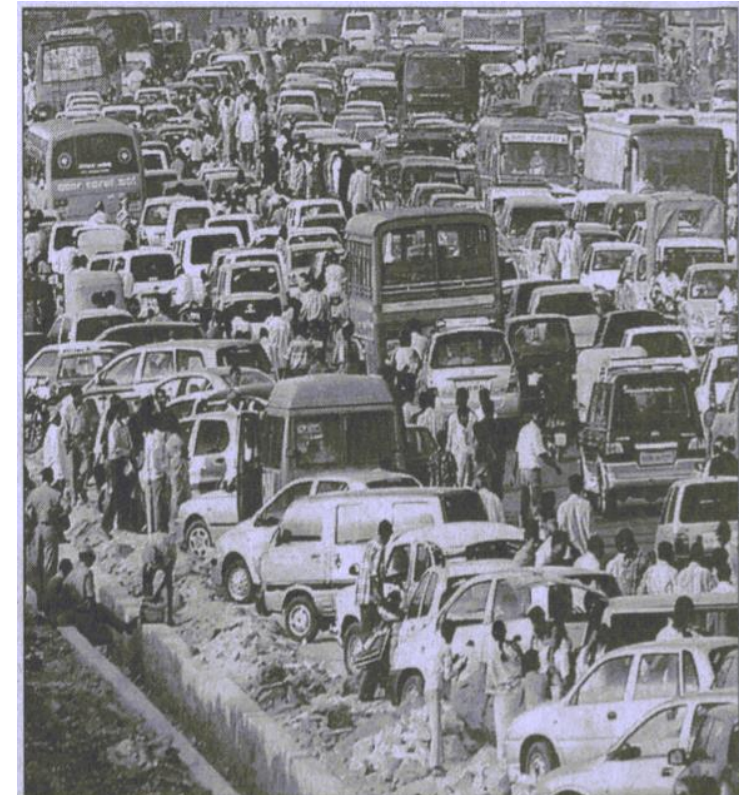
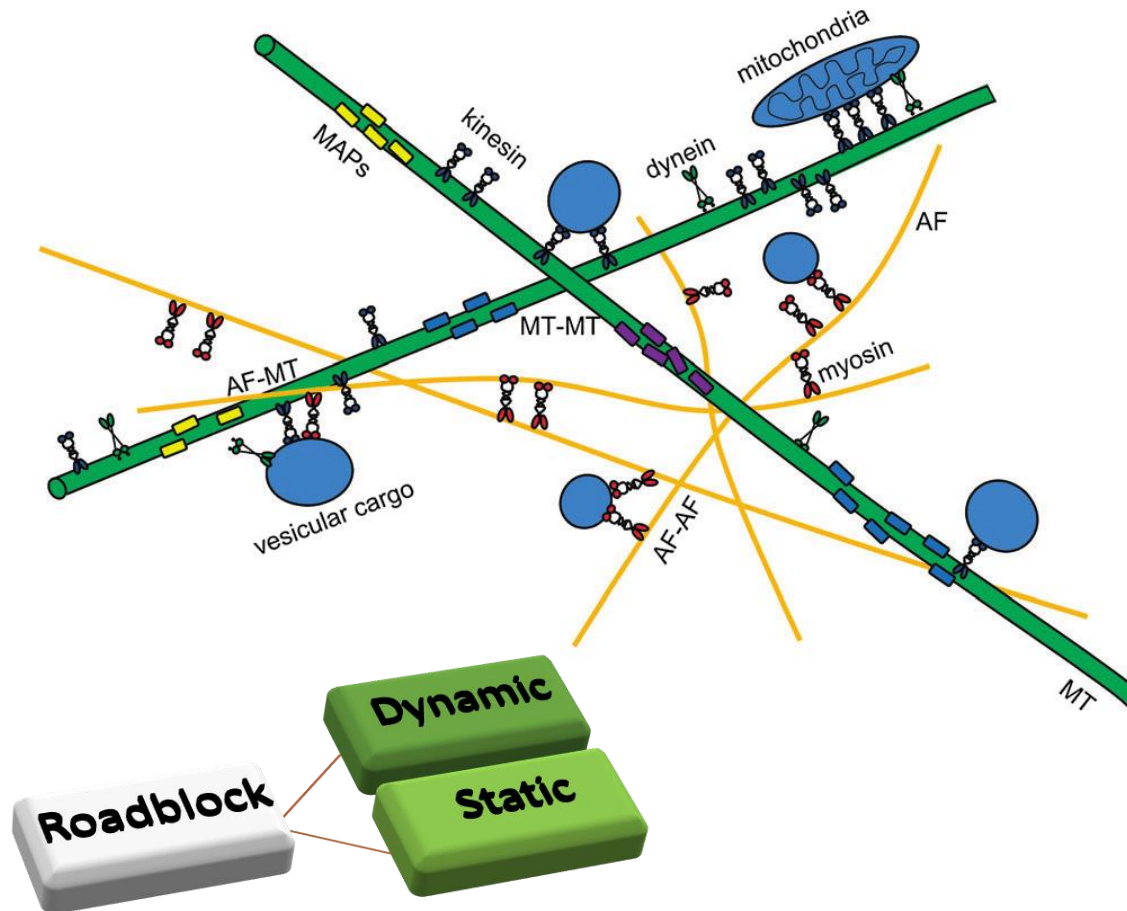


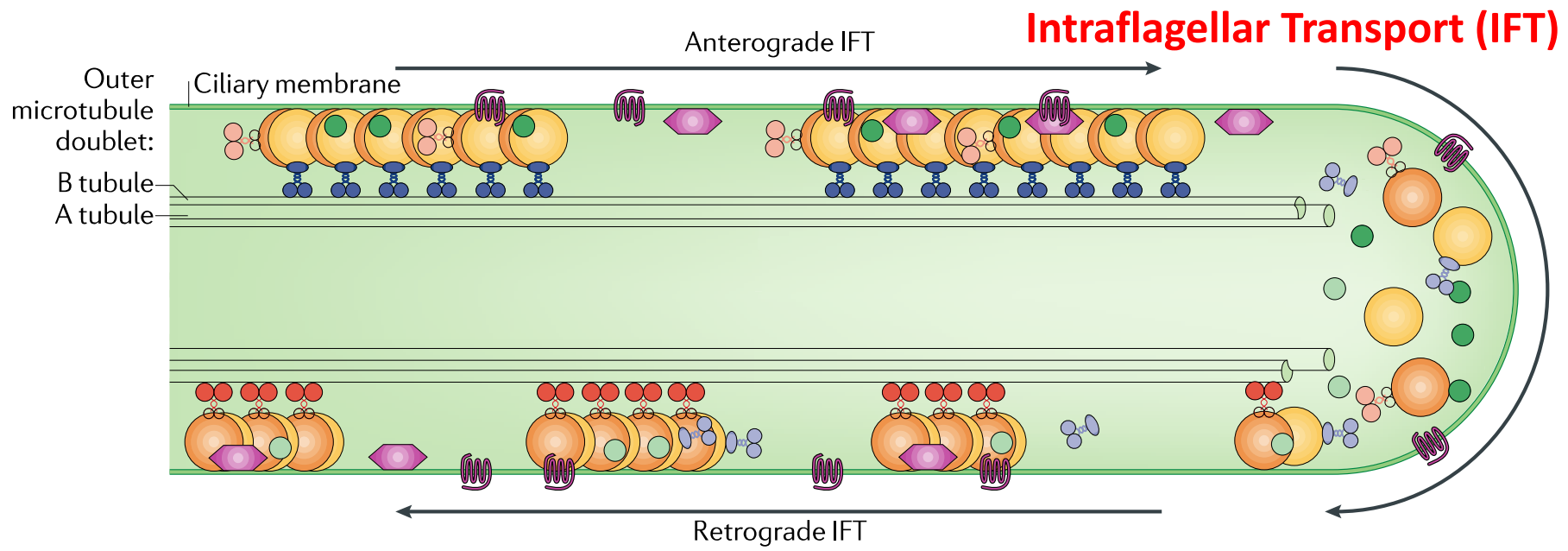
Fig 2. Illustration of the ribosome drop-off model. Ribosomes bind to the first codon of the mRNA with rate α and leave the lattice representing the mRNA strand at the stop codon, with rate β . Throughout the mRNA lattice they can either hop to the next codon, if it is free, with rate k , or drop off the lattice with rate γ .

“Like cities, cells have developed diverse transport systems to ensure that the right components are delivered to, or manufactured at, the right location at the right time. What are these transport systems? What are the intracellular roads or tracks, what are the engines and motors, and how do they operate? How are the various types of cargo shipped, and how is such transport tailored to demand? What determines whether it is the finished product that is shipped, or rather smaller parts or subunits for local on-site assembly? How are such mechanisms regulated to maintain cellular function, react to physiological stimuli, and ensure flexible adaptation to changing environments?”

Tiedge et al., PNAS (2001)

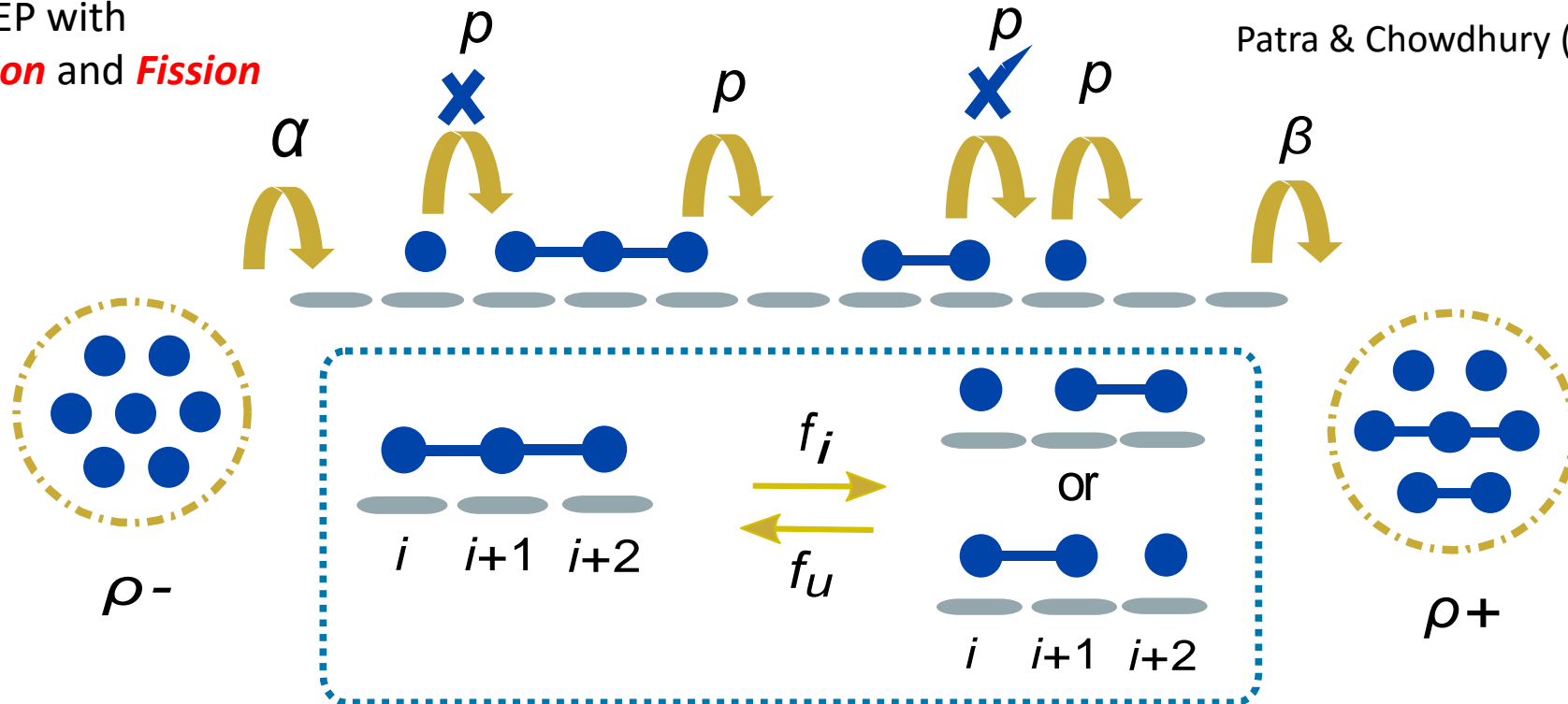


Times of India, Feb 16 (2007)



TASEP with
Fusion and **Fission**

Patra & Chowdhury (2017)



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