

Genetic Oscillations and Feed-Backs in NF- κ B, p53 and Wnt Systems

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1. Four eukaryotic systems with oscillatory gene expressions:

- *Hes1-mRNA* protein network (Hirata et al (2002))
- *P53-mdm2* network (Oren et al (2000), Lahav et al (2006))
- NF- κ B transcription factor (Nelson, White et al)
- Wnt-Notch segmentation network (Goldbeter, Pourquie)

→ Identify the ‘simplest’ negative feed-back loop.

→ Oscillating regimes: Ultradian time period (2-3 hours)

2. **NF- κ B, p53, Wnt systems: Regulated by negative feed-back loops: inflammation , apoptosis, segmentation.**
3. **NF- κ B - I κ B feed-back loop:**
 - Reduce 26-dimensional dynamics to three/nine variables**
 - Spiky oscillations:**
 - Saturated degradation.**
 - A20 change period. Chaotic response**
4. **External stresses and responses in p53-Mdm2:**
 - DNA damage, hypoxia, nutlin, etc**
5. **Somite segmentation in embryos (space):**
 - Oscillating proteins: Wnt signaling feed-back loop**
6. **Coupled feed-back loops in space:**
 - Cell-to-cell communications.**

Collaborators:

- Sandeep Krishna, Guido Tiana (Milan), Simone Pigolotti
- Kim Sneppen
- Peter B. Jensen, Lykke Pedersen, Alex Hunziker, Benedicte Mengel,

S. Krishna, M.H. Jensen and K. Sneppen, "Spiky oscillations in NF- κ B signalling", Proc.Nat.Acad.Sci. 103, 10840-10845 (2006).

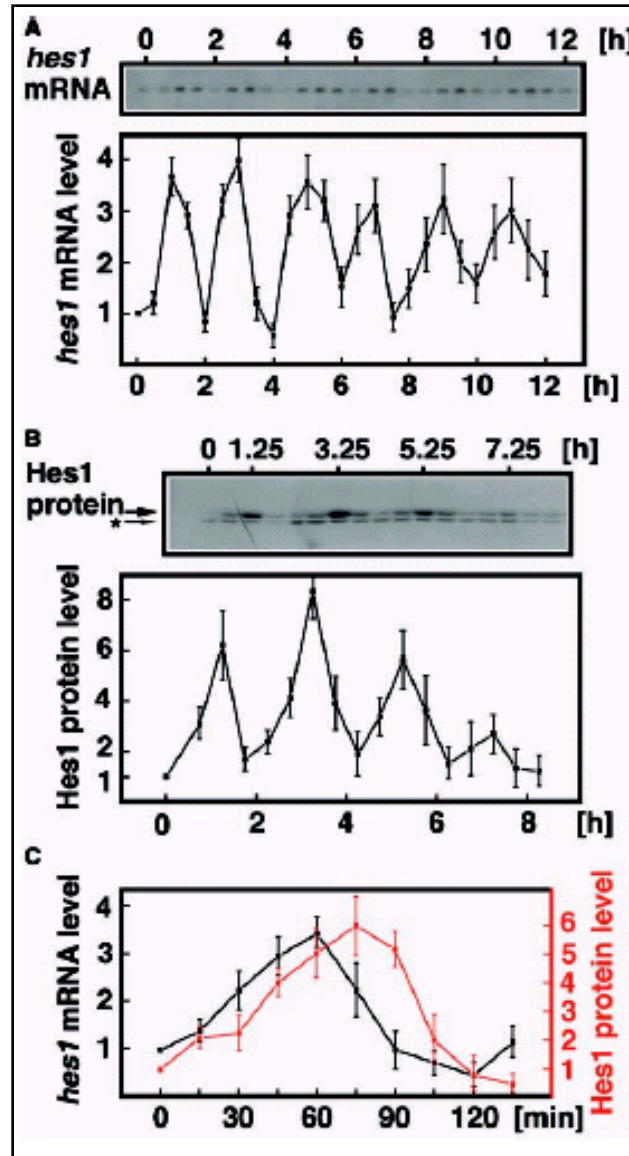
S. Pigolotti, S. Krishna, M.H. Jensen, "Oscillation patterns in negative feedback loops", Proc.Nat.Acad.Sci., 104, 6533-6537 (2007).

G. Tiana, S. Krishna, S. Pigolotti, M.H. Jensen and K. Sneppen, "Oscillations and temporal signalling in cells", Physical Biology 4, R1-R17 (2007).

S. Pigolotti, S. Krishna, M.H. Jensen "Symbolic Dynamics of Biological Feedback Networks", Phys. Rev. Lett. 102, 088701 (2009); 103, 118101 (2009).

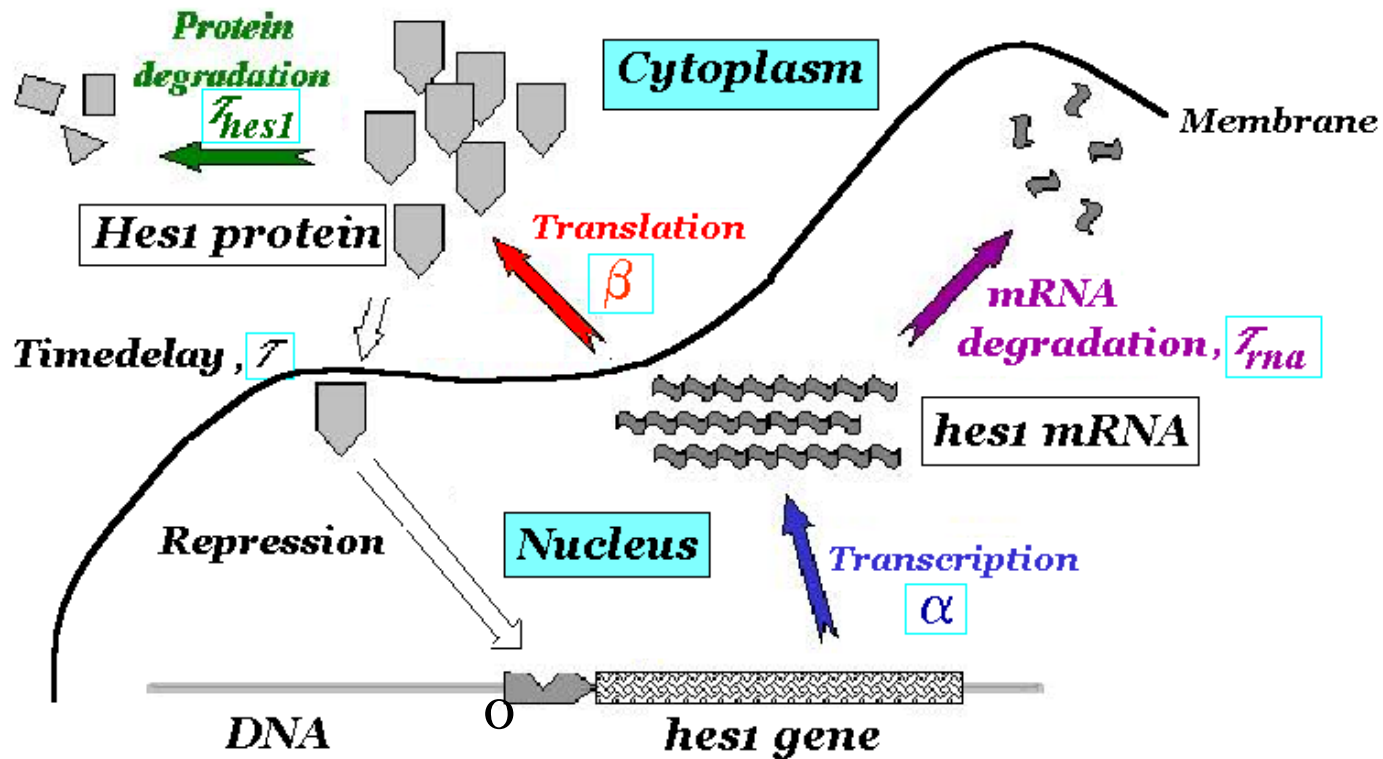
P.B. Jensen, L. Pedersen, S. Krishna, M.H. Jensen "A Wnt Oscillator Model for Somitogenesis", Biophys. Journ. 98, 943-50 (2010). Several new preprints

'Typical' Oscillating data: Hes1 - segmentation



(Hirata et al, 2002)

Simplest negative feed-back loop: Hes1



$$\frac{d[mRNA]}{dt} = \alpha \cdot [o_{free}] - \frac{[mRNA(t)]}{\tau_{rna}}$$

$$\frac{d[Hes1]}{dt} = \beta \cdot [mRNA(t)] - \frac{[Hes1(t)]}{\tau_{hes1}}$$

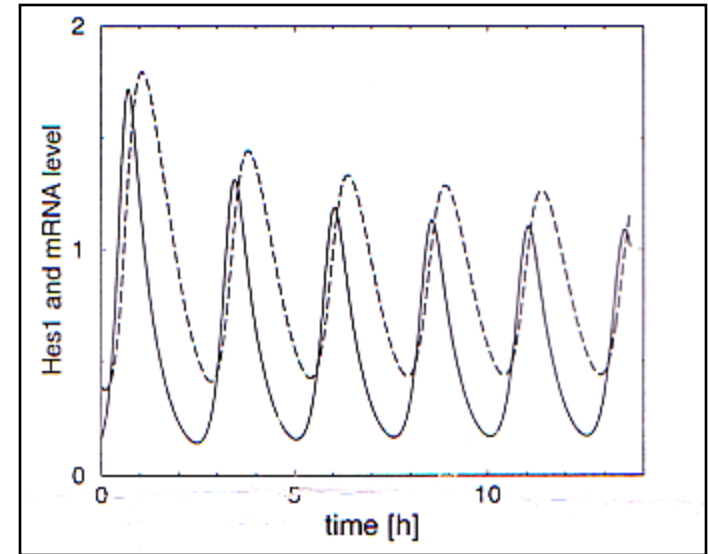
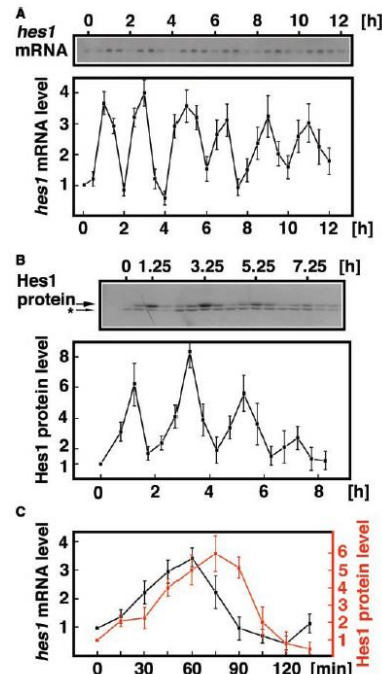
Jensen et al 2003

$$\frac{d[mRNA]}{dt} = \alpha \cdot \frac{K_M}{K_M + [Hes1(t - \tau)]^n} - \frac{[mRNA(t)]}{\tau_{rna}}$$

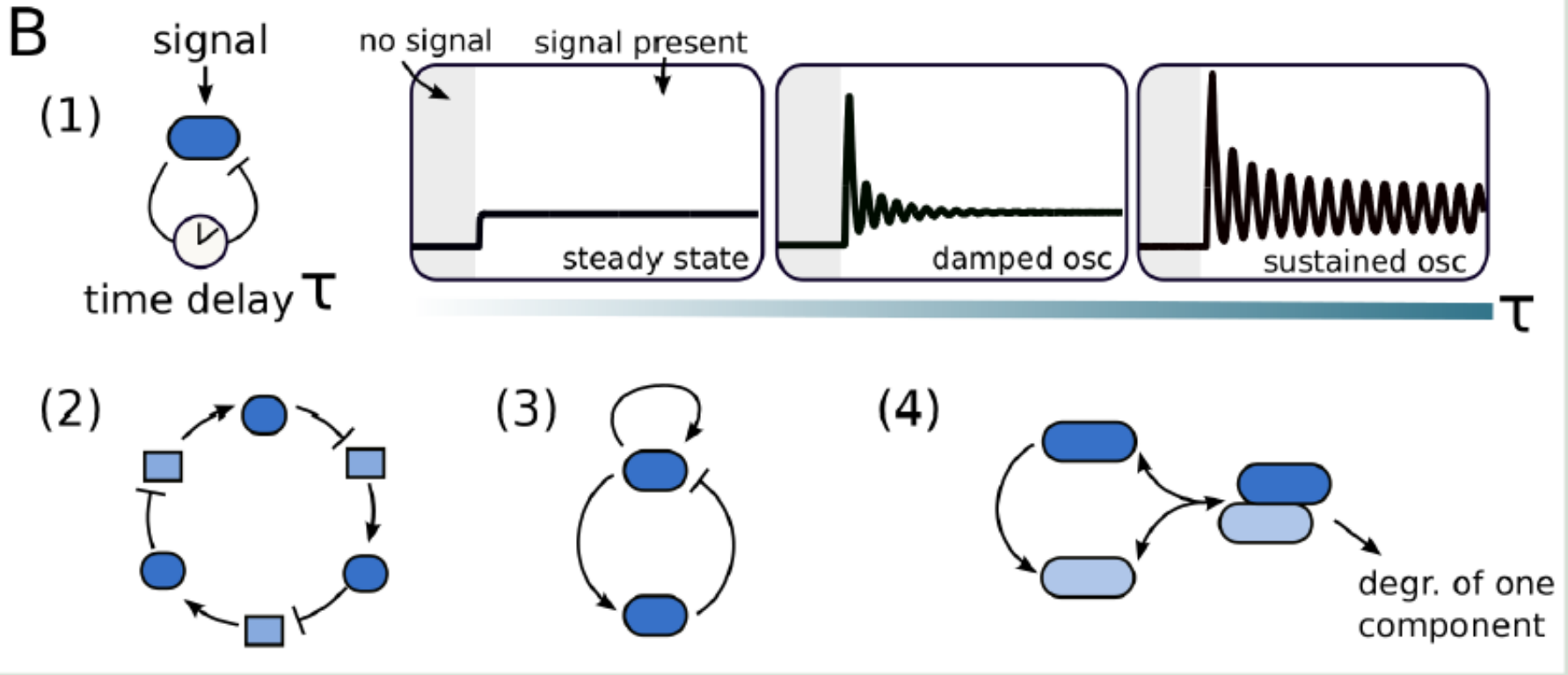
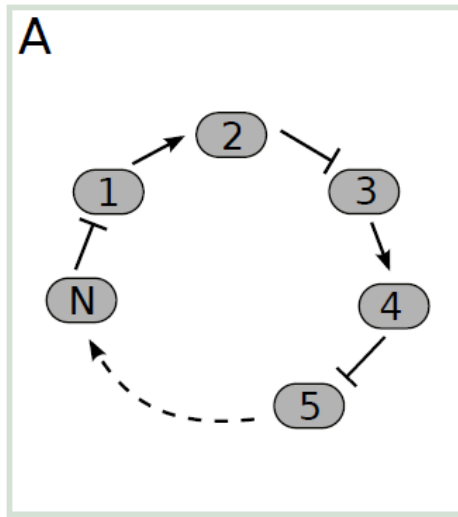
$$\frac{d[Hes1]}{dt} = \beta \cdot [mRNA(t)] - \frac{[Hes1(t)]}{\tau_{hes1}}$$

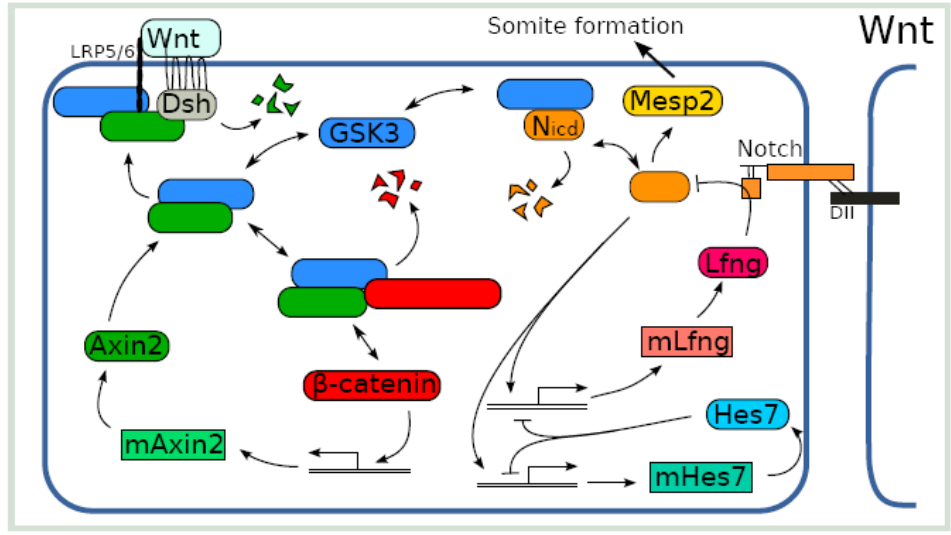
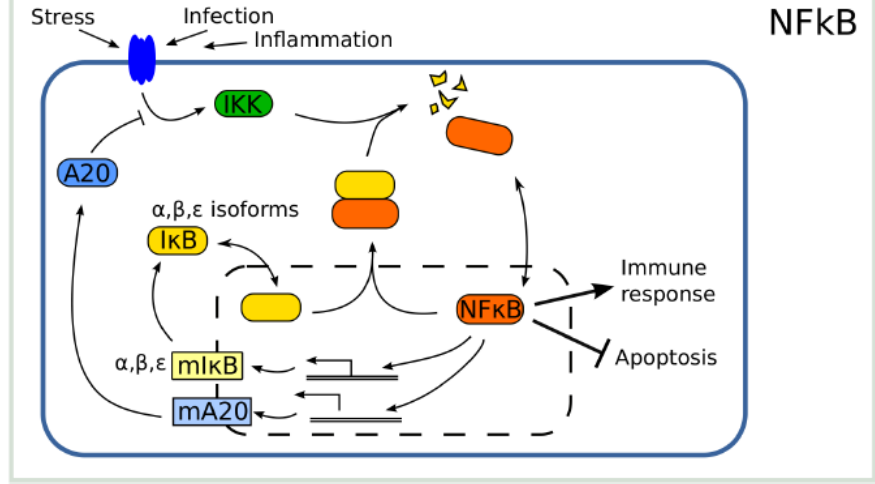
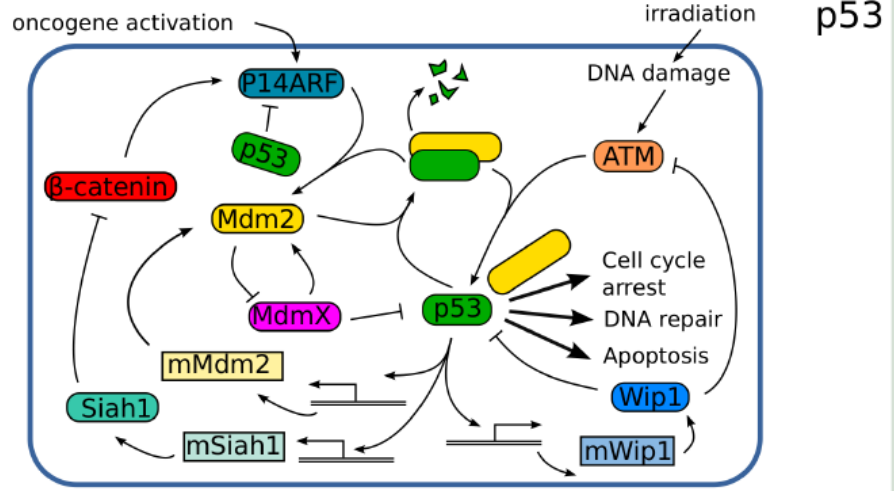
- Dashed curve [Hes1]
- Solid curve [mRNA]

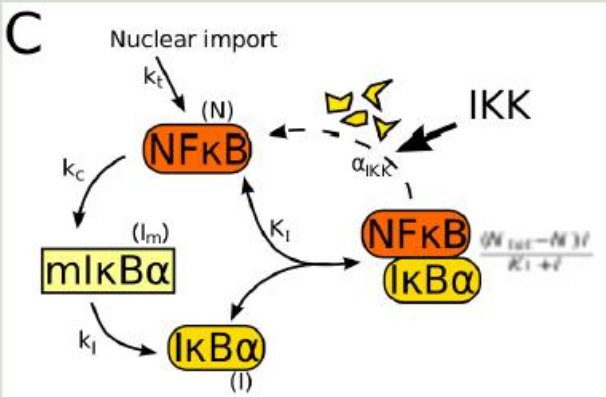
- $\tau_{rna} = 24.1$ min
- $\tau_{hes1} = 22.3$ min
- $\tau = 24$ min
- $\alpha = 20 [R]_0 \text{ min}^{-1}$
- $\beta = 1/20 \text{ min}^{-1}$
- $K_M = (0.1 [R]_0)^n$
- $n = 4$



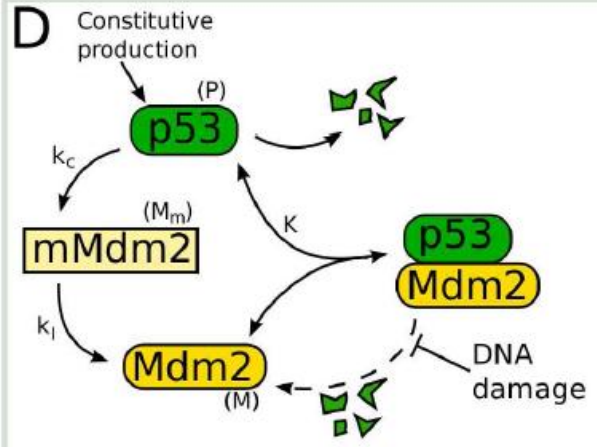
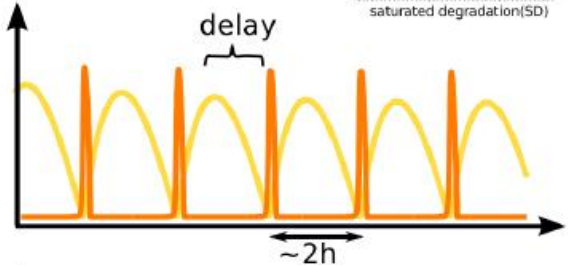
Negative Feed-Back loops:



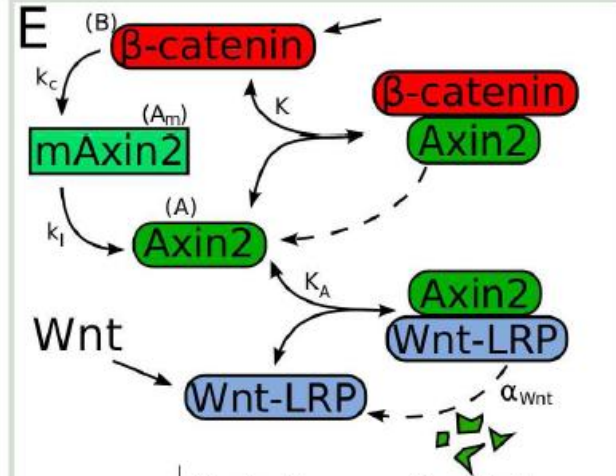
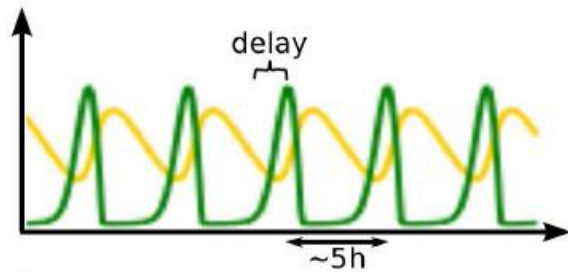




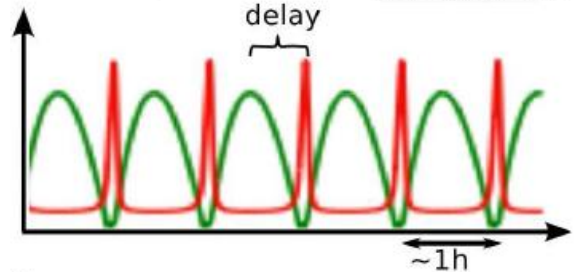
	Production	Degradation
$\frac{dN}{dt} =$	$k_t \frac{(N_{tot} - N) K_I}{K_I + I}$	$\delta \frac{I N}{K + N}$
$\frac{dI_m}{dt} =$	$k_c N^2$	βI_m
$\frac{dI}{dt} =$	$k_l I_m$	$\alpha_{IKK} \frac{(N_{tot} - N) I}{K_I + I}$ saturated degradation (SD)



	Production	Degradation
$\frac{dP}{dt} =$	k_s	$\delta M \frac{P}{K + P}$ SD
$\frac{dM_m}{dt} =$	$k_c P^2$	βM_m
$\frac{dM}{dt} =$	$k_l M_m$	αM



	Production	Degradation
$\frac{dB}{dt} =$	k_s	$\delta B \frac{A}{K + A}$
$\frac{dA_m}{dt} =$	$k_c B^2$	βA_m
$\frac{dA}{dt} =$	$k_l A_m$	$\alpha_{Wnt} \frac{A}{K_A + A}$ SD



Why oscillations ?

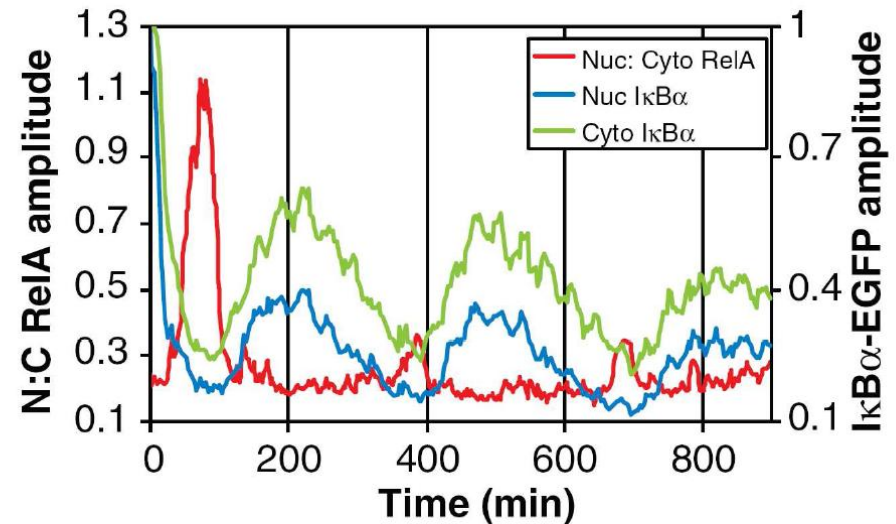
- Importance for DNA-repair and apoptosis
- Essential in segmentation
- Spiky oscillations →
important for **sharp responses**,
fast regulations,
high Hill coefficients
→ hormones also come in spikes

Mathematically: The most 'simple' dynamics !

The NF- κ B System in Mammalian Cells

- NF- κ B family: dimeric transcription factors
- Regulates immune response, inflammation, apoptosis
- Over 150 triggering signals, over 150 targets
- Each NF- κ B has a partner inhibitor I κ B
- Fluorescence imaging of NF- κ B and I κ B in human S-type neuroblastoma cells.

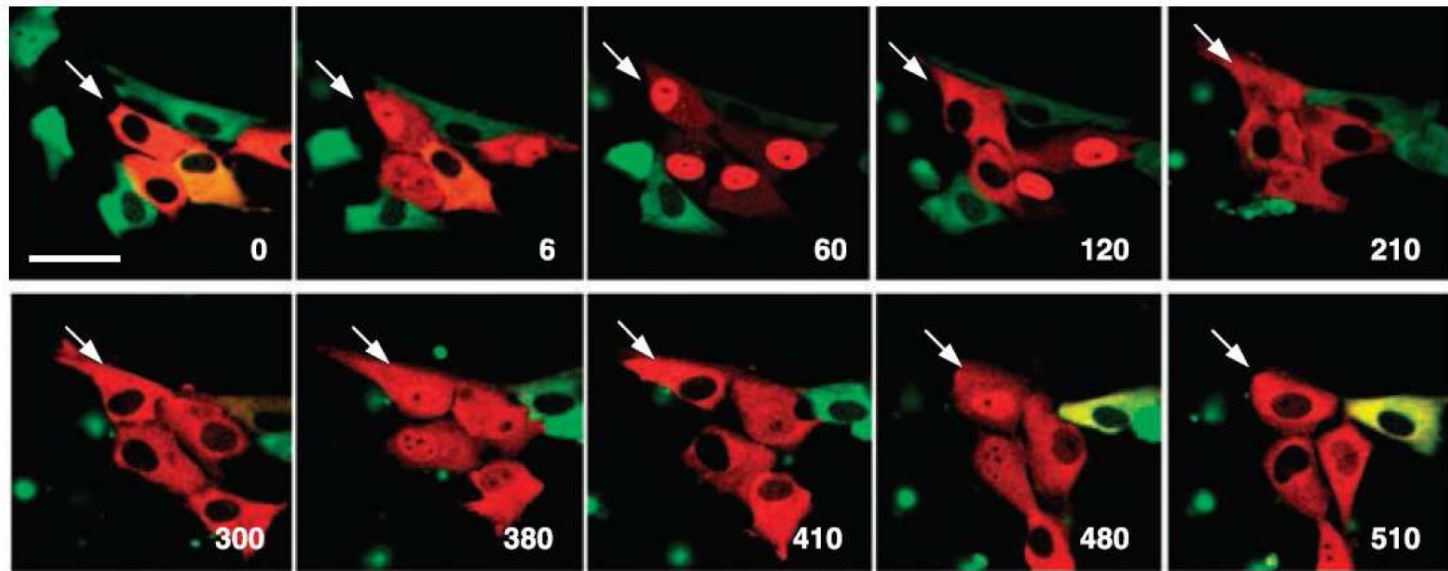
Nelson et al. (2004) *Science* 306, 704.



How does the network produce oscillations?

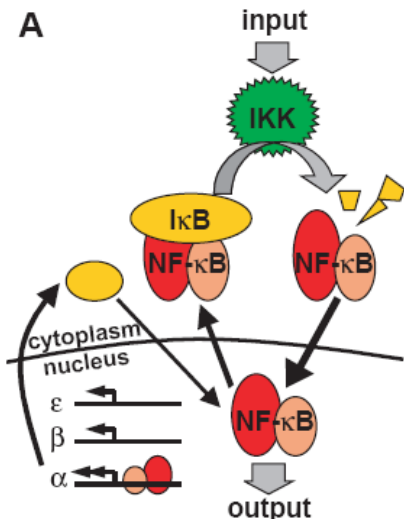
Why does the cell need the oscillations?

'Direct' observations of oscillations in nucleus

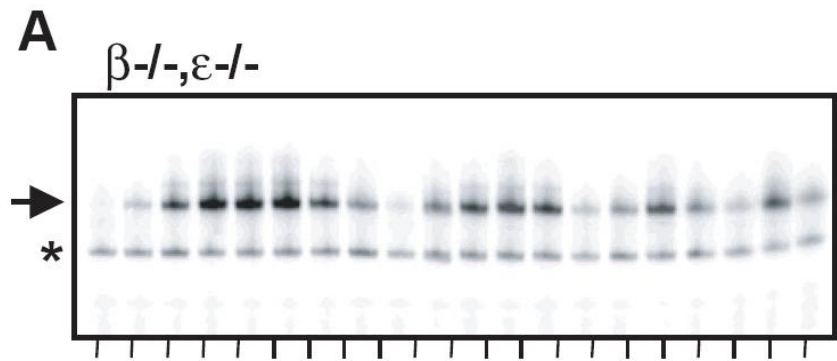


Oscillations in the nuclear localization of an NF- κ B transcription factor in human cells

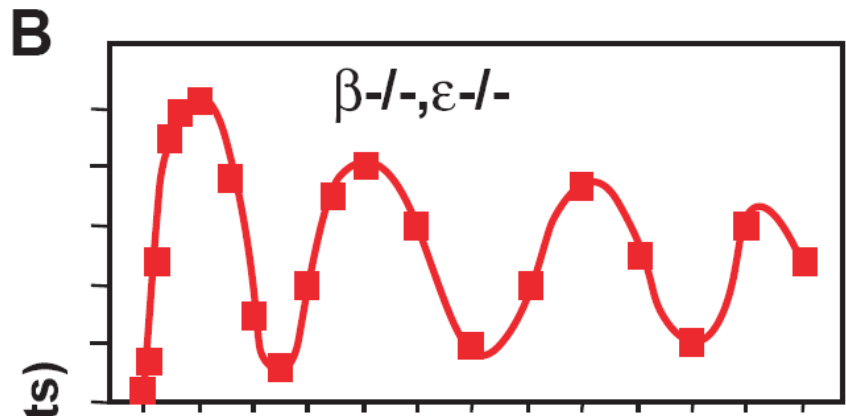
Nelson et al. (2004) *Science* 306, 704.



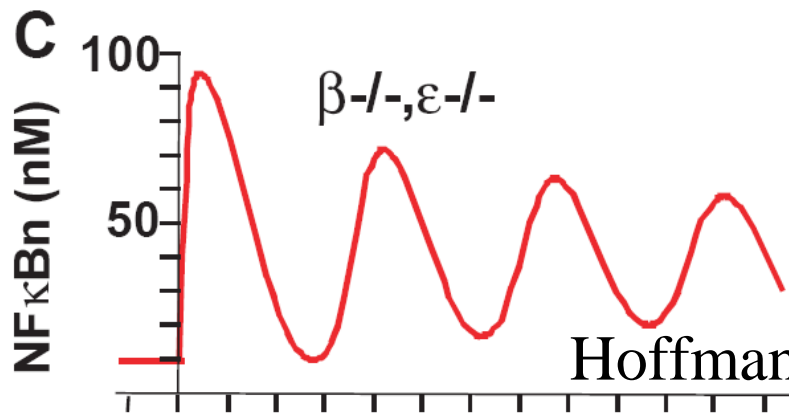
The NF-κB feed-back network



NF-κB oscillations

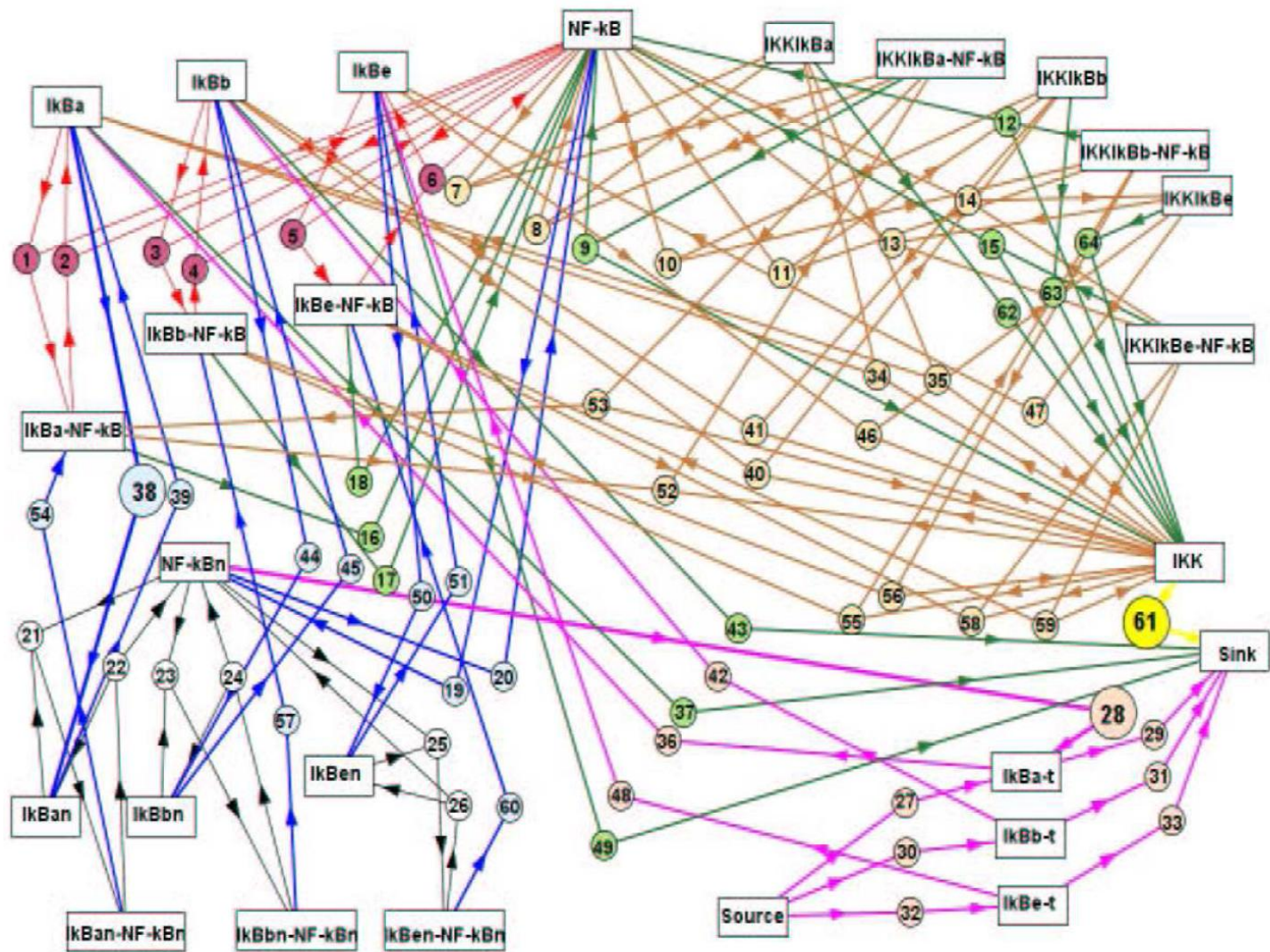


IκBβ/ε knocked out



Hoffmann, Baltimore et al (2002)

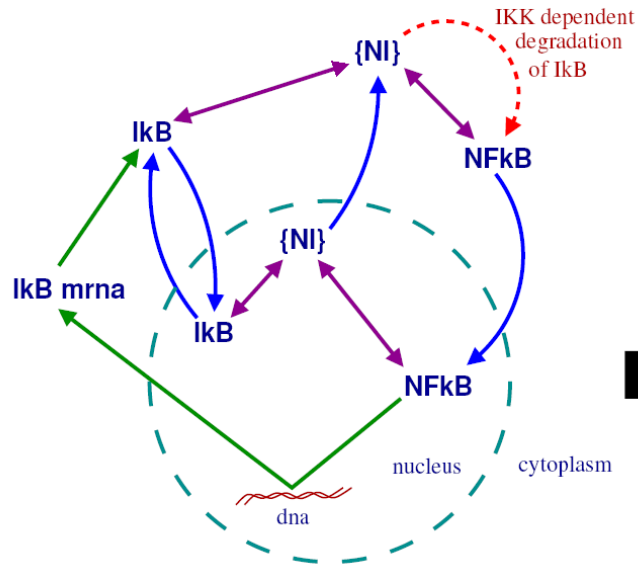
The NF- κ B System in Mammalian Cells



Nelson et al. (2004) *Science* 306, 704.

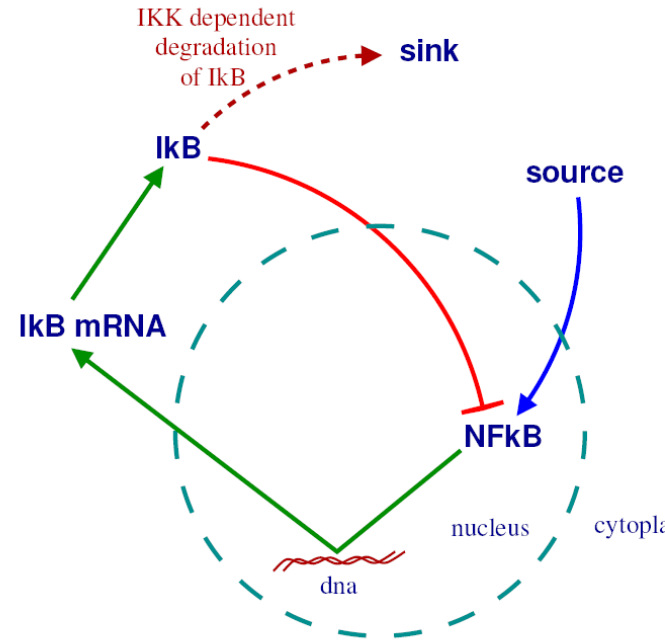
Reduction of the NF- κ B system

7-variable model



- complex formation/dissociation
- transport into/out of nucleus
- transcription & translation

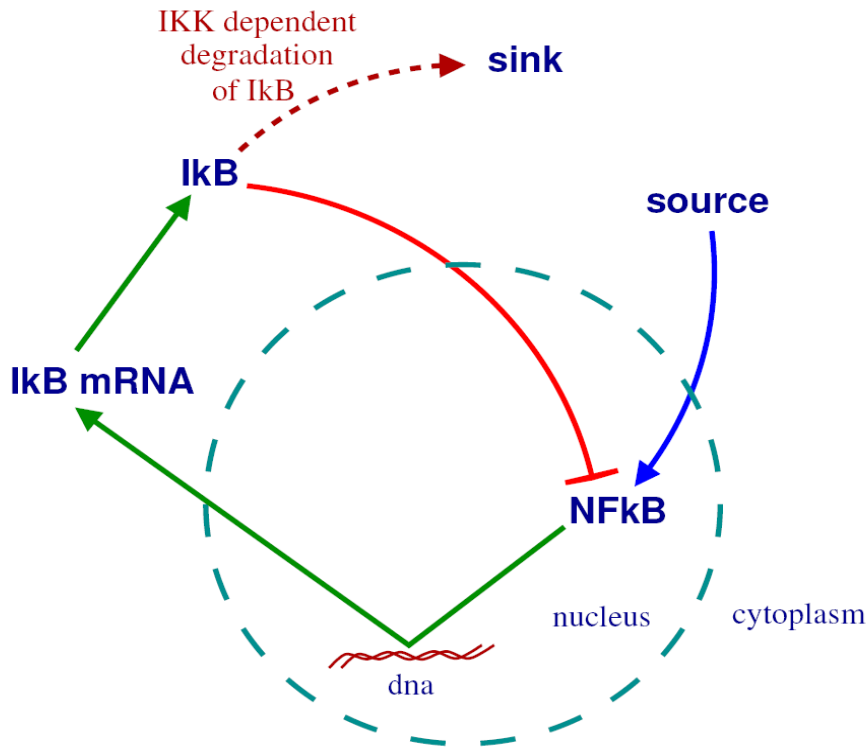
3-variable model



Remove very slow transport reactions
Assume complexes are in equilibrium

Assume certain concentrations
ratios are constant

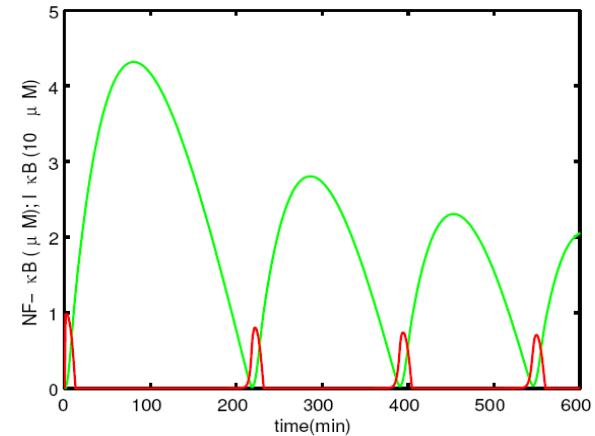
Simple Model for Protein Oscillations



$$\frac{dN_n}{dt} = A \frac{(1 - N_n)}{\epsilon + I} - B \frac{IN_n}{\delta + N_n},$$

$$\frac{dI_m}{dt} = N_n^2 - I_m,$$

$$\frac{dI}{dt} = I_m - C \frac{(1 - N_n)I}{\epsilon + I}.$$

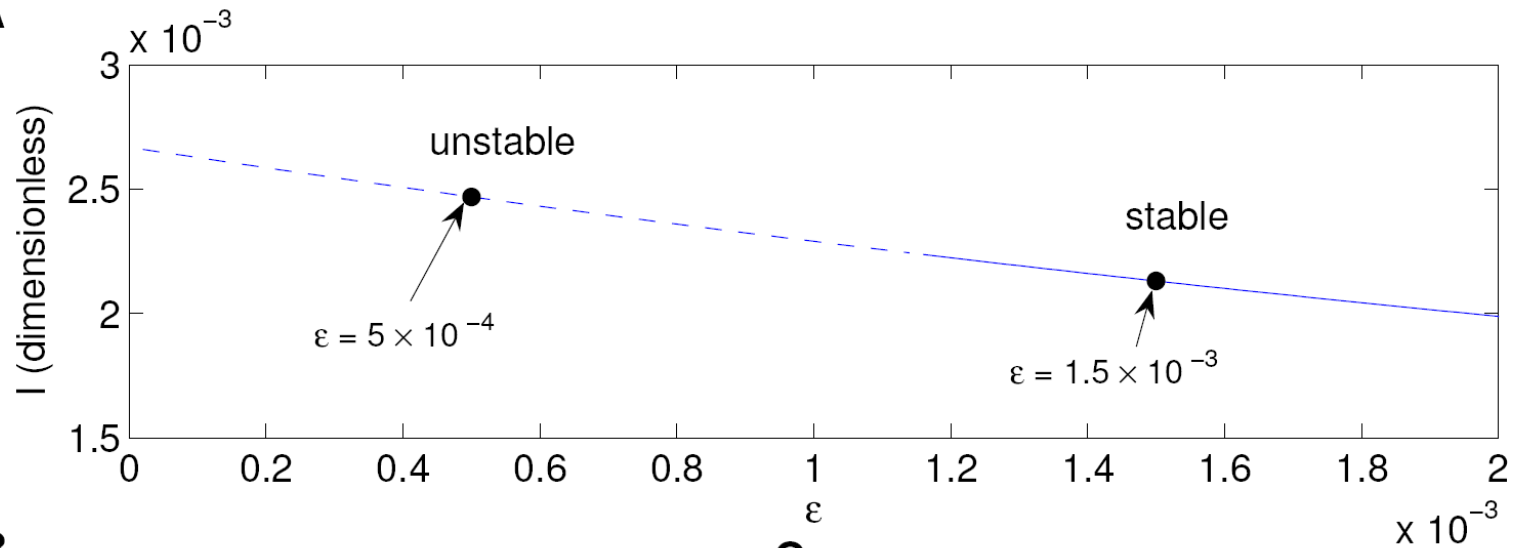


$$A = 0.007, B = 954.5, C = 0.035,$$

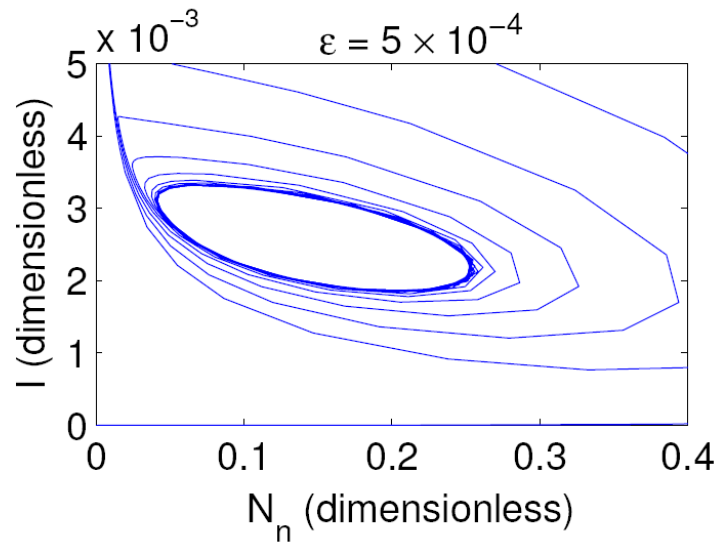
$$\delta = 0.029, \epsilon = 2 \times 10^{-5}$$

Hopf bifurcation

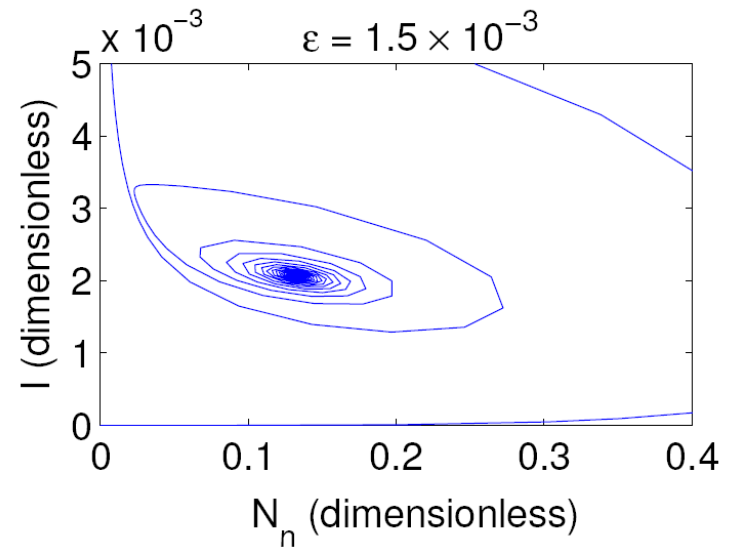
A



B

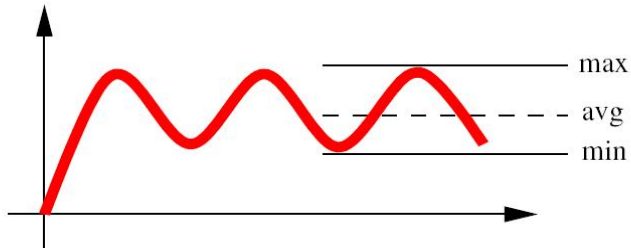
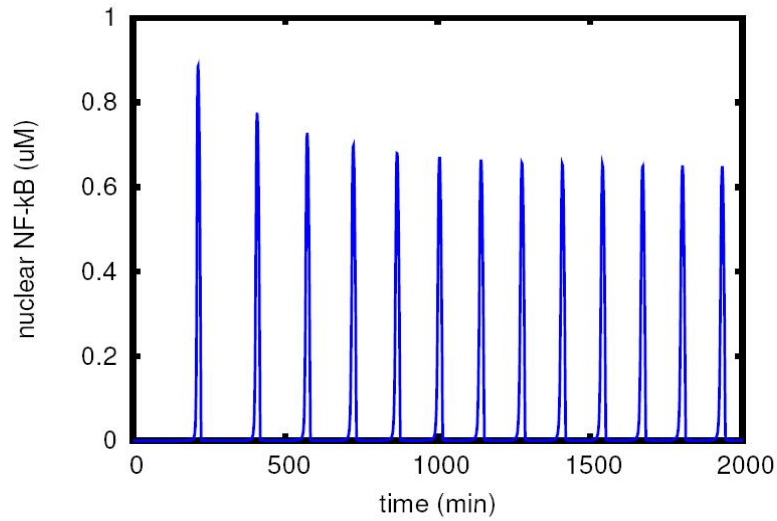


C

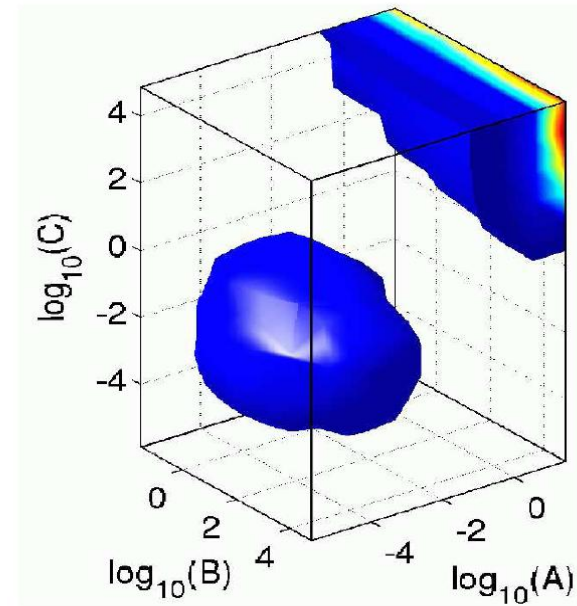
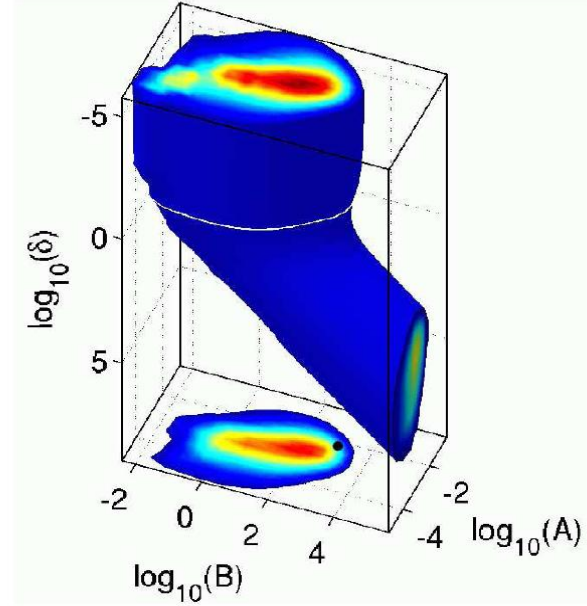


Robust Spiky Oscillations

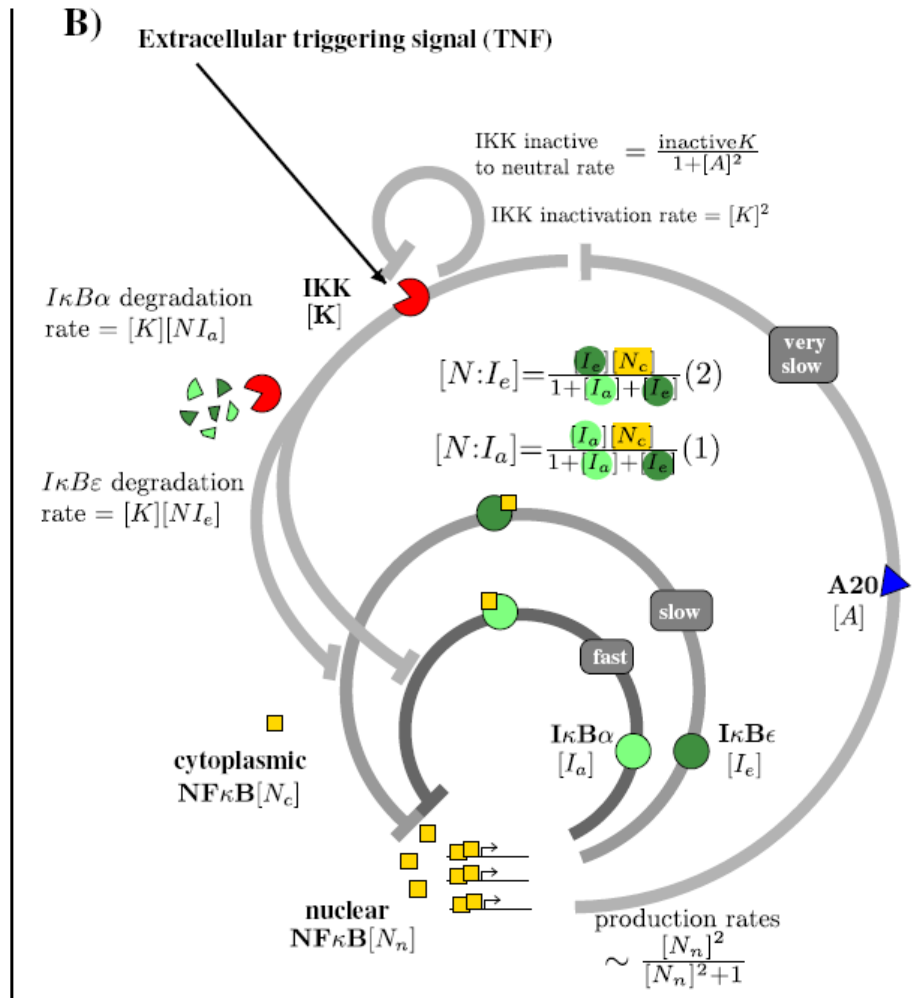
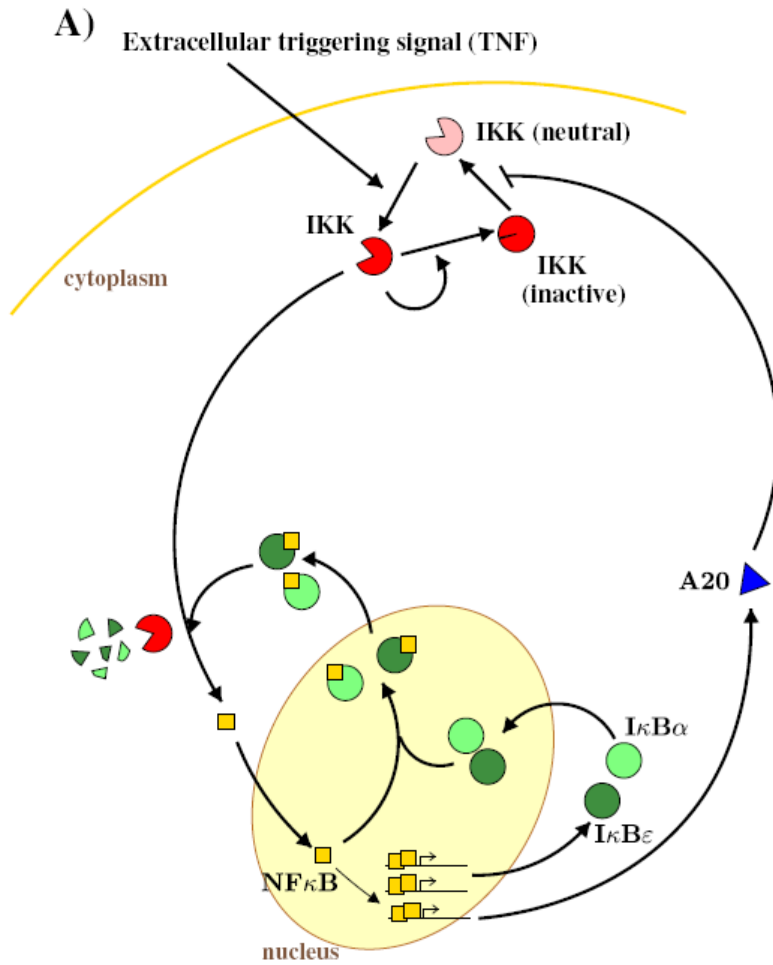
Spiky Oscillations, $Z > 2$



$$Z = \frac{\text{max} - \text{min}}{\text{avg}}$$



Nested feed-back loops

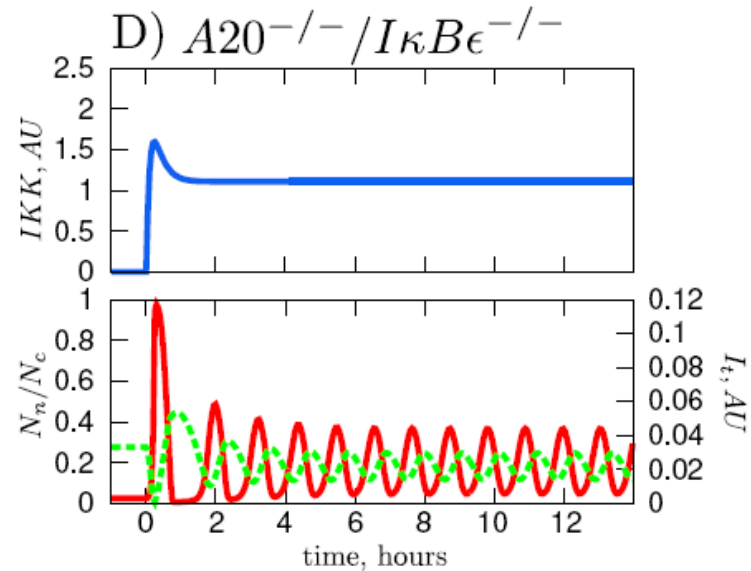
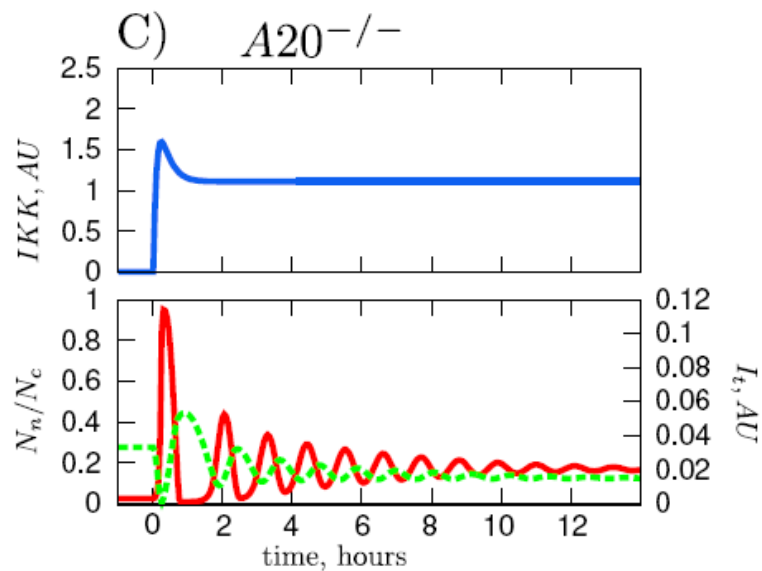
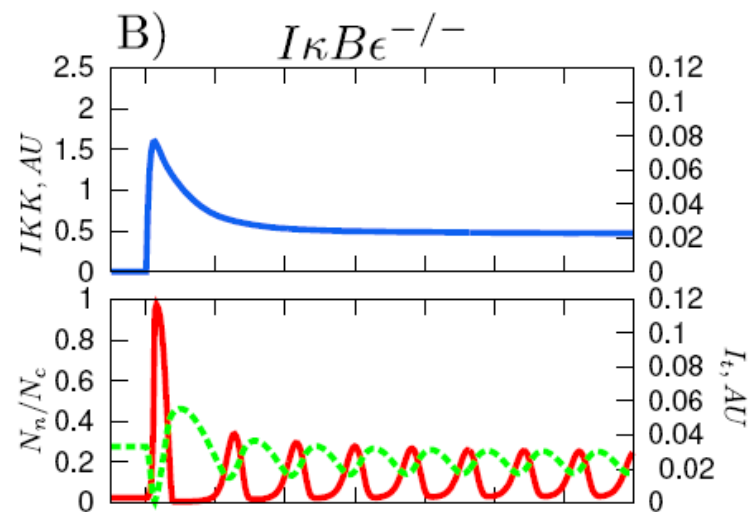
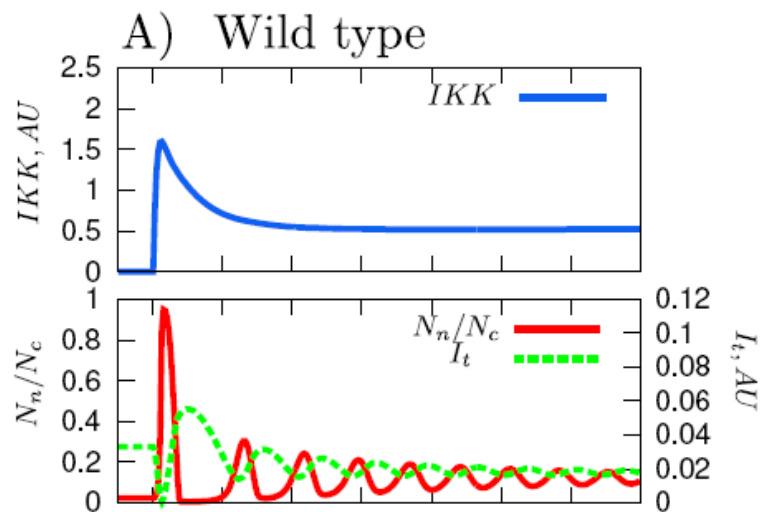


$\xrightarrow{\text{NF}\kappa\text{B}}$
regulated promoters

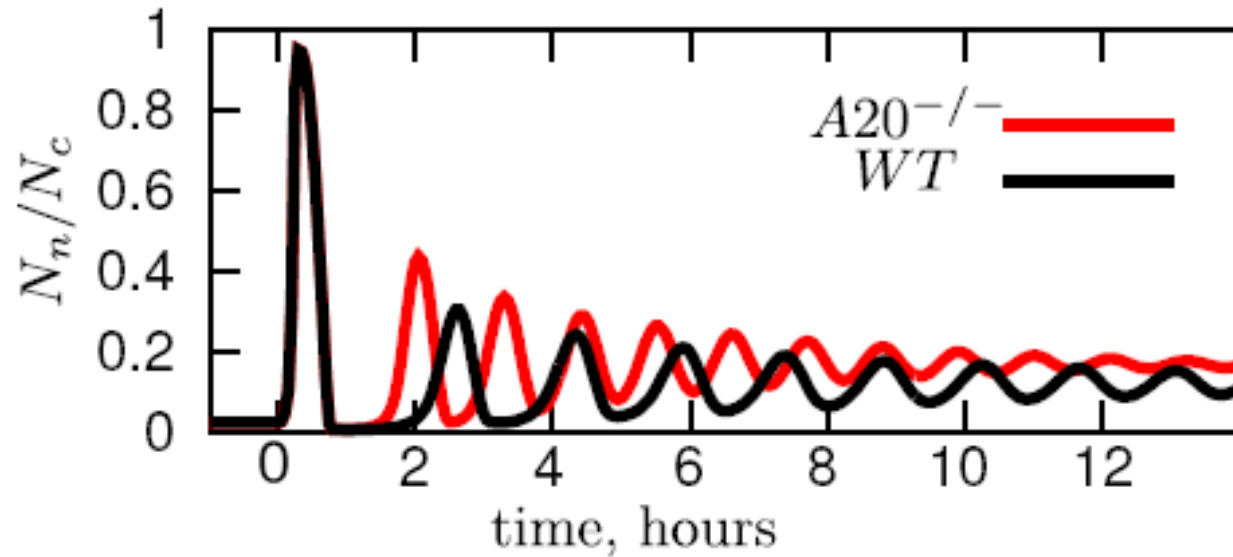
Interactions
 \longrightarrow activating
 $\longrightarrow|$ inhibiting

$\longrightarrow|$ Effective negative regulations

(...) Concentrations



A20 regulates period of NF- κ B oscillations



Single cells !

IKK has been observed to oscillate itself

$$\frac{dN_n}{dt} = A \frac{(1 - N_n)}{\epsilon + I} - B \frac{IN_n}{\delta + N_n},$$

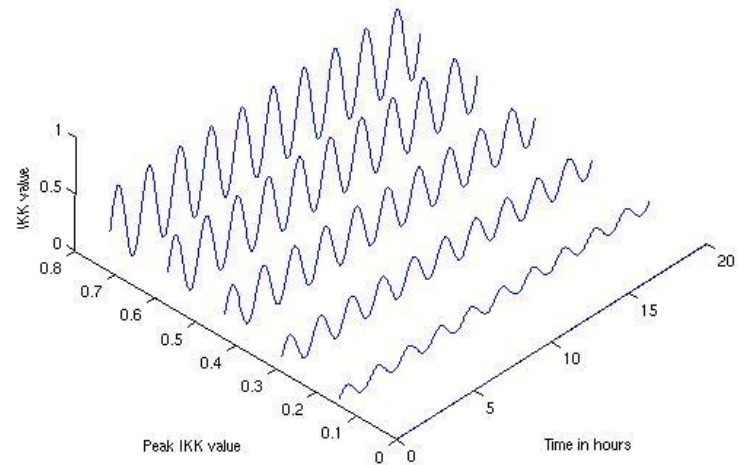
$$\frac{dI_m}{dt} = N_n^2 - I_m,$$

$$\frac{dI}{dt} = I_m - C \frac{(1 - N_n)I}{\epsilon + I}.$$

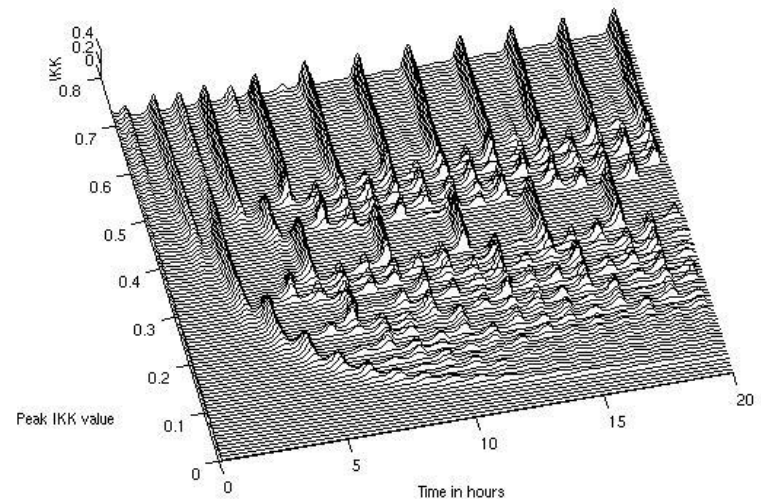
$$C \rightarrow C(1 + \sin 2\pi\omega t)$$

(Kristian Rud, Jesper Fonslet)

IKK profiles (2).



NF-kB response to oscillating IKK (3).



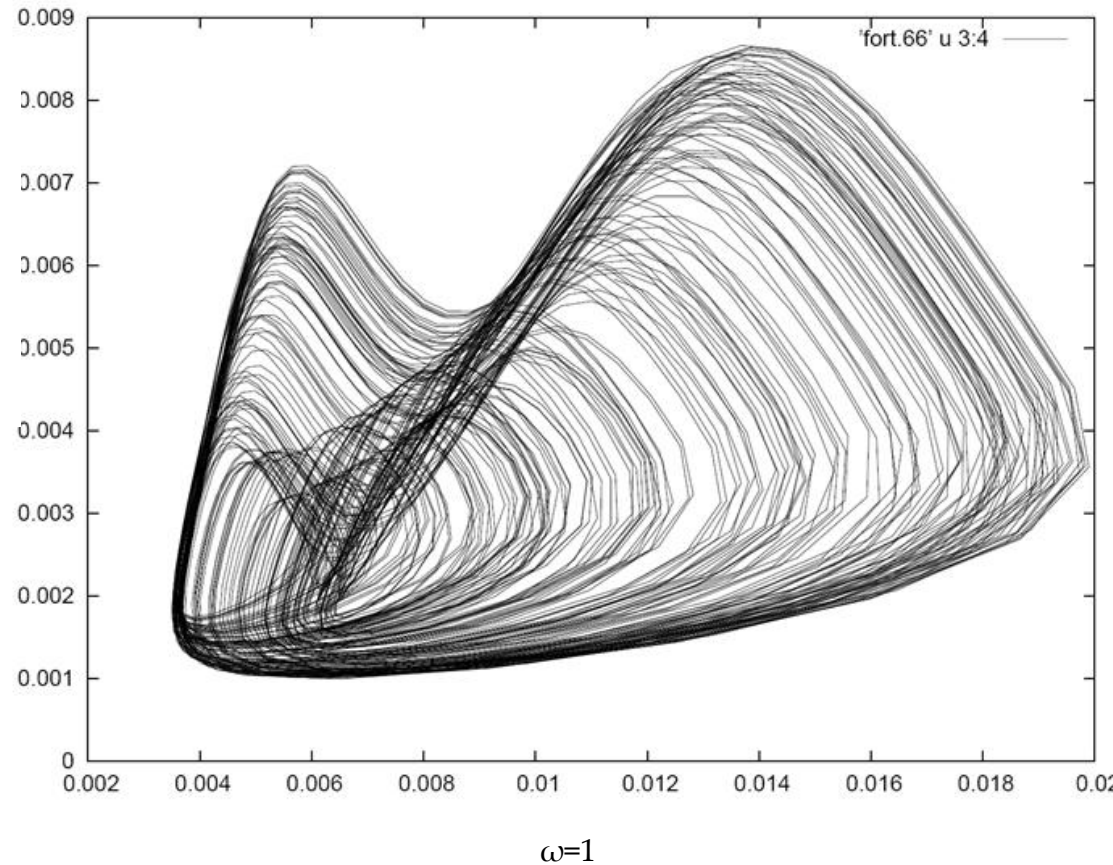
Strange attractor of periodically 'forced' NF-κB system

$$\frac{dN_n}{dt} = A \frac{(1 - N_n)}{\epsilon + I} - B \frac{IN_n}{\delta + N_n},$$

$$\frac{dI_m}{dt} = N_n^2 - I_m,$$

$$\frac{dI}{dt} = I_m - C \frac{(1 - N_n)I}{\epsilon + I}.$$

$$C \rightarrow C(1 + \sin 2\pi\omega t)$$

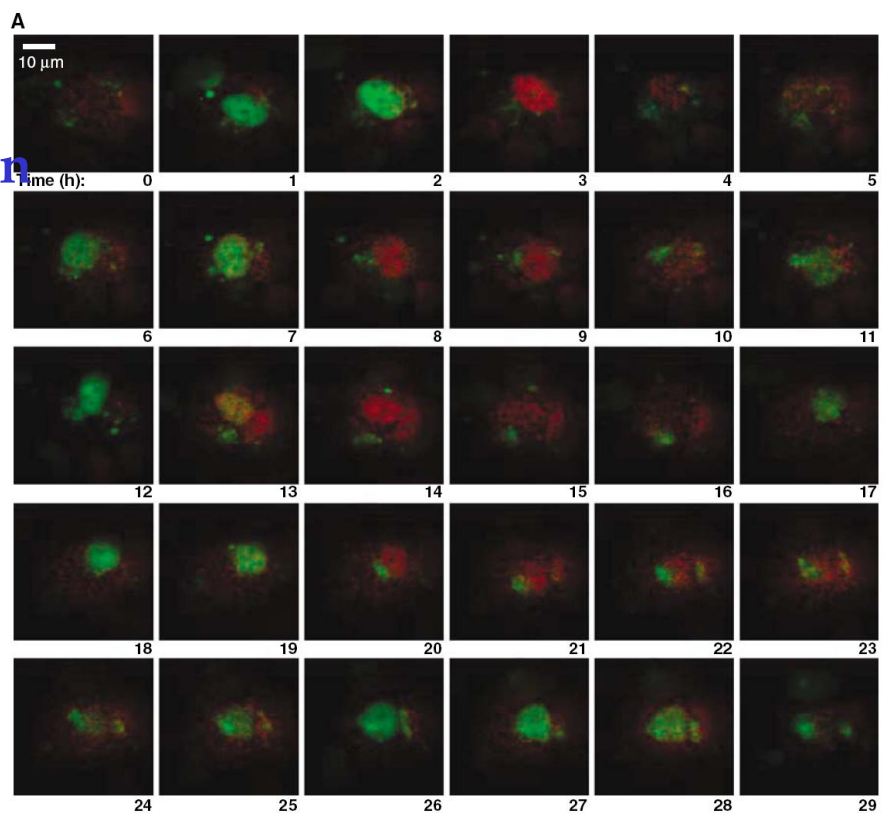


$$A = 0.007, B = 954.5, C = 0.035,$$

(2-3 hour period)

$$\delta = 0.029 \text{ and } \epsilon = 2 \times 10^{-5}$$

Response to irradiation in single cells



Often time series
are very noisy !
→ Then what ?

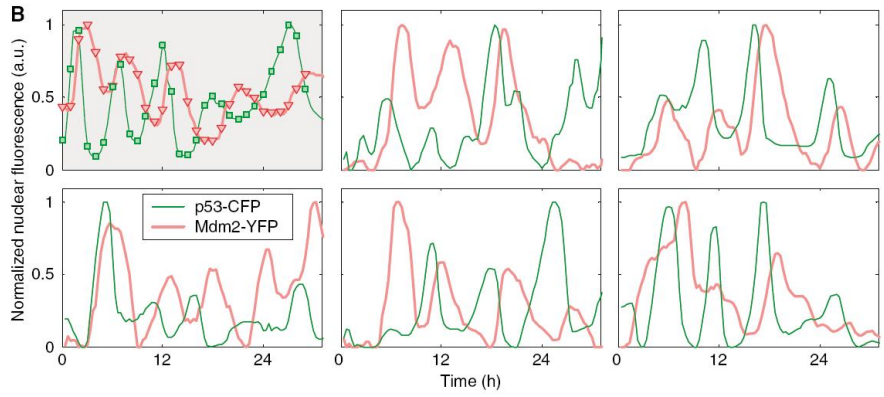
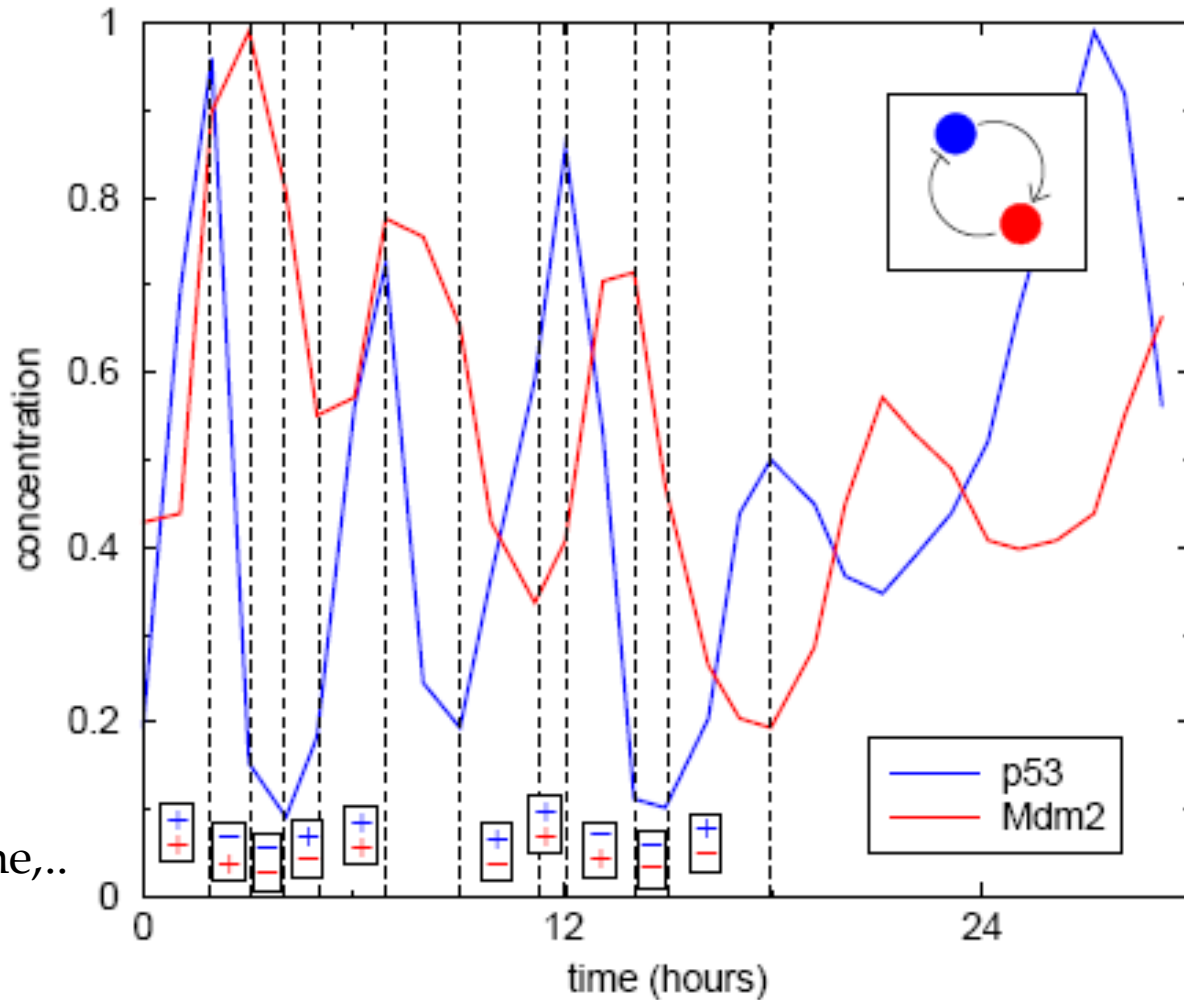


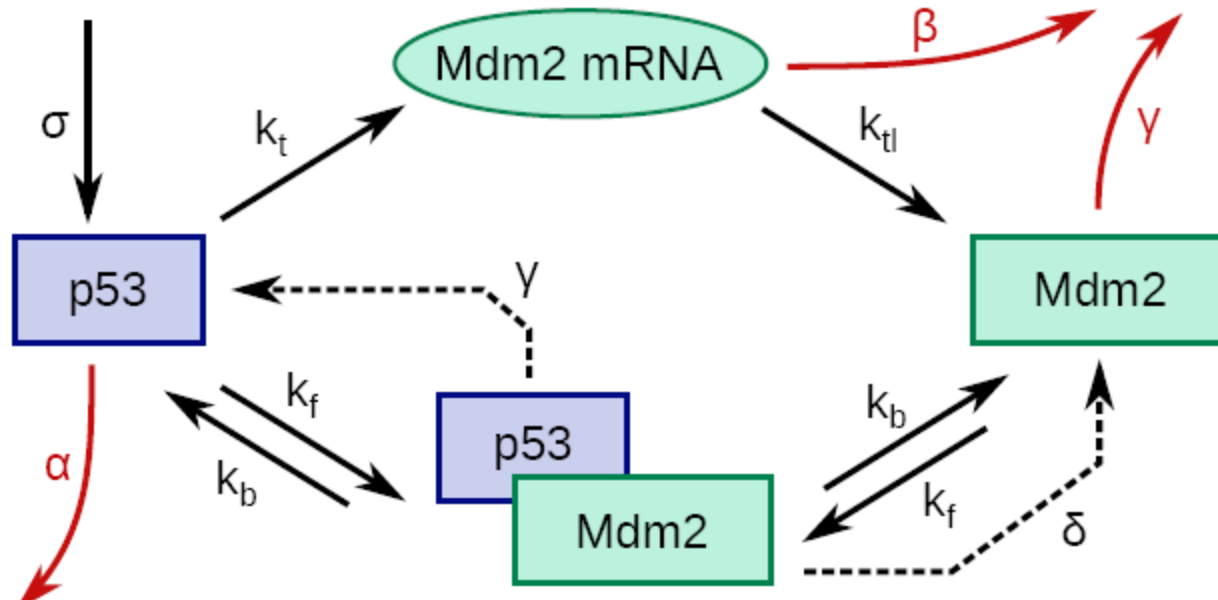
Figure 1 Prolonged oscillations in the nuclear levels of fluorescently tagged p53 and Mdm2 in individual MCF7, U280, cells following gamma irradiation. (A) Time-lapse fluorescence images of one cell over 29 h after 5 Gy of gamma irradiation. Nuclear p53-CFP and Mdm2-YFP are imaged in green and red, respectively. Time is indicated in hours. (B) Normalized nuclear fluorescence levels of p53-CFP (green) and Mdm2-YFP (red) following gamma irradiation. Top left: the cell shown in panel A. Other panels: five cells from one field of view, after exposure to 2.5 Gy gamma irradiation.

Apoptosis

(a) N. Geva-Zatorsky, *Mol. Sys. Biol.* 2:2006.0033 (2006)



Lahav, Alon, Levine,...



$$\frac{dp}{dt} = \sigma - \alpha p - k_f p m + k_b c + \gamma c$$

$$\frac{dm_m}{dt} = k_t p^2 - \beta m_m$$

$$\frac{dm}{dt} = k_{tI} m_m - k_f p m + k_b c + \delta c - \gamma m$$

$$\frac{dc}{dt} = k_f p m - k_b c - \delta c - \gamma c$$

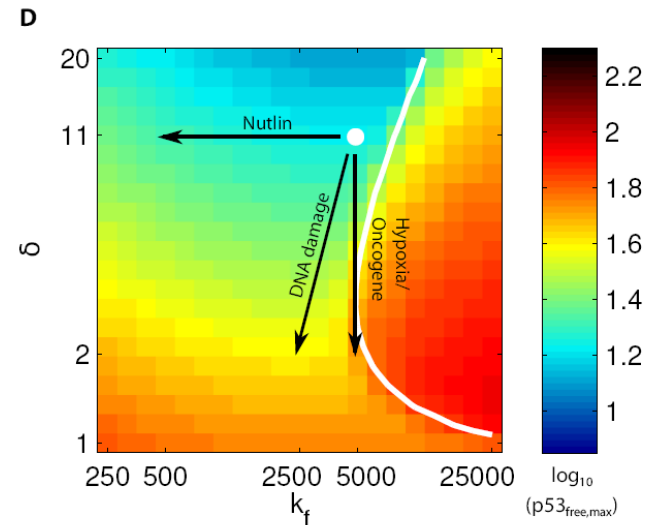
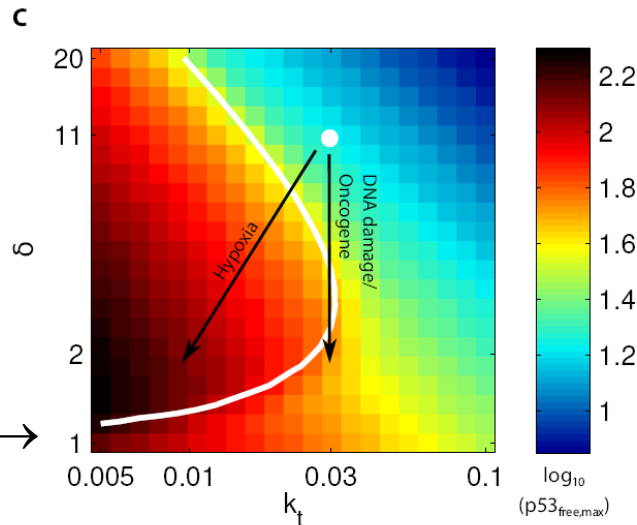
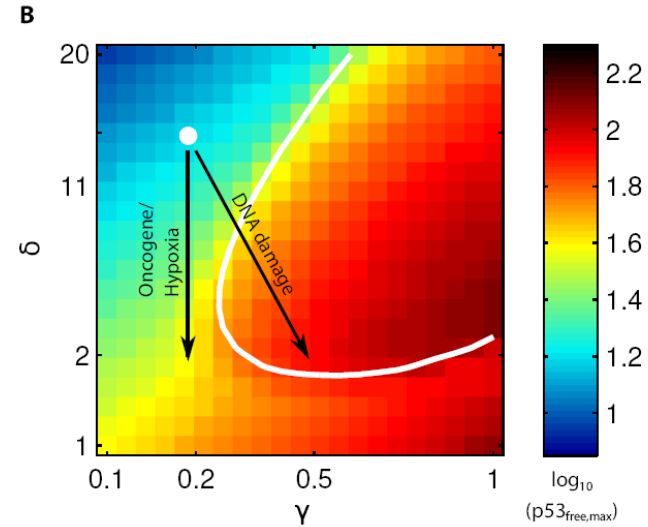
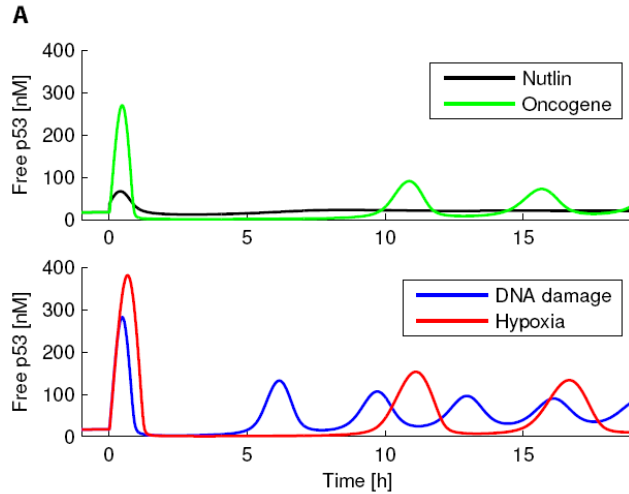
Mdm2 regulates both activity and stability

tions: nuclear-p53, p ; Mdm2, m ; Mdm2 mRNA, m_m ; and the p53-Mdm2 complex, c . The tempo-

Four different stresses

- DNA damage: Irradiation triggers oscillations: increase auto-ubiq. of Mdm2, decrease ubiq. of p53 by Mdm2, weak p53-Mdm2 binding
- Hypoxia: Deprive oxygen, apoptosis: decrease transactivation, prevents degrad. of p53
- Oncogenes: trigger 53 pathway: decrease Mdm2-dependent degrad.
- Nutlin (chemical), cell-cycle arrest (not apoptosis): reduces p53-Mdm2 binding

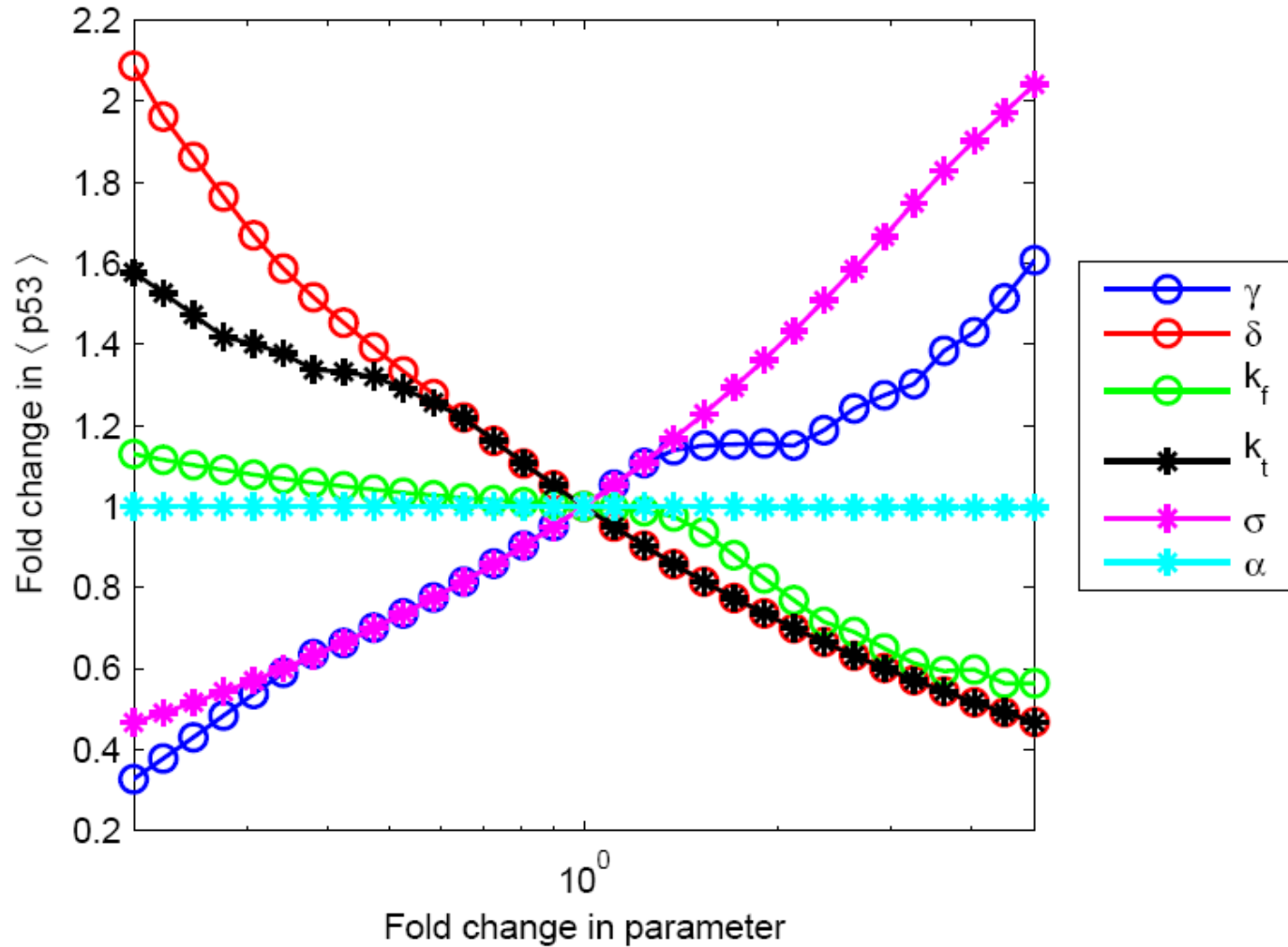
Stress variations in parameters



Spikyness \rightarrow

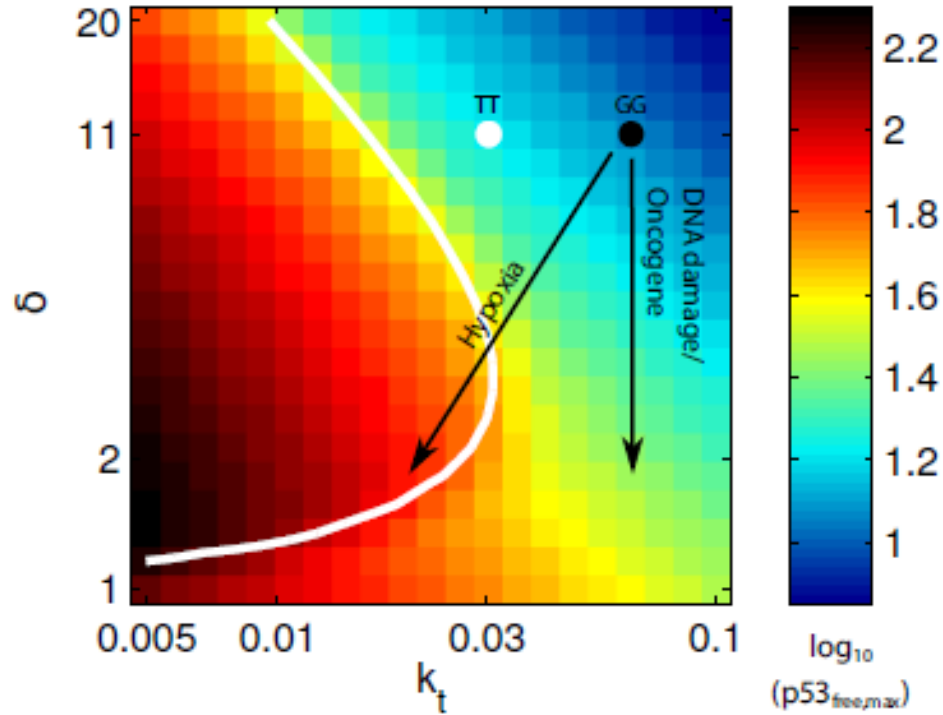
p53 concentrations (peak)

Responses:



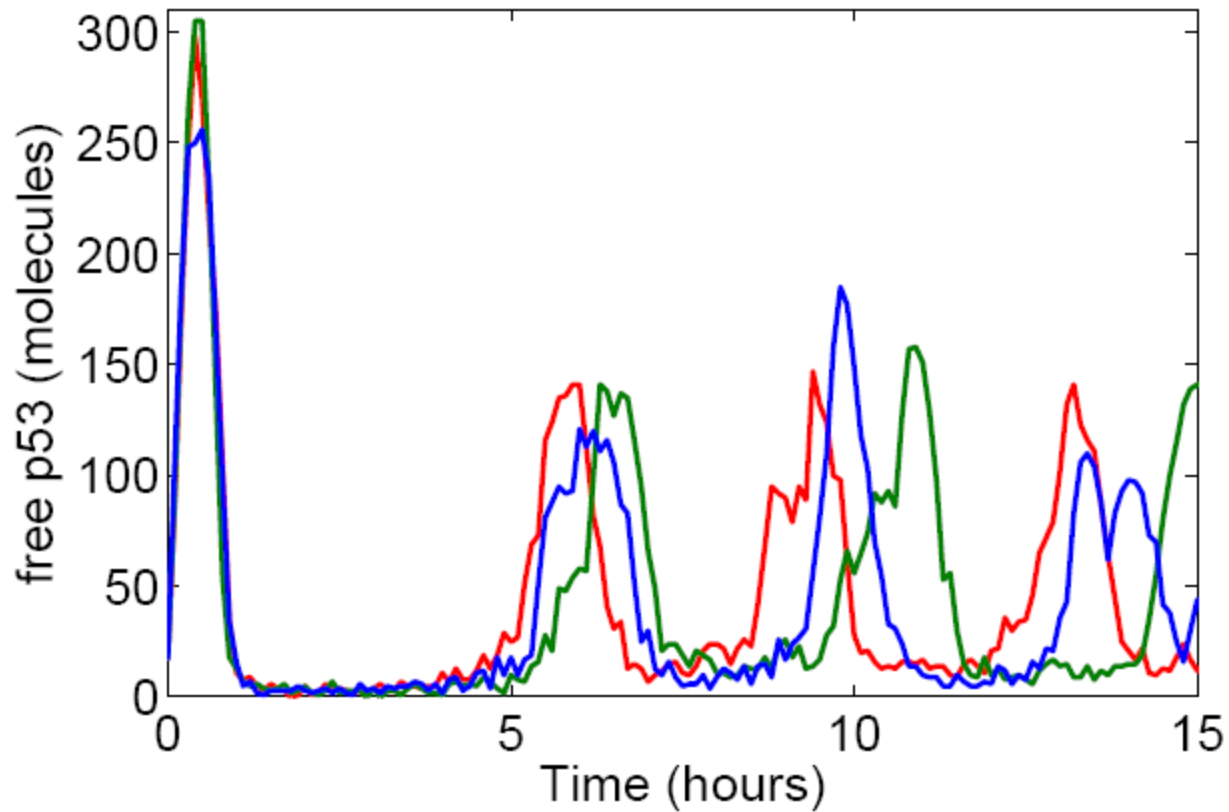
δ : DNA damage, hypoxia, oncogenes

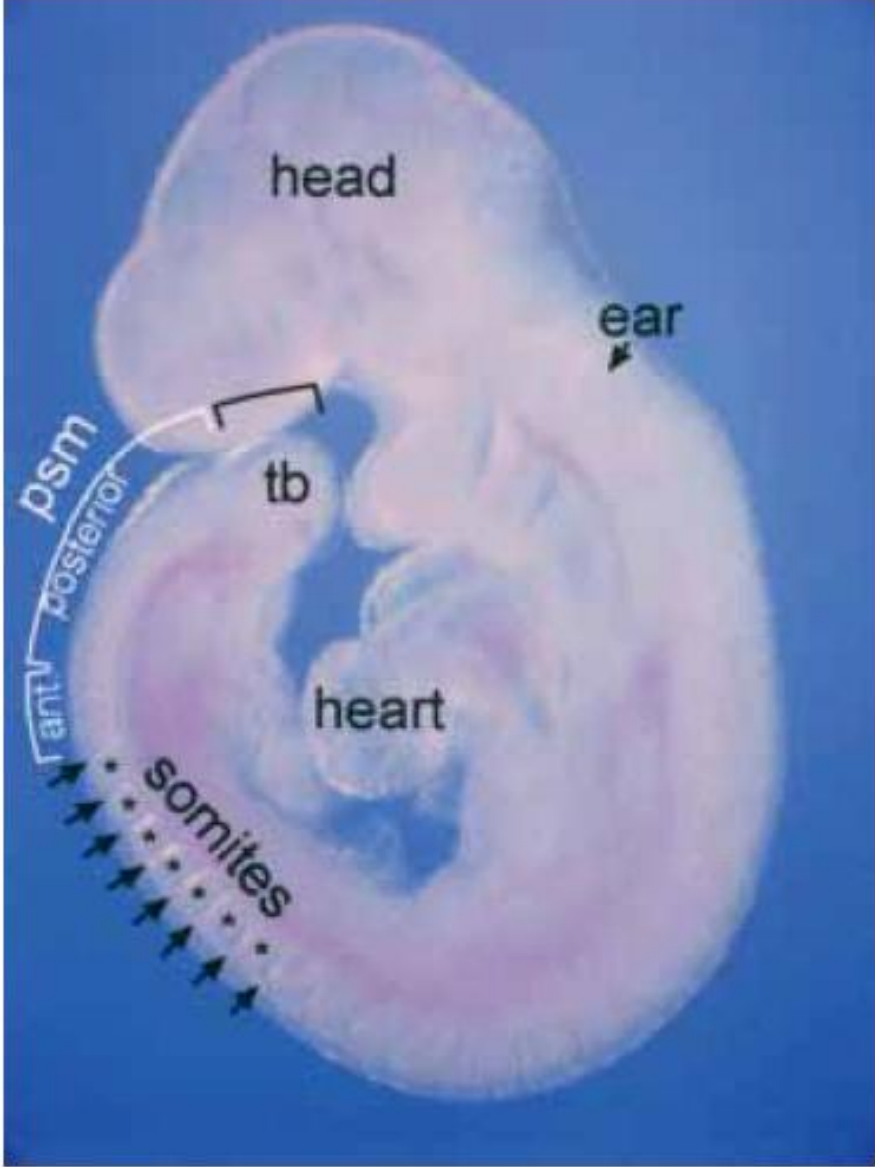
Mdm2 SNP309 allele

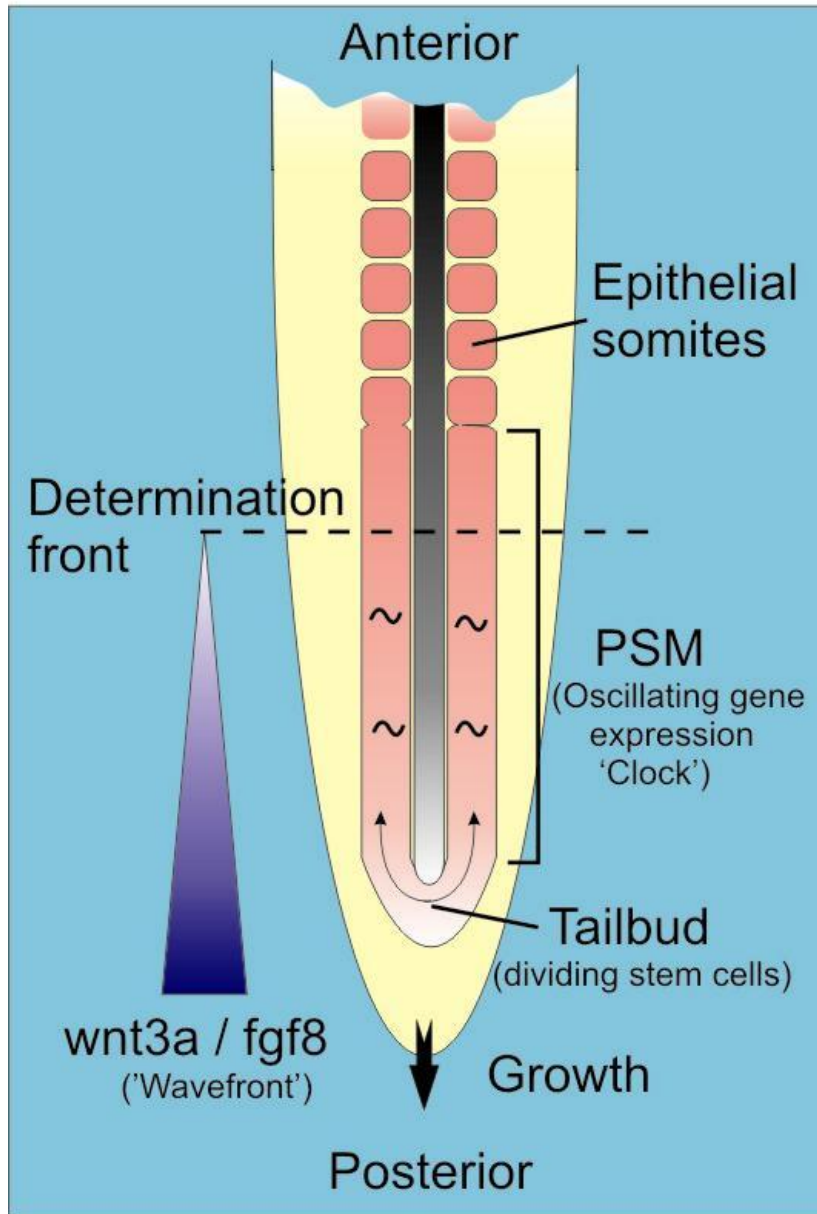


Weaker response for GG than for TT

Stochastic simulations (for p53!!)







A clock and wavefront

(Cooke and Zeeman 1976)

The presomitic mesoderm (PSM) segments anterior-posterior as somites bud off from the anterior end

Dividing stem cells in the tailbud supply cells to posterior PSM and elongates the embryo

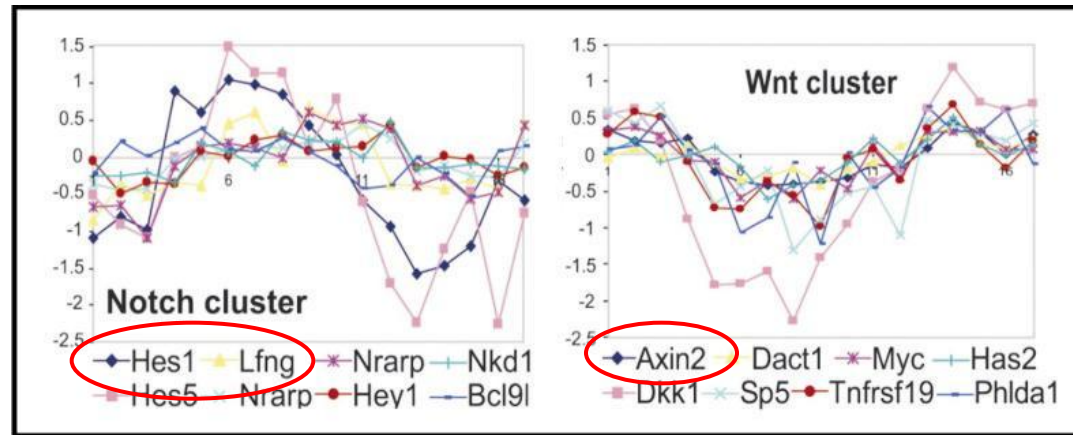
PSM cells have locally synchronized oscillating expression patterns with periods matching somite formation (90 min in chick) – **Clock**

A morphogen gradient (**Wavefront**) determines onset of segmentation program

Clock determines susceptibility to **wavefront**, which ensures groupwise incorporation into somites

Several signaling pathways are involved

Oscillating transcripts are mainly targets of Notch and Wnt pathways



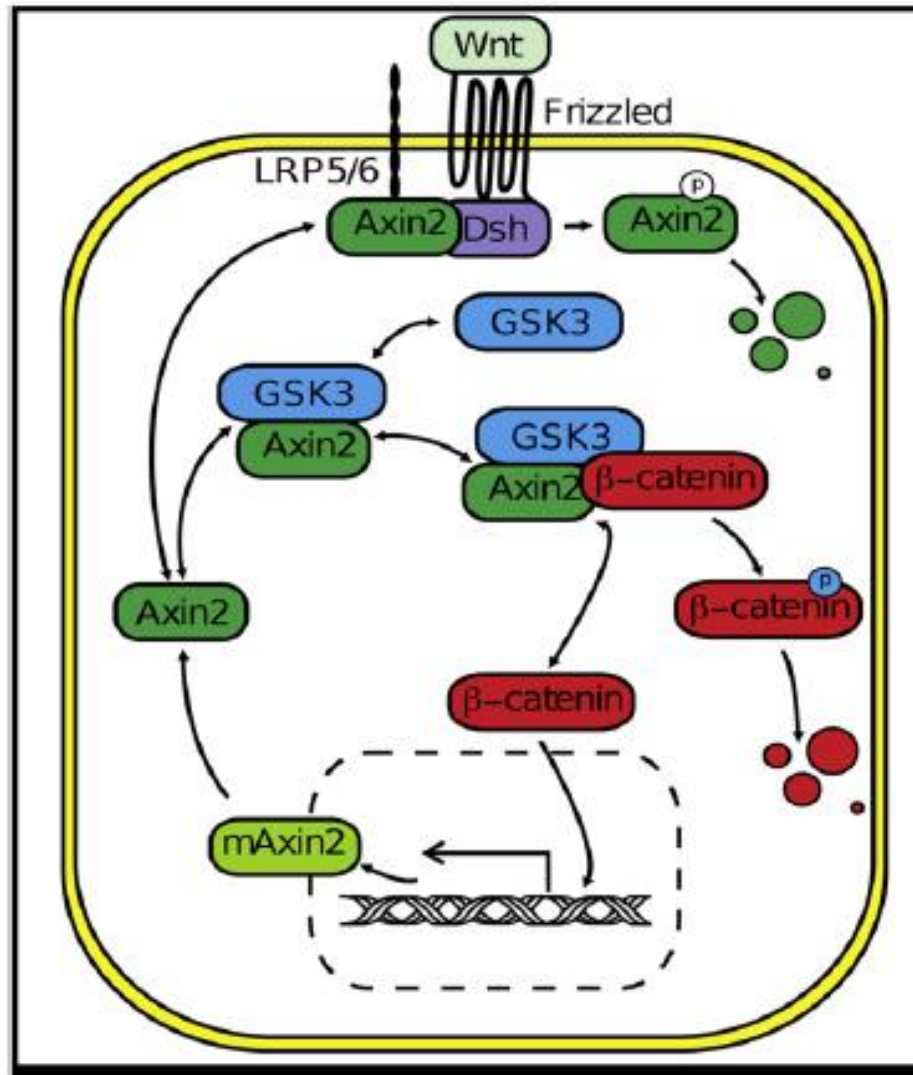
Dequèant et al. 2006

Notch and Wnt targets have same period but are 180 out of phase
➔ Crosstalk & possible hierarchical relationship

Focus is on *Hes1/7*, *Lfng* and *Axin2* that have all been associated with feedback loops

One crosstalk candidate (out of many possible) is GSK3 β , which can bind and phosphorylate both β -catenin and Notch_{ICD}

The Wnt systems



Goldbeter,
Pourquie

Equations for the Wnt system

Destruction complex $\frac{dC}{dt} = c_{fC}B[GA] - c_{bC}C - \alpha C,$ (1)

GA-complex $\frac{d[GA]}{dt} = c_{f[GA]}GA - c_{b[GA]}[GA] - c_{fC}B[GA] + c_{bC}C + \alpha C,$ (2)

β -catenin $\frac{dB}{dt} = S - c_{fC}B[GA] + c_{bC}C,$ (3)

GSK $\frac{dG}{dt} = -c_{f[GA]}GA + c_{b[GA]}[GA],$ (4)

Axin2 $\frac{dA}{dt} = -c_{f[GA]}GA + c_{b[GA]}[GA] + c_{tIA}A_m - c_{f[AL]}AL$
 $+ c_{b[AL]}[AL],$ (5)

$$\frac{dA_m}{dt} = c_{tsA}B^2 - \frac{A_m}{\tau_{Am}},$$
 (6)

$$\frac{d[AL]}{dt} = c_{f[AL]}AL - c_{b[AL]}[AL] - \nu[AL],$$
 (7)

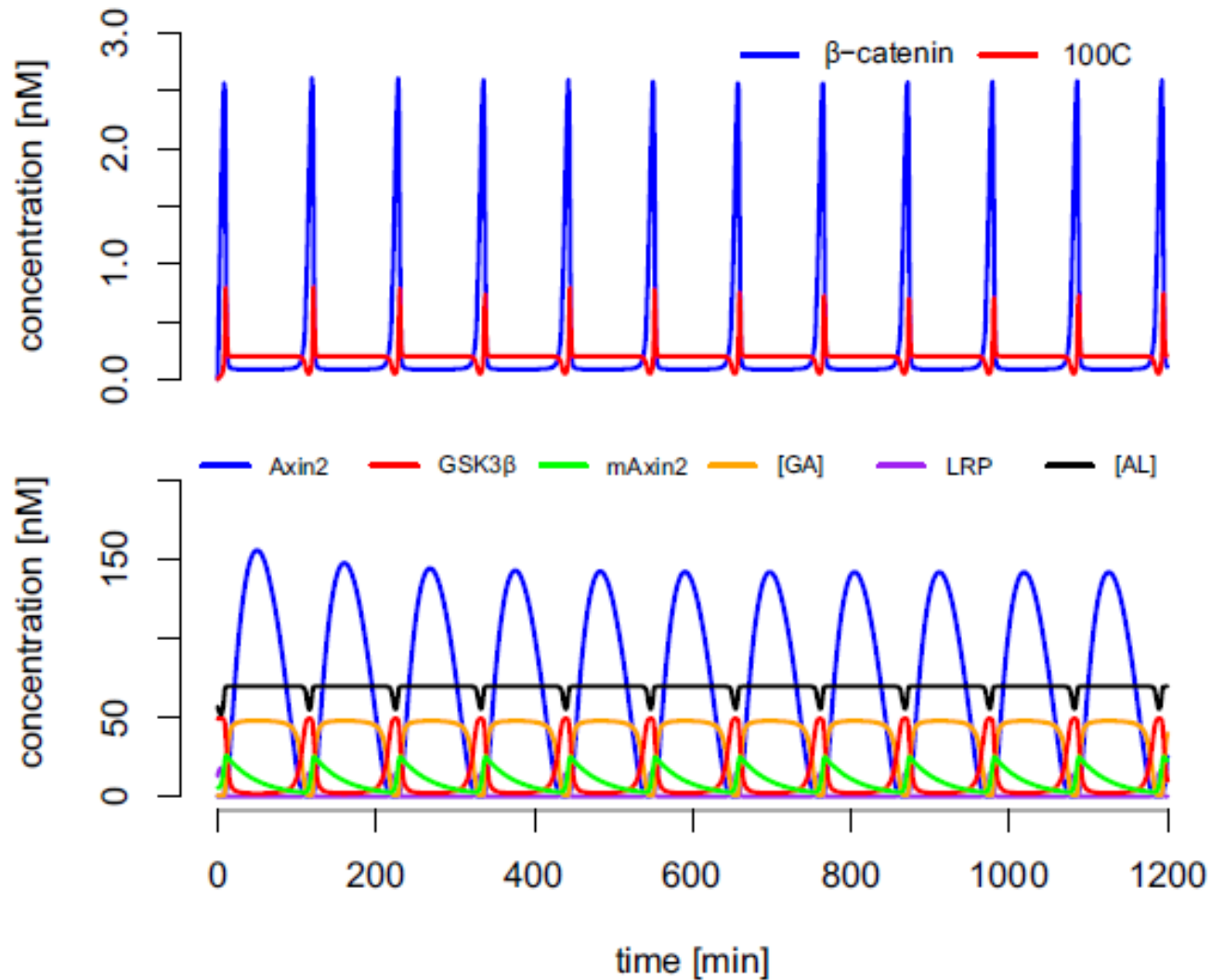
Axin2-Wnt

$$\frac{dL}{dt} = -c_{f[AL]}AL + c_{b[AL]}[AL] + \nu[AL],$$
 (8)

TABLE 1 Parameters in the eight-variable model of the Wnt system and their default values [4]

Parameter	Process	Default value
c_{fC}	Binding of B to [GA] to form destruction complex C	$0.1 \text{ nM}^{-1} \text{ min}^{-1}$
c_{bC}	Dissociation of C into B and [GA]	7 min^{-1}
α	Dissociation of C due to destruction of β -catenin	200 min^{-1}
$c_{f[GA]}$	Binding of G to A to form [GA]	$0.2 \text{ nM}^{-1} \text{ min}^{-1}$
$c_{b[GA]}$	Dissociation of [GA] into G and A	1.2 min^{-1}
S	Constant source of β -catenin	0.4 nM min^{-1}
$c_{f[AL]}$	Binding of A to L	$10 \text{ nM}^{-1} \text{ min}^{-1}$
$c_{b[AL]}$	Dissociation of [AL] into A and L	0.08 min^{-1}
c_{tsA}	Transcription of <i>axin2</i> gene	$0.7 \text{ nM}^{-1} \text{ min}^{-1}$
c_{tlA}	Translation of Axin2 mRNA	0.7 min^{-1}
τ_{Am}	Average lifetime of Axin2 mRNA	40 min
ν	Degradation of Axin2 in [AL] complex	0.1 min^{-1}
$GSK3\beta_{tot}$	Total G level	50 nM
L_{tot}	Total L level	70 nM

The Wnt reference state



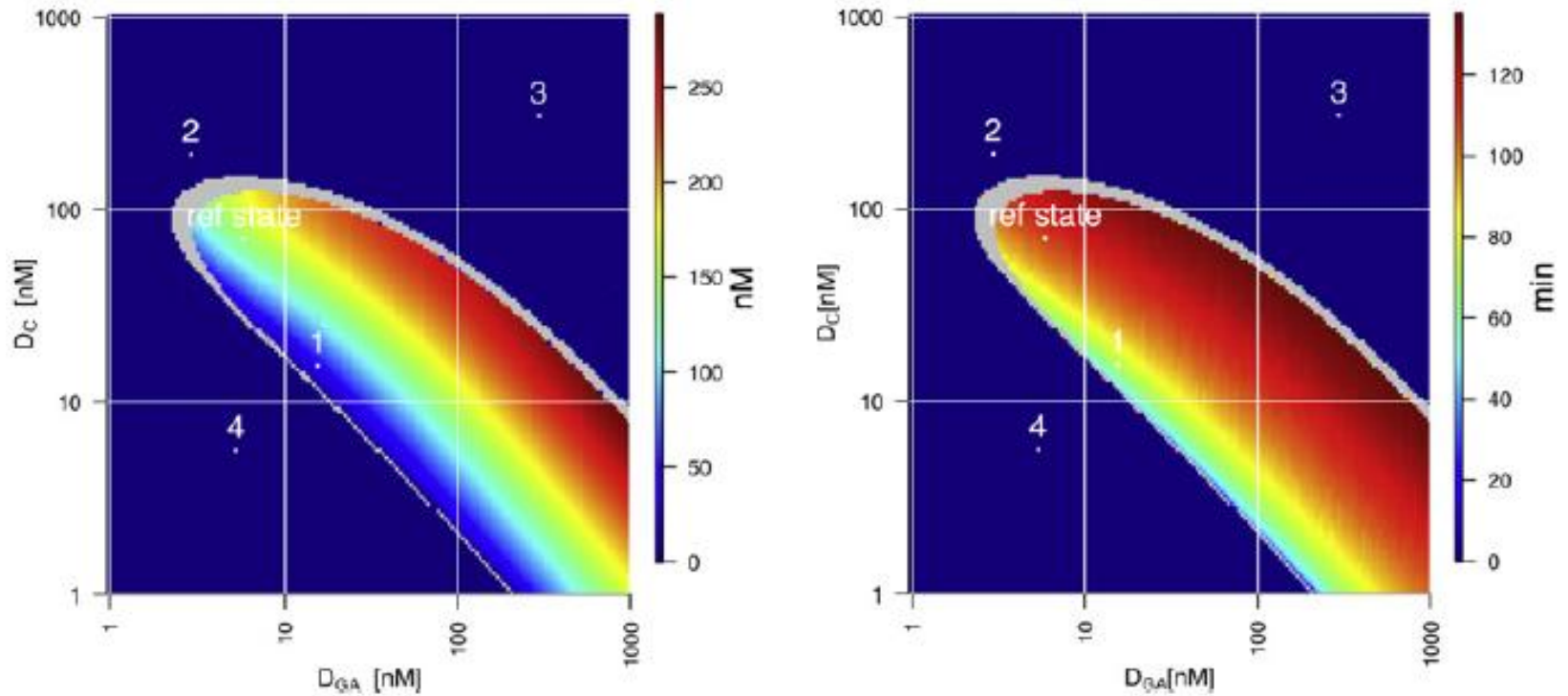
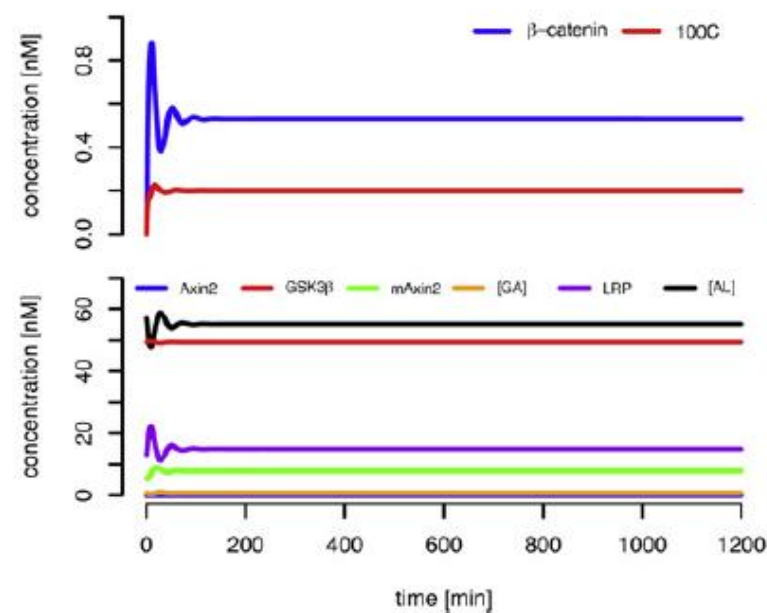
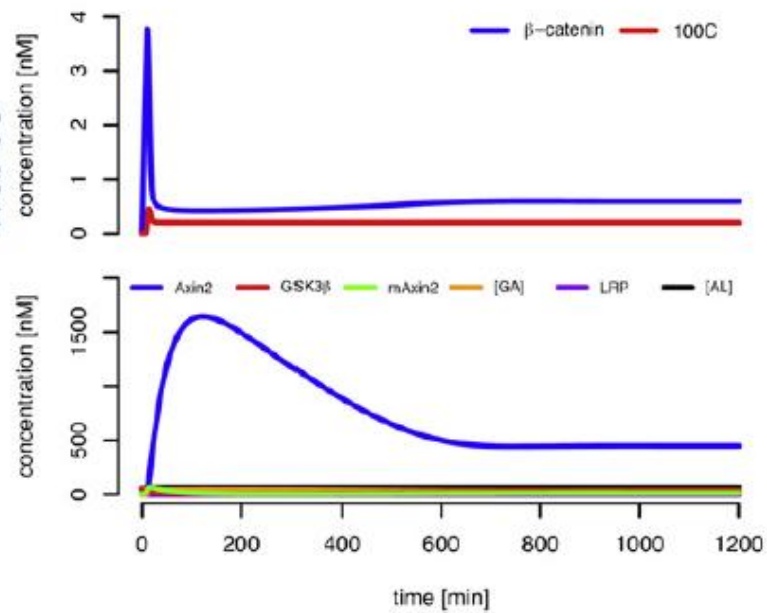
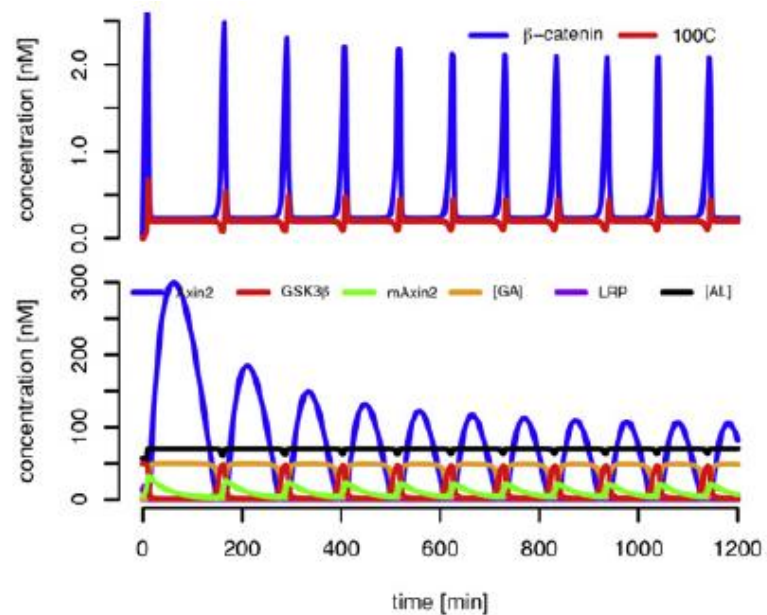
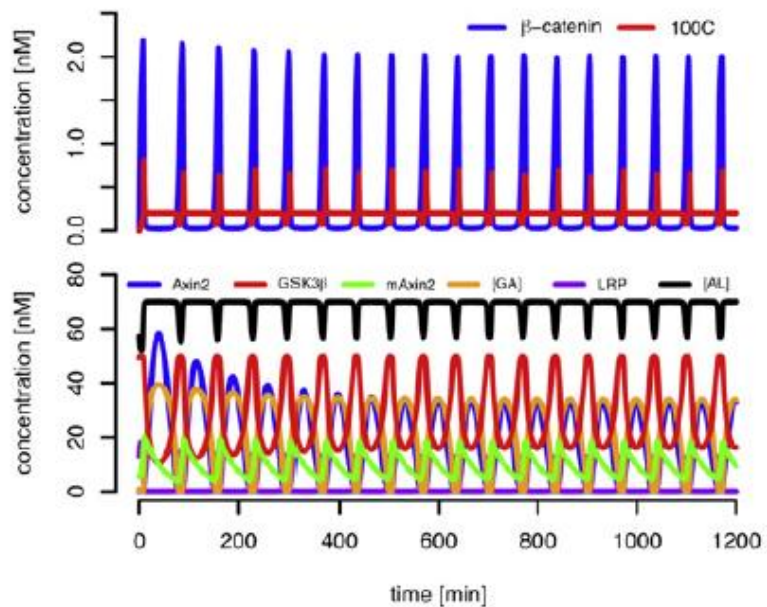


FIGURE 3 The D_C and $D_{[GA]}$ parameter plane. (*Left panel*) The amplitude of the Axin2 oscillations. (*Right panel*) Oscillation period of Axin2. The shaded borderline indicates the boundary of sustained oscillations, derived from a stability analysis of the system. The reference state (see Fig. 2) is indicated by the dot labeled “ref state”. Timeseries corresponding to the four numbered dots are shown in Fig. 4.

$$D_C = \frac{c_{bC}}{c_{fC}}$$

$$D_{[GA]} = \frac{c_{b[GA]}}{c_{f[GA]}}$$



Spatial gradient of Wnt:

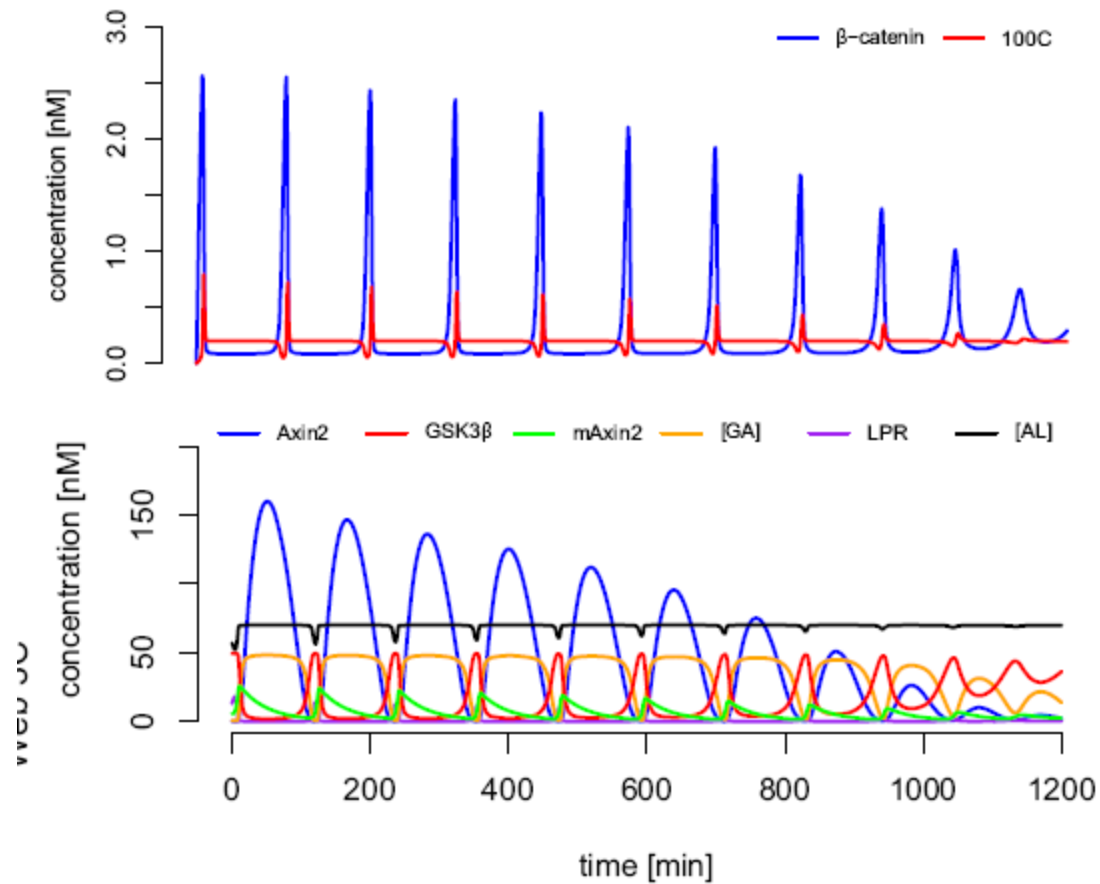
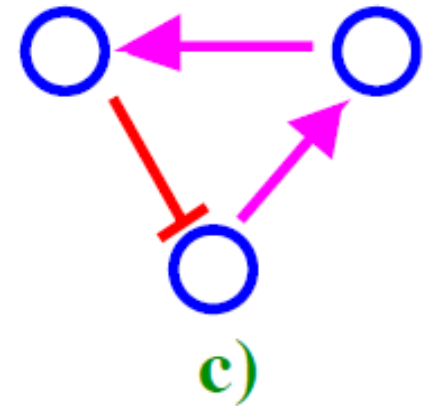
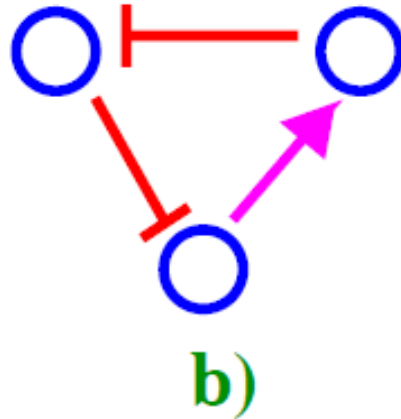
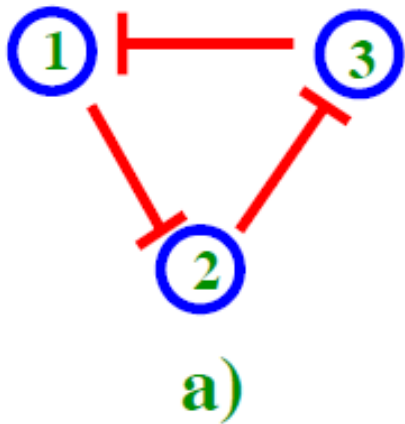


FIGURE 6 Oscillations of the Wnt variables as ν is linearly decreased from 0.1 to 0.03 min^{-1} in 1200 min. Note that the oscillations cease when the Wnt signal falls below a certain threshold level.

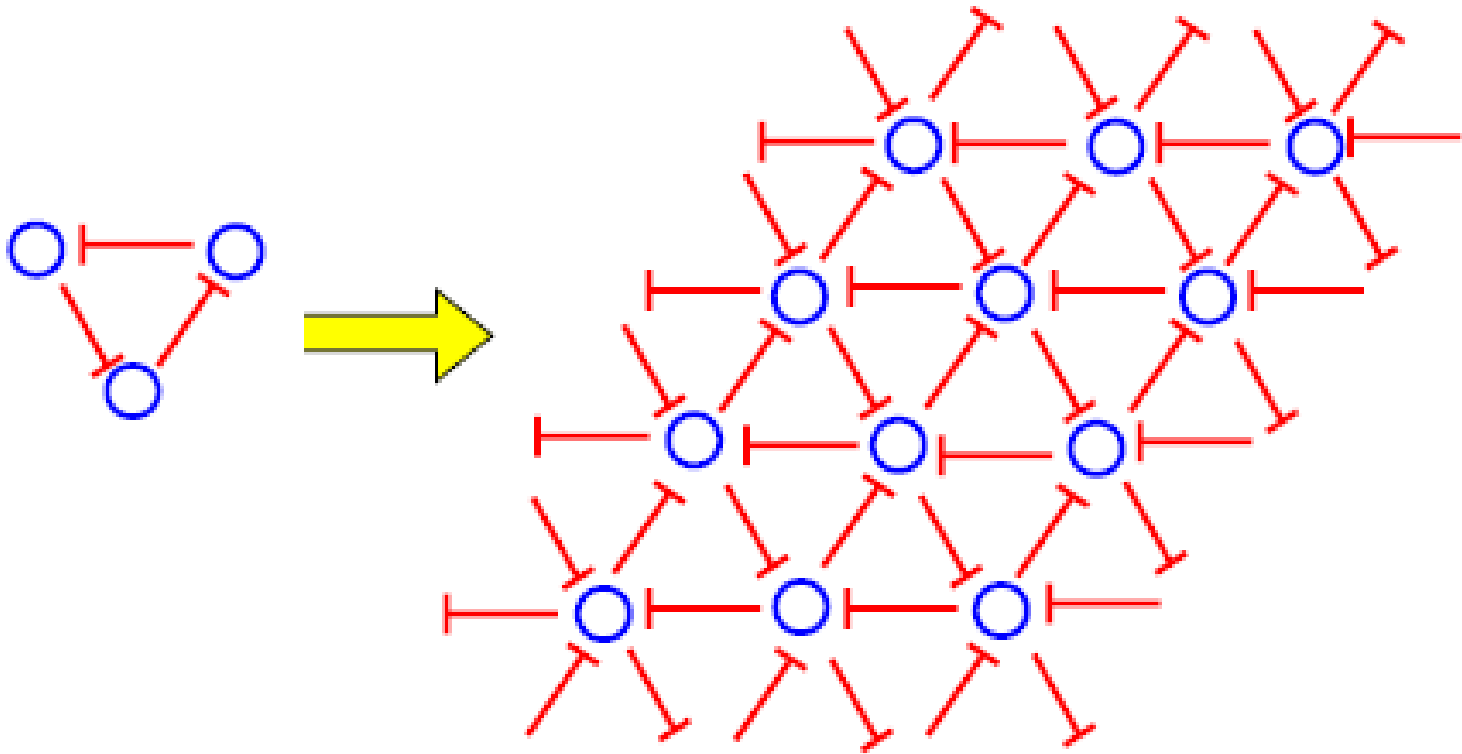
Three node motifs: Oscillations and Switches



Repressilator:
oscillations

$$\frac{dx}{dt} = c - \gamma x + \alpha \frac{1}{1 + \left(\frac{\tilde{x}}{K}\right)^h}$$

A model for cell-to-cell communications:



$$\frac{dx_{m,n}}{dt} = c - \gamma x_{m,n} + \alpha F_{\text{int}}$$

Dynamical equation for site (m,n):

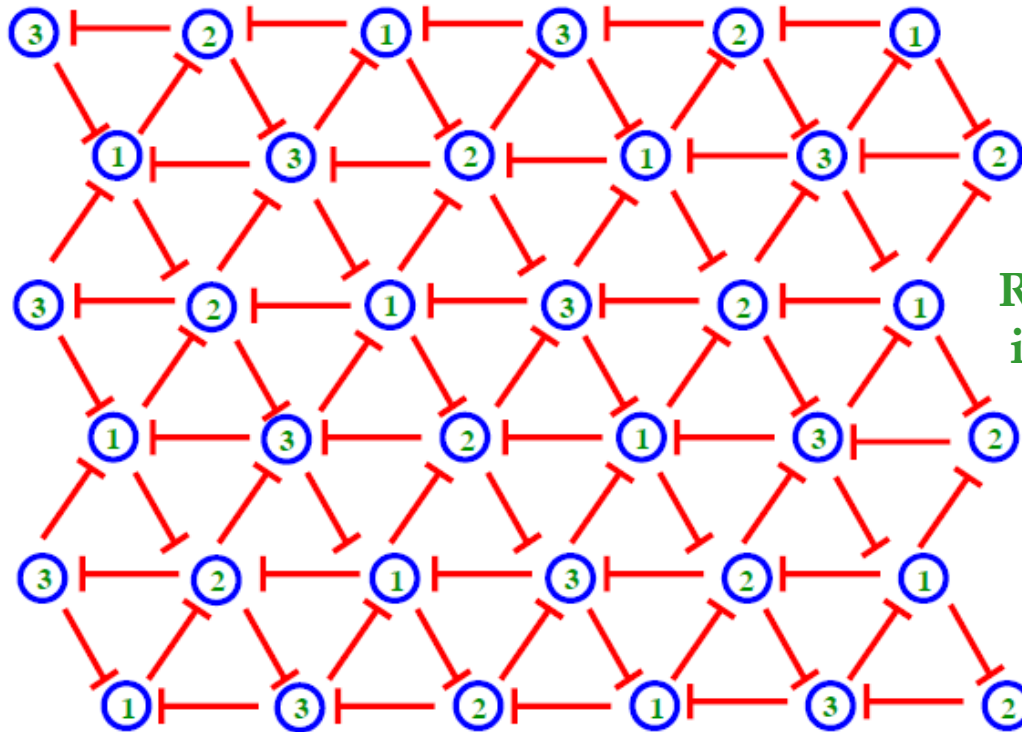
We consider two types of interaction terms—either an additive repression (an “OR gate”),

$$F_{\text{int}} = \frac{1}{1 + \left(\frac{x_{m+1,n}}{K}\right)^h} + \frac{1}{1 + \left(\frac{x_{m,n-1}}{K}\right)^h} + \frac{1}{1 + \left(\frac{x_{m-1,n+1}}{K}\right)^h}, \quad (2)$$

or a multiplicative repression (an “AND gate”),

$$F_{\text{int}} = \frac{1}{1 + \left(\frac{x_{m+1,y}}{K}\right)^h} \frac{1}{1 + \left(\frac{x_{m,n-1}}{K}\right)^h} \frac{1}{1 + \left(\frac{x_{m-1,n+1}}{K}\right)^h}. \quad (3)$$

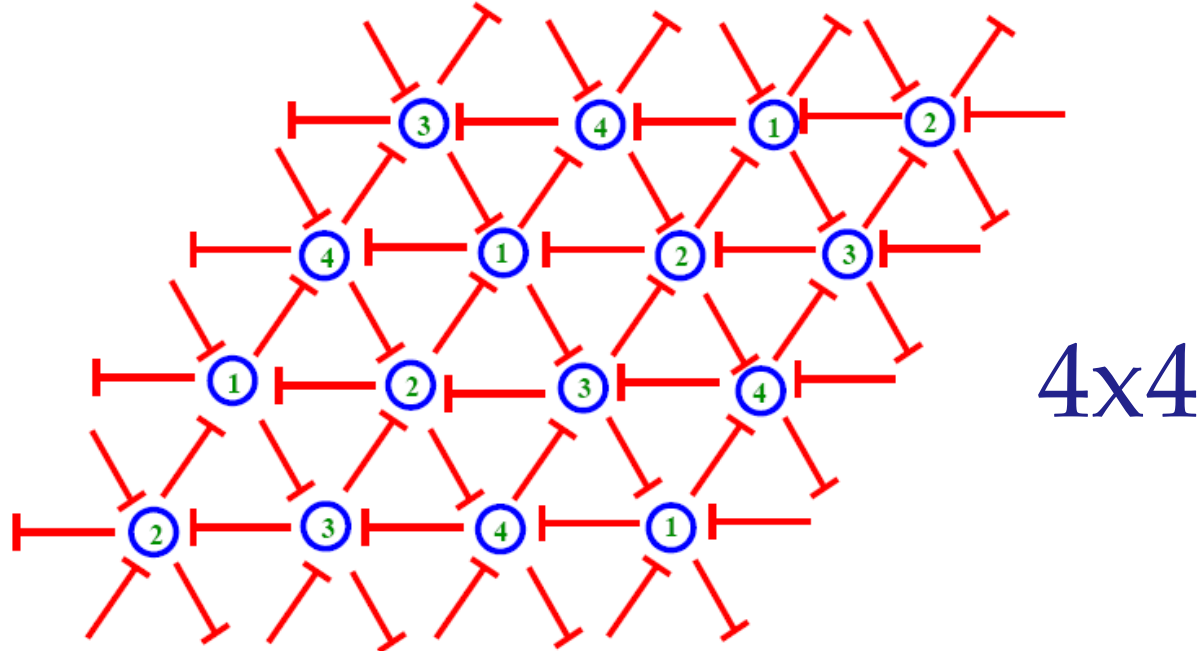
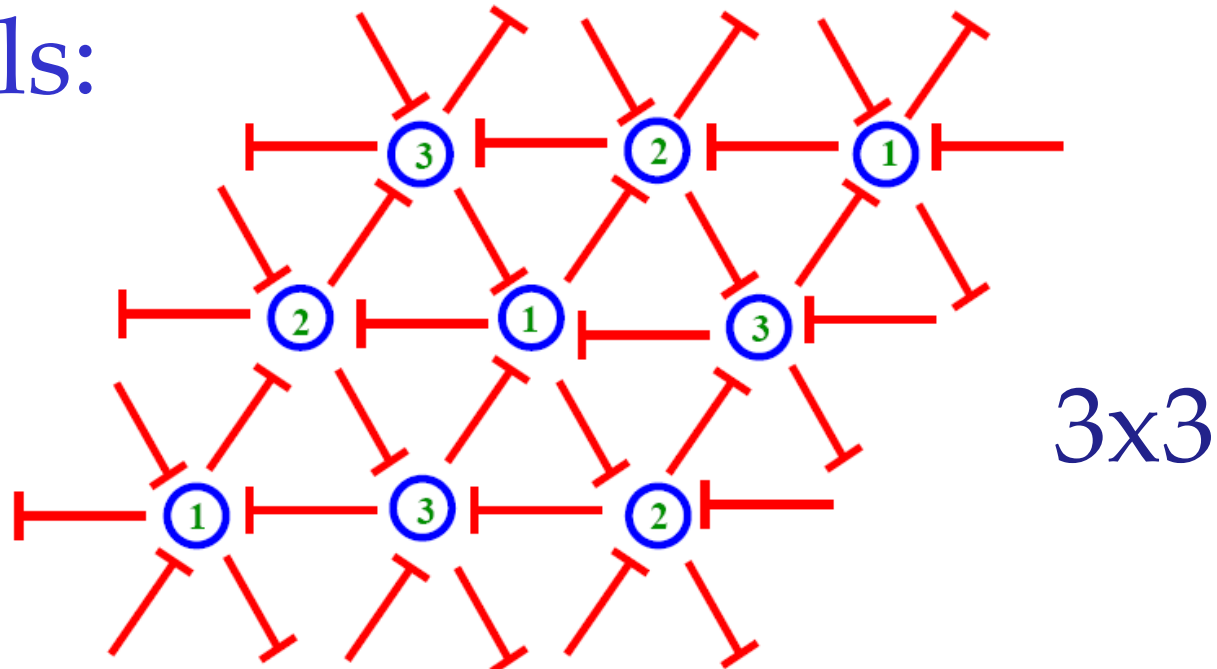
Coupled Repressilators: A Repressor-Lattice



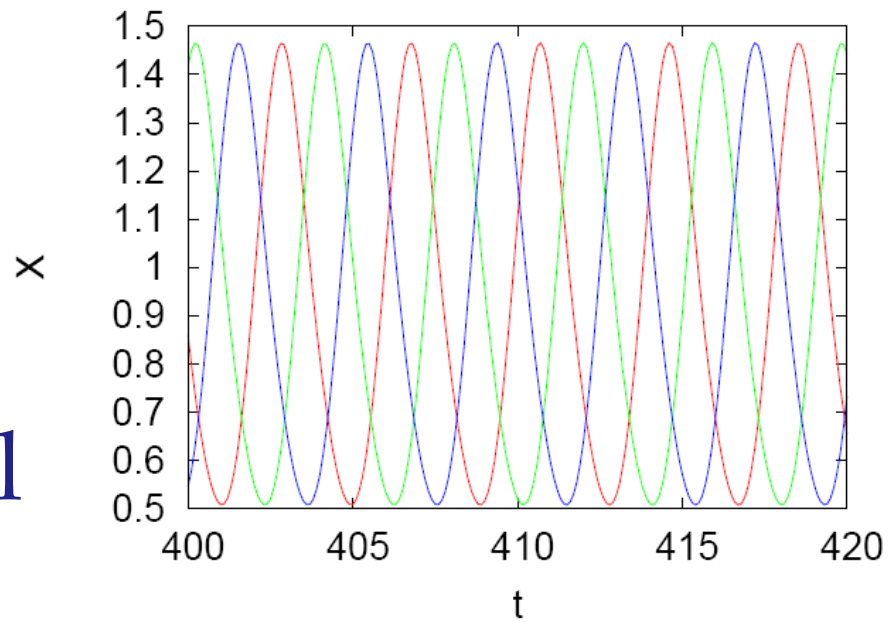
Repressing genes/proteins:
in a cell or between cells !

'Natural' phases: No frustration !

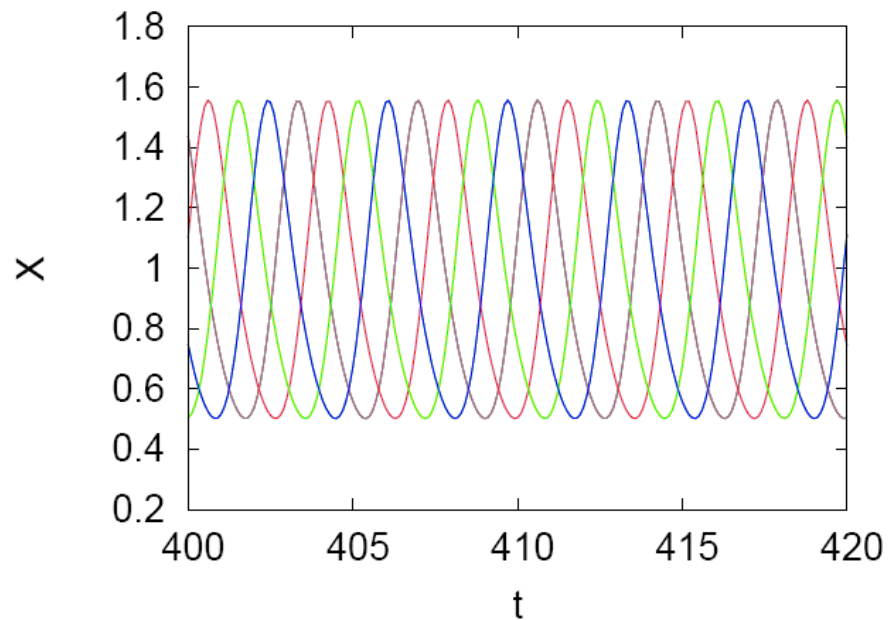
Unit cells:



Dynamical solutions



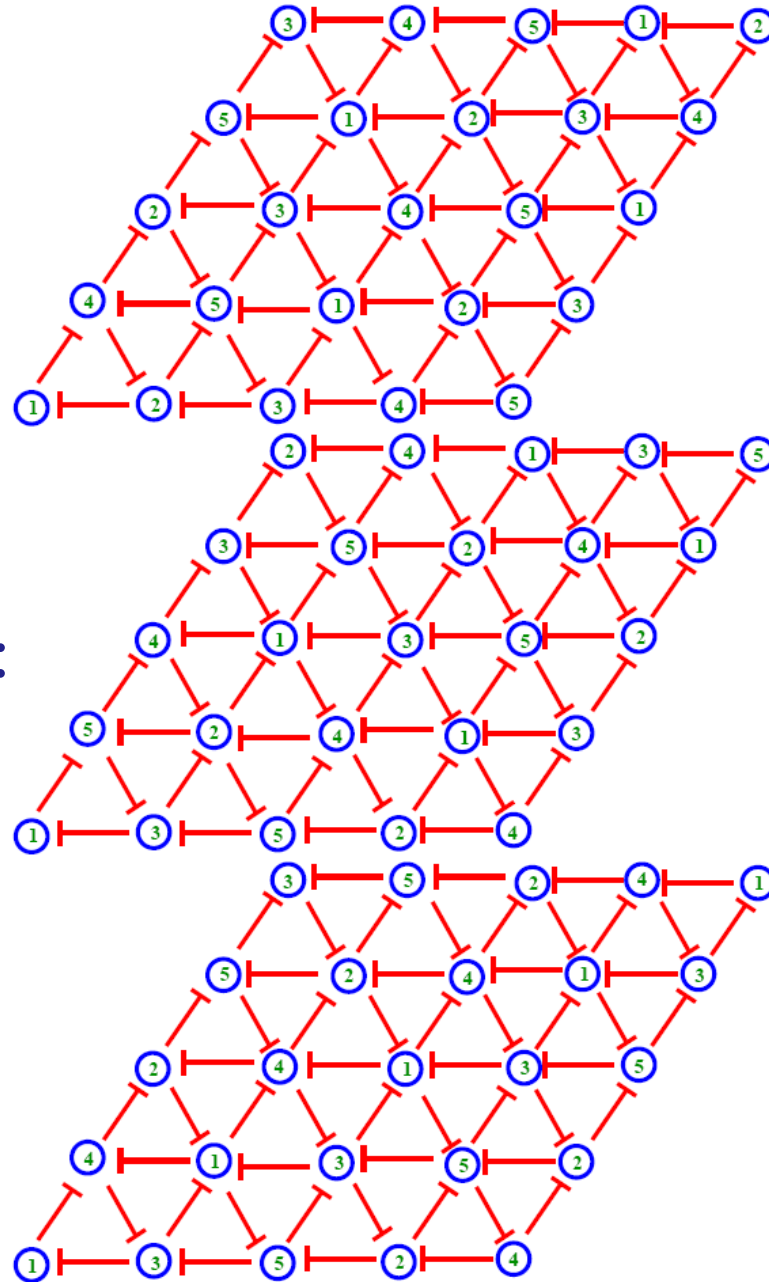
3×3



4×4

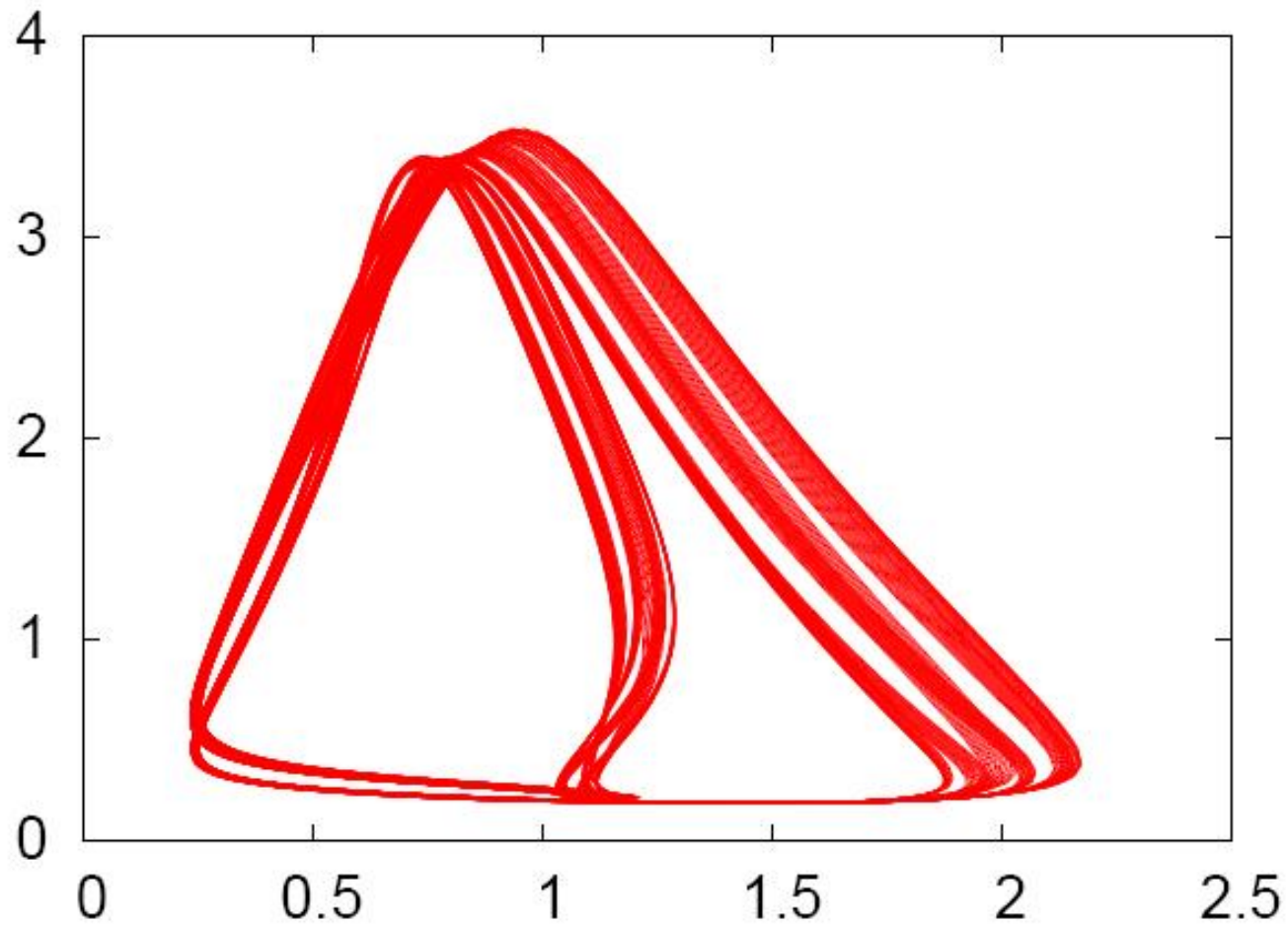
5x5

Solutions from
rotational symmetry:
Group theory !

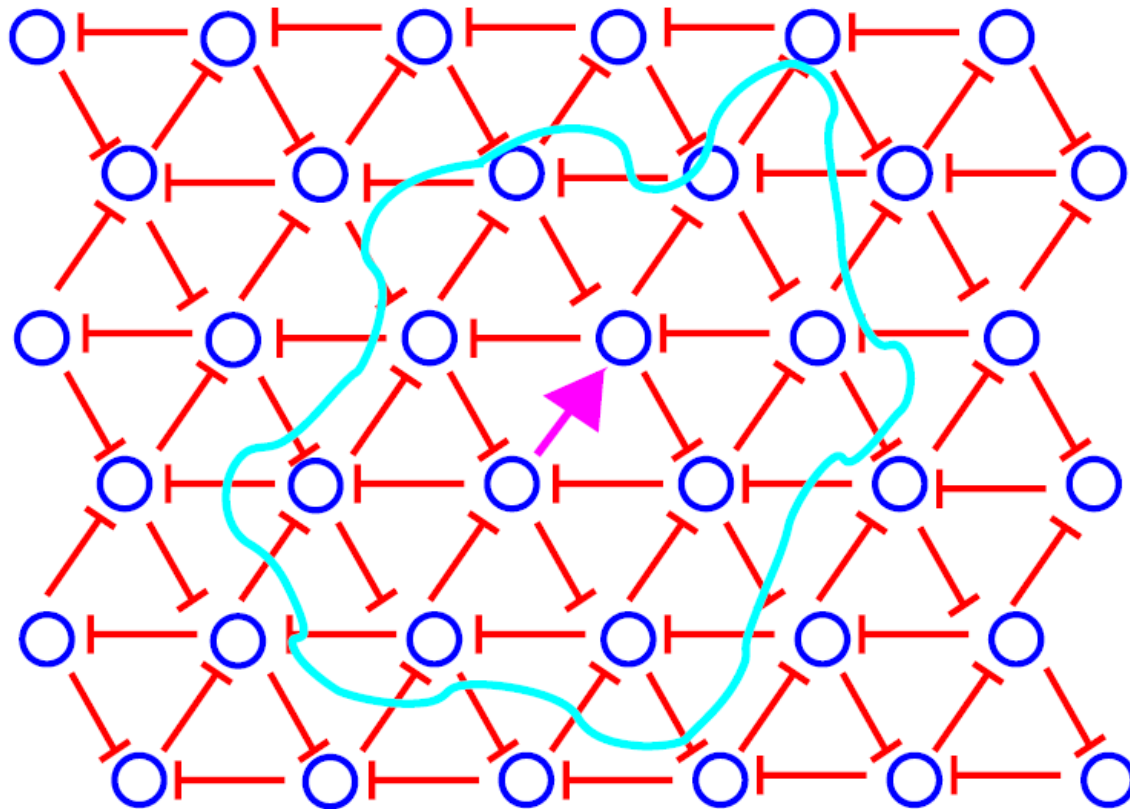


Dynamical frustrations: Chaotic solutions

5x5



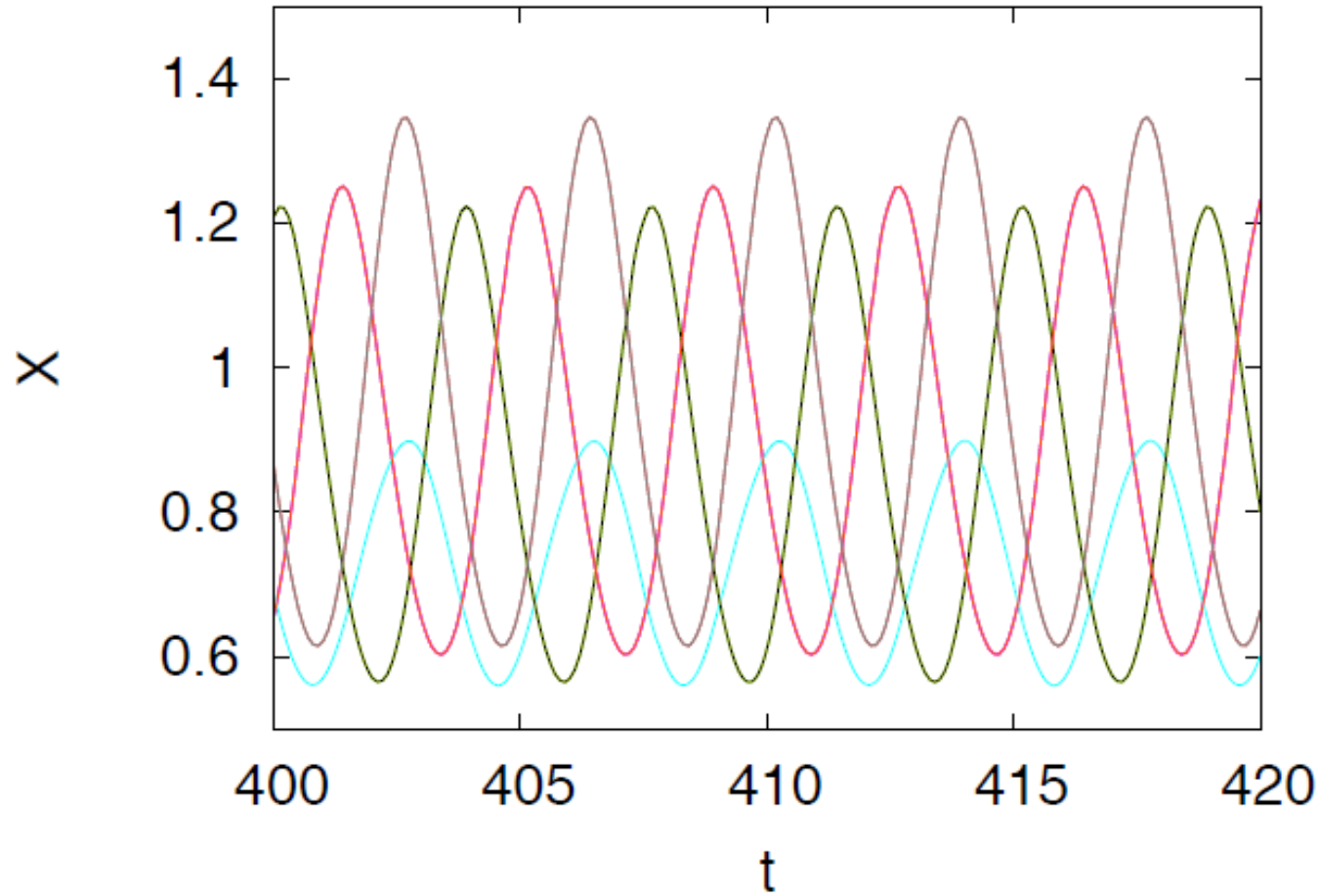
Non-balanced repressor-lattice



One activator

frustration !

Frustrated solutions:



3 x 3 lattice

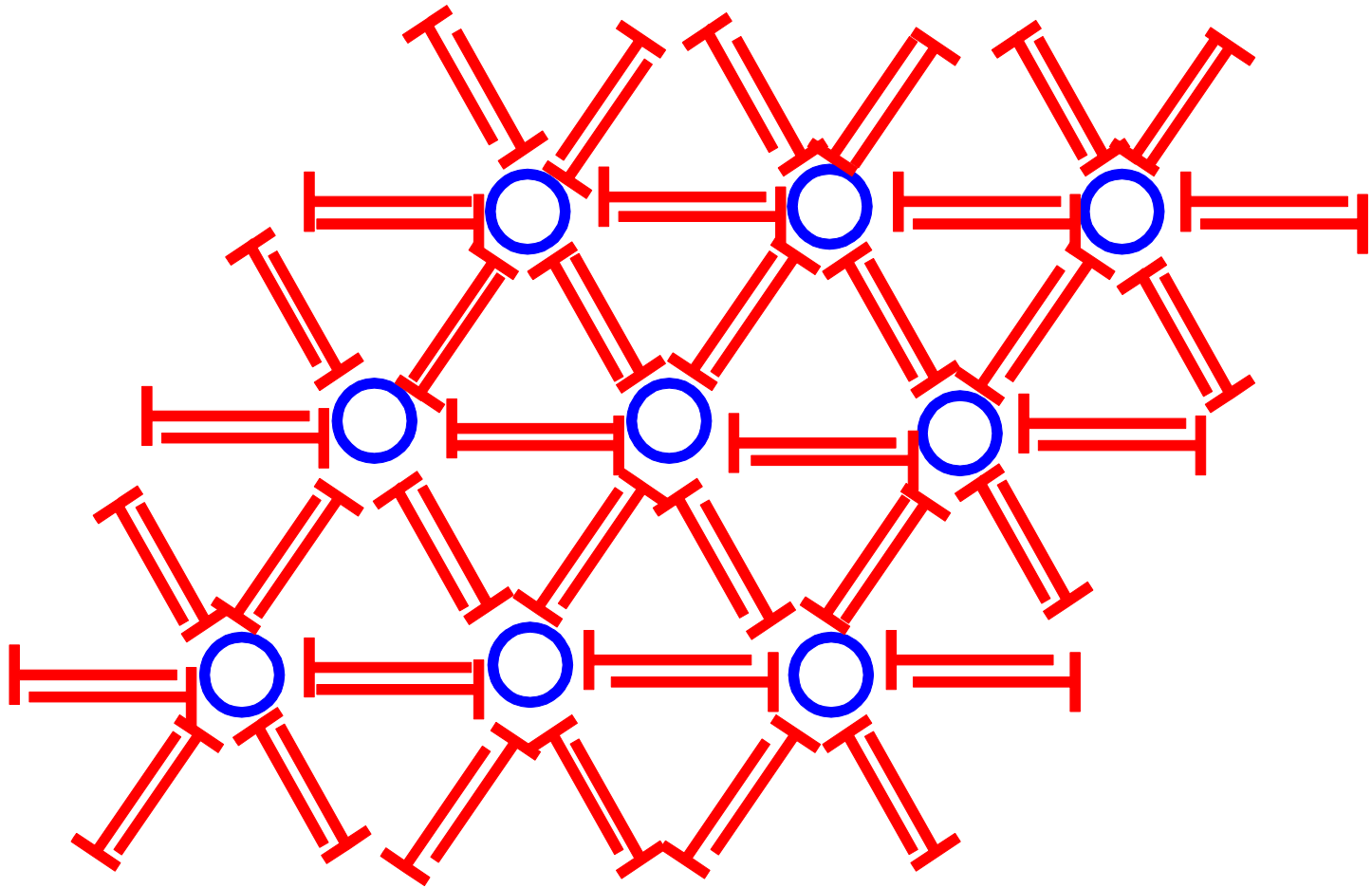
Phases are fixed !

Cell-to-cell communication:

- Cell touches: gap junctions → Directed (or bidirected?) interactions ! **Oscillations !**
- Cells do not touch: Send out signalling molecules → bi-directed interactions:
‘Local’ switches → High or low state !

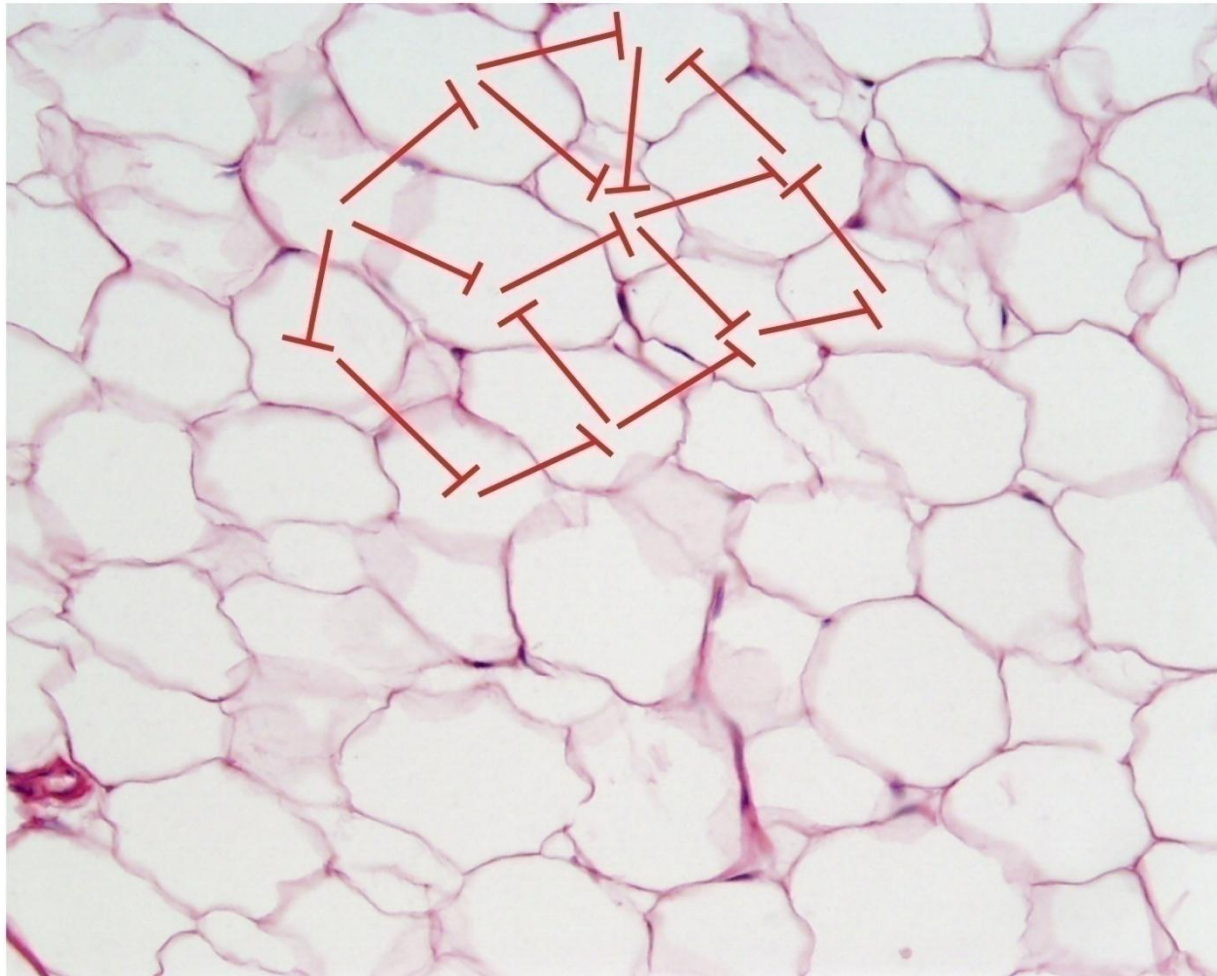
Relevant for ordered tissues: fat tissue, onion skin, human skin, etc.

Bi-directional interactions



No oscillations: hi or low states. Mutations !

Cell-to-cell communicatios in space:
Fatty tissues: organized on a “lattice”



Negative feed-back loops

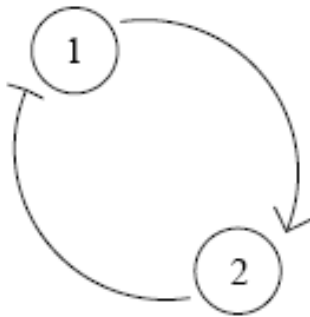
- Guess the feed-back loop from an experimental time series: Algorithm
- Derive symbolic dynamics
- From fixed points to Hopf bifurcations

with Simone Pigolotti and Sandeep Krishna

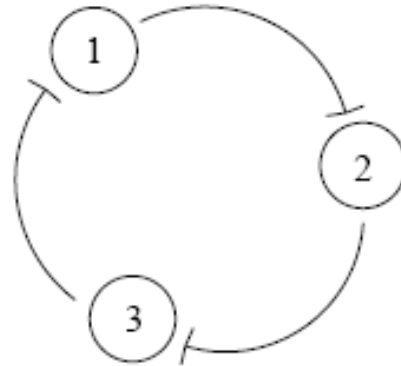
Proc. Nat. Acad. Sci. (2007)

Negative Feed-Back Loops:

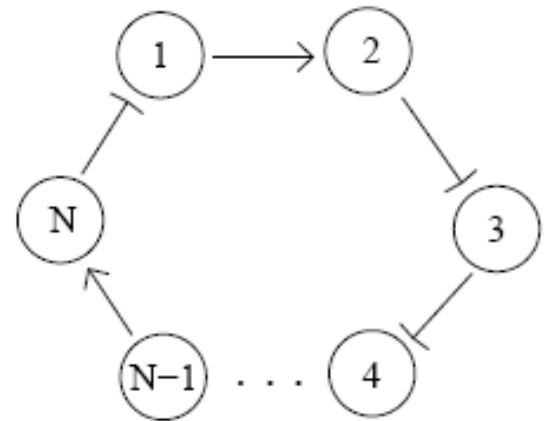
(a)



(b)



(c)



- **Always odd number of repressor links**
- **A 'closed' loop: No cross-links**
- **Node: concentration, expression level, etc**

Determined by the following equation:

$$\frac{dx_i}{dt} = g_i^{(A,R)}(x_i, x_{i-1})$$

Assume $g_i^{(A;R)}$ are monotonic

Example:
$$\frac{dx_i}{dt} = c - \gamma x_i + \alpha \frac{1}{1 + (x_{i-1}/K)^h}$$

- c : basal production
- γ : degradation \rightarrow linear
- α : production rate: activation \leftrightarrow repression

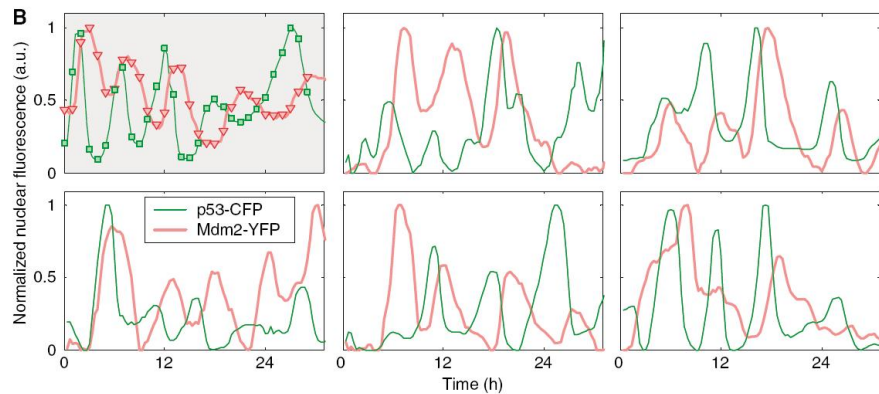
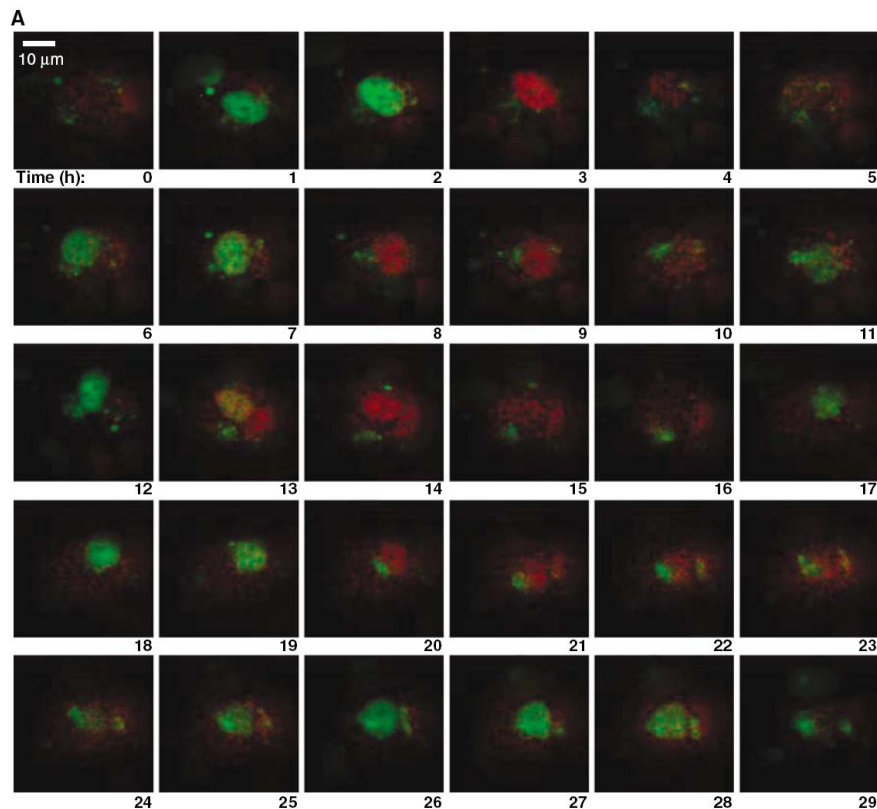
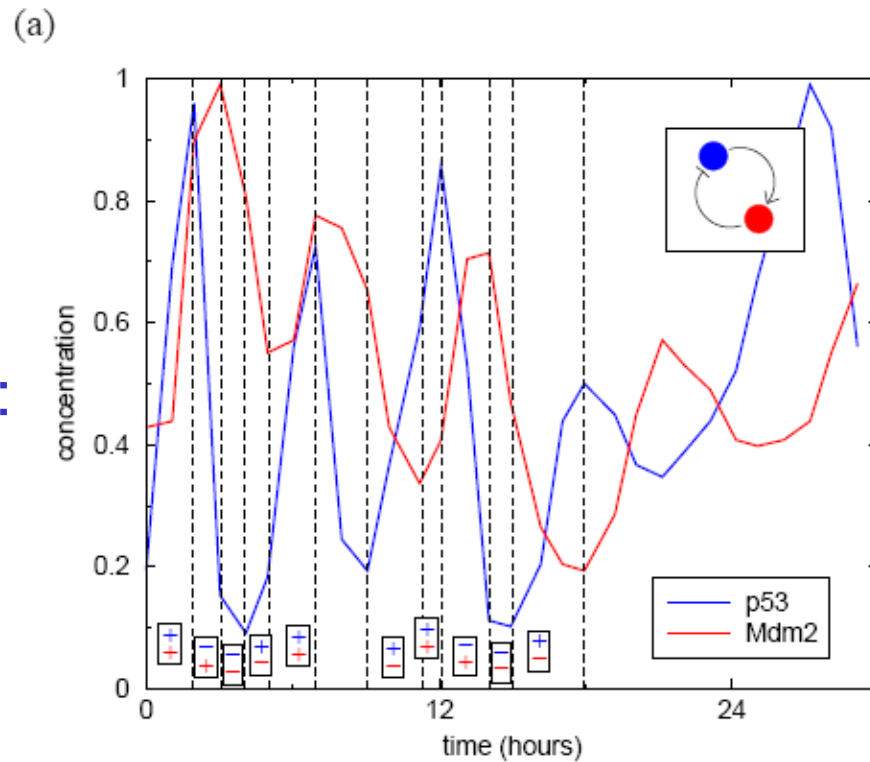


Figure 1 Prolonged oscillations in the nuclear levels of fluorescently tagged p53 and Mdm2 in individual MCF7, U280, cells following gamma irradiation. **(A)** Time-lapse fluorescence images of one cell over 29 h after 5 Gy of gamma irradiation. Nuclear p53-CFP and Mdm2-YFP are imaged in green and red, respectively. Time is indicated in hours. **(B)** Normalized nuclear fluorescence levels of p53-CFP (green) and Mdm2-YFP (red) following gamma irradiation. Top left: the cell shown in panel A. Other panels: five cells from one field of view, after exposure to 2.5 Gy gamma irradiation.

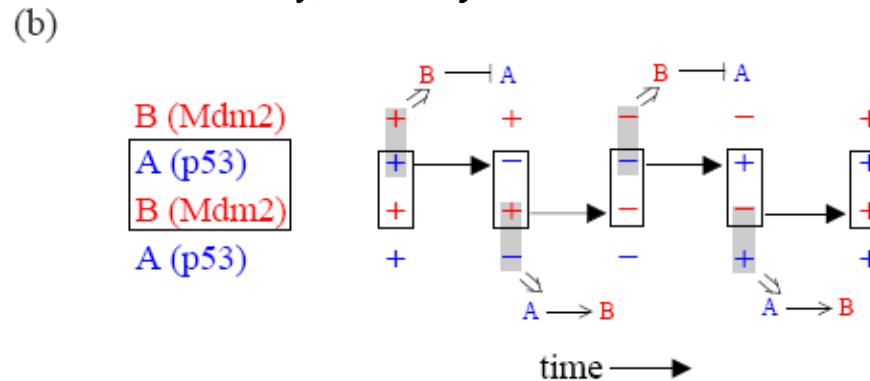
Often time series
are very noisy !
→ Then what ?

Experimental time series:
(transient is OK !)



N. Geva-Zatorsky, *Mol. Sys. Biol.* 2:2006.0033 (2006)

Symbolic Dynamics:




Stationary point:

$$g_i^{(A,R)}(x_i^*, x_{i-1}^*) = 0 \quad \Rightarrow \quad x_i^* = f_i^{(A,R)}(x_{i-1}^*)$$

Notice: $f_i^{(A,R)} \rightarrow$ same monotonicity as $g_i^{(A,R)}$
(when $g_i^{(A,R)}$ is decreasing in x_i)

Fixed point equation:

$$\begin{aligned} x_i^* &= f_i(x_{i-1}^*) = f_i(f_{i-1}(x_{i-2}^*)) = \dots = \\ &= f_i \circ f_{i-1} \circ f_{i-2} \circ \dots \circ f_{i+1}(x_i^*) \equiv F_i(x_i^*) \end{aligned}$$

 I.e. measures how species 'i' interacts with itself through the loop

Using chain rule: $F'_i(x) = \prod_j f'_j(x_j)|_{x_i=x}$

Therefore: If even number of repressors \rightarrow
positive \rightarrow multiple fixed points (bistability)

If odd number of repressors \rightarrow
negative \rightarrow only one fixed point !

Eigenvalue equation: $\prod_{i=1}^N \left(\frac{\lambda}{h_i} + 1 \right) = F'(x^*)$

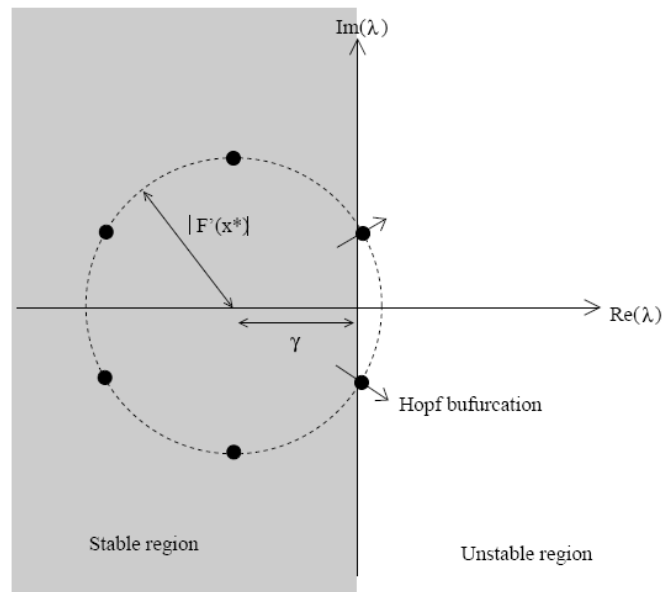
where $h_i = -\partial_x g_i(x_i, x_{i=1})|_{x^*}$ (degradation rates at fixed point)

$$\prod_{i=1}^N \left(\frac{\lambda}{h_i} + 1 \right) = F'(x^*)$$

Right hand side: **Negative**

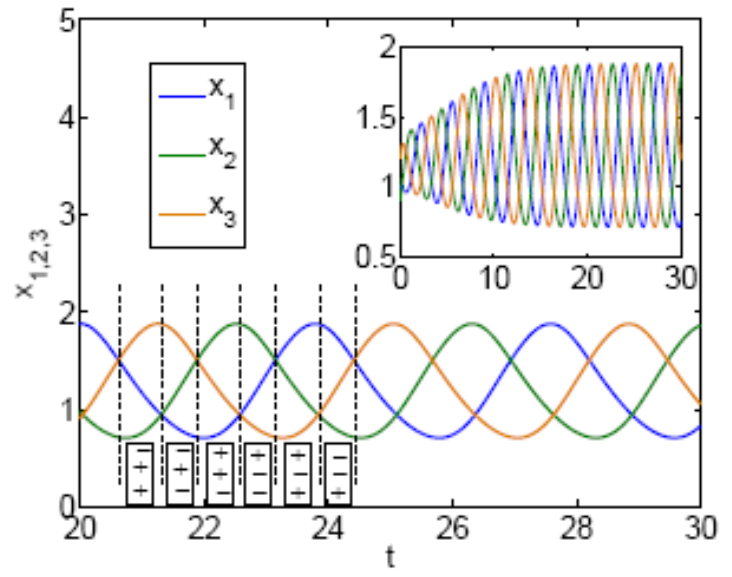
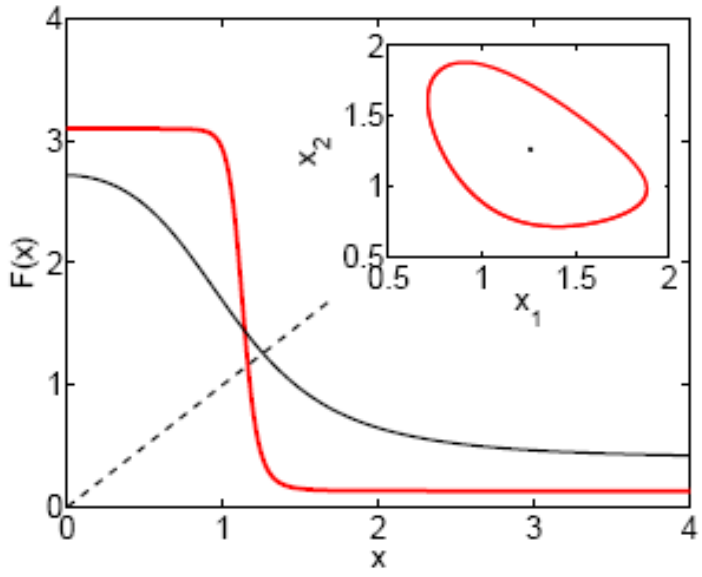
Left hand side: **Positive coefficients**

: Hopf bifurcation



Example:

$\alpha=3.0, c=0.1,$
 $K=1, \gamma=1, h=2$
 $h=4$



Three repressors:

$$\frac{dx_i}{dt} = c - \gamma x_i + \alpha \frac{1}{1 + (x_{i-1}/K_i)^h} \quad i = 1 \dots 3.$$

We denote by x^* the solution to the equation $\gamma x = c + \alpha/(1 + (x/K)^h)$. Then the characteristic polynomial is simply:

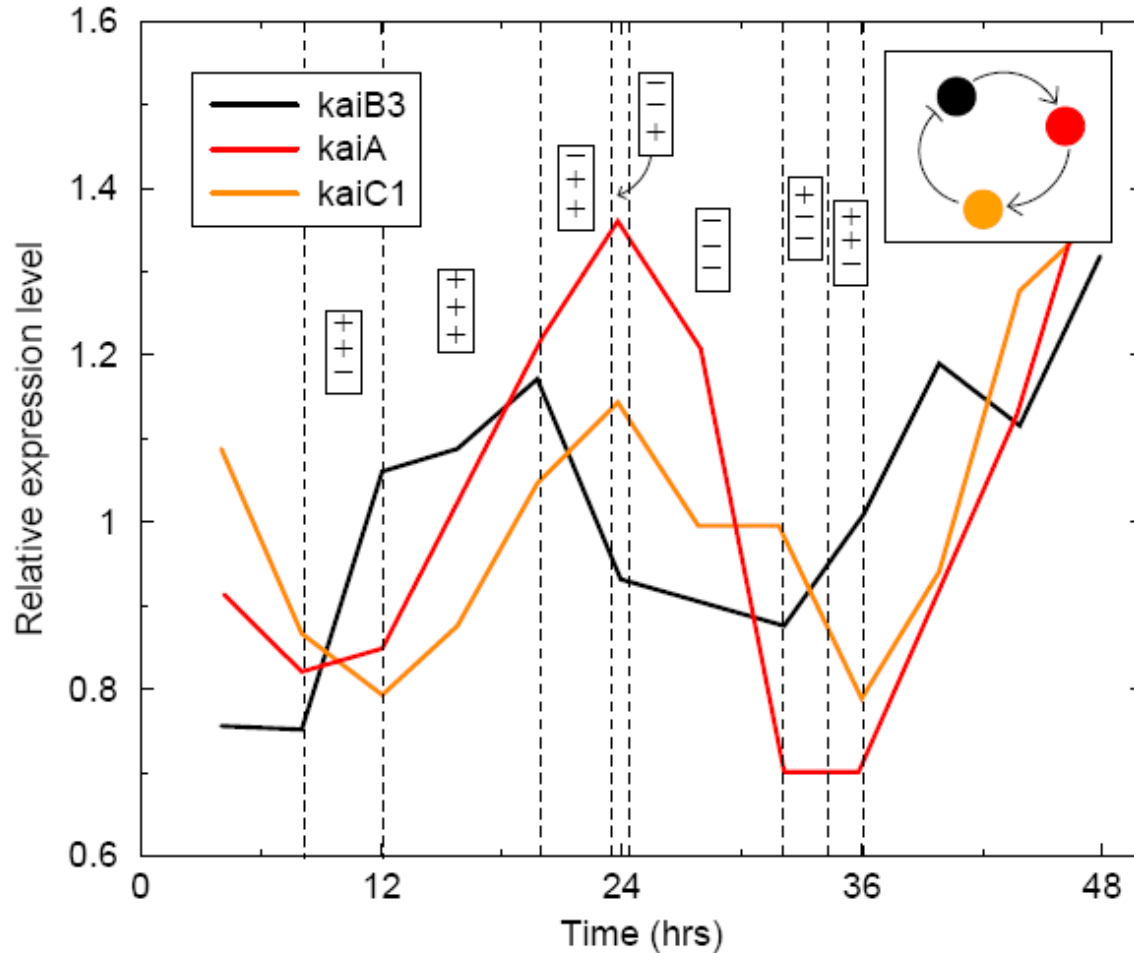
$$(\lambda + \gamma)^3 = - \left(\frac{\alpha}{1 + (x^*/K_i)^h} \right)^3$$

$$\Rightarrow \left(\frac{\lambda}{\gamma} + 1 \right)^3 = F'(x^*) \Rightarrow |F'(x^*)| \cos(\pi/3) < \gamma \quad \begin{cases} \text{OK: } h=2 \\ \text{Not OK: } h=4 \end{cases}$$

Stability condition

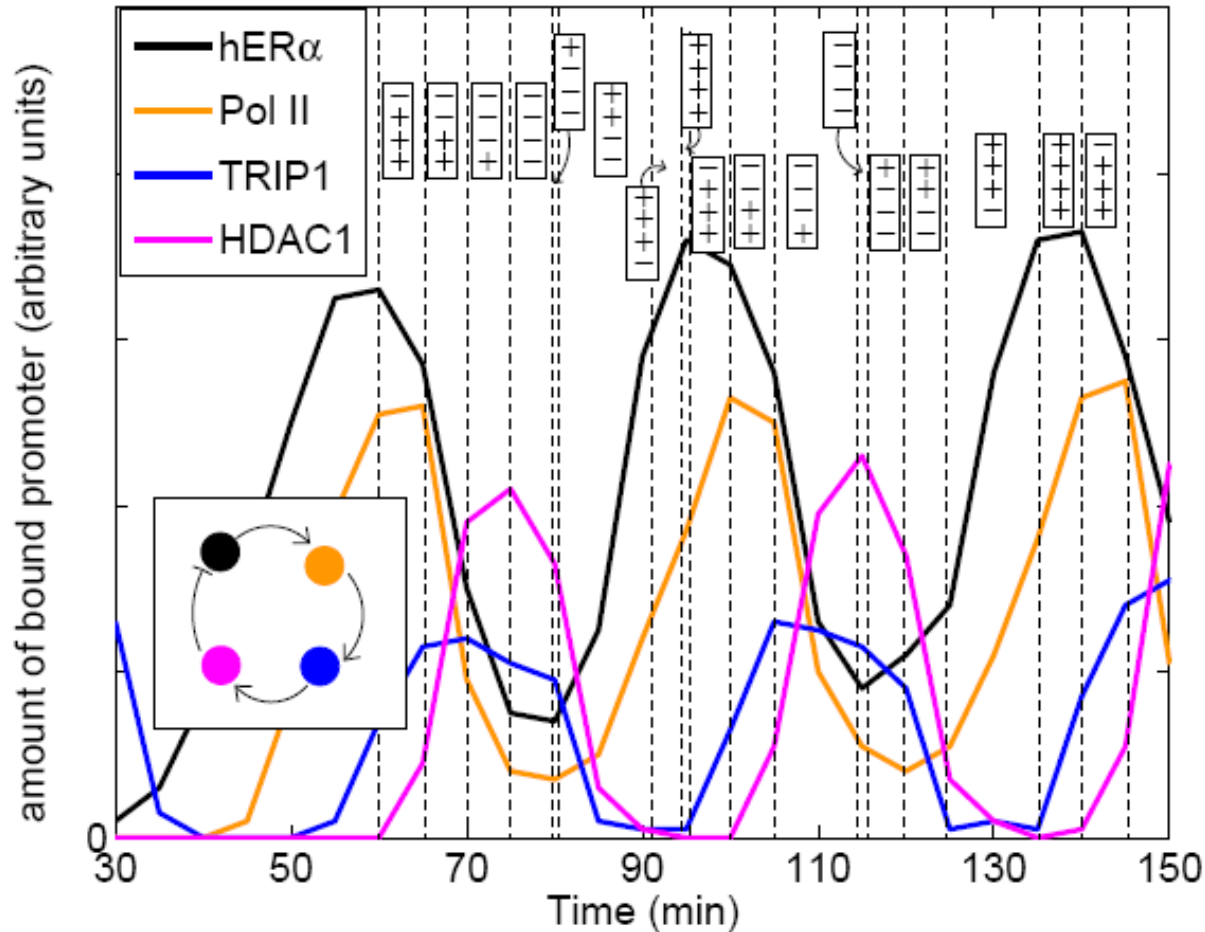
Circadian rhythms of kai genes in cyanobacteria

(a)



Binding of four proteins to pS2 promoter

(b)



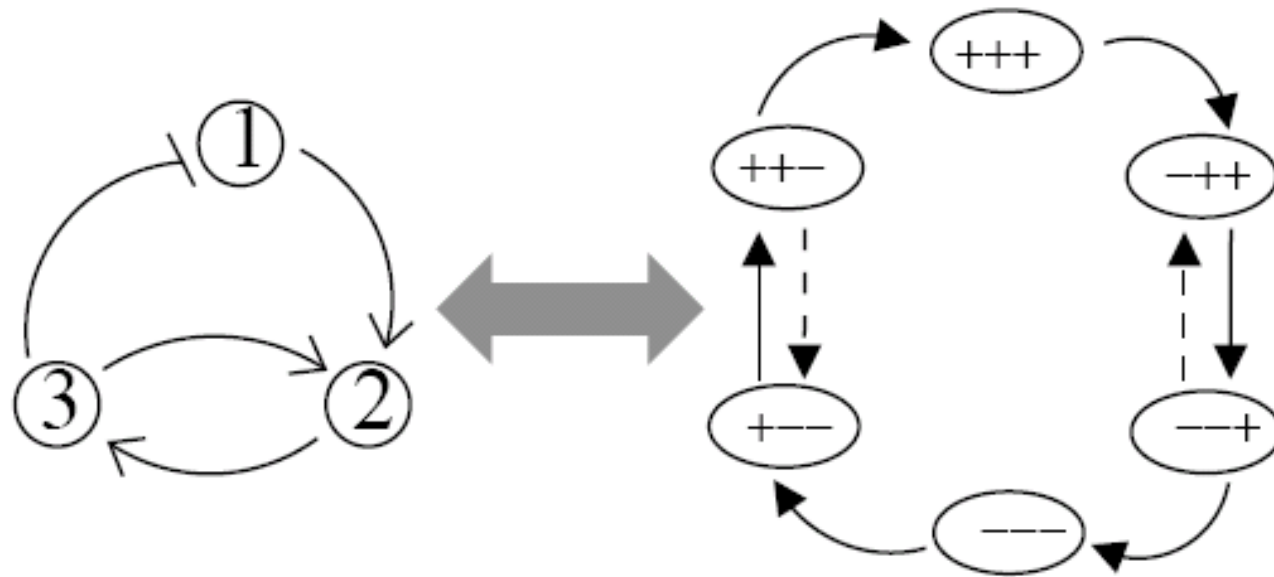
Adding estradiol:

ER: estradiol receptor, initiates transcription

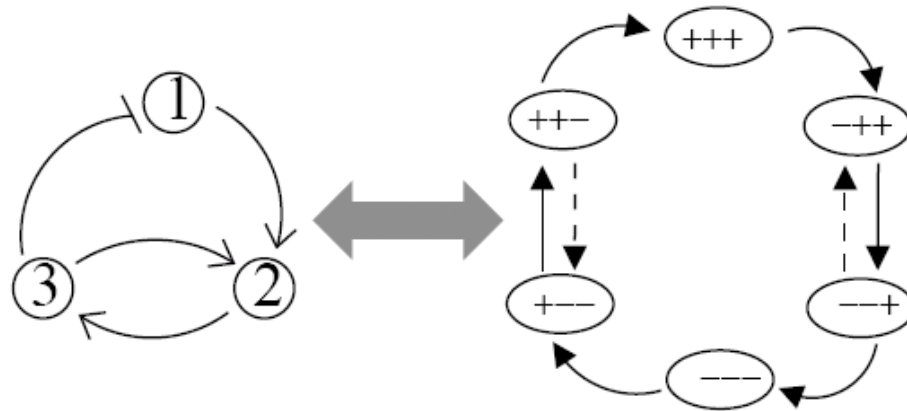
Pol: RNA polymerase

Now with possible cross-links !

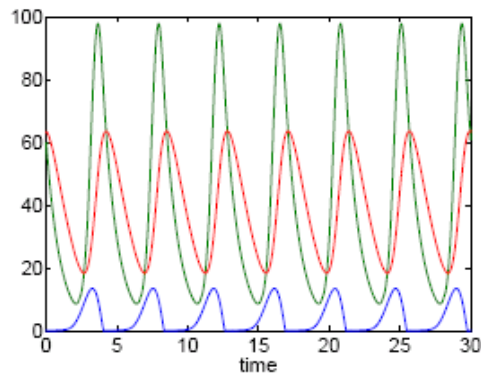
a)



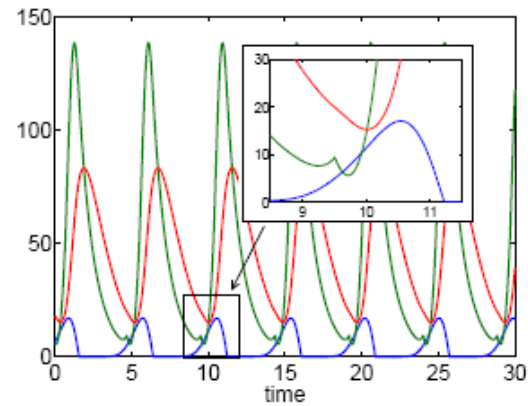
a)



b)



c)



‘Normal’ symbolic dynamics

Kick !

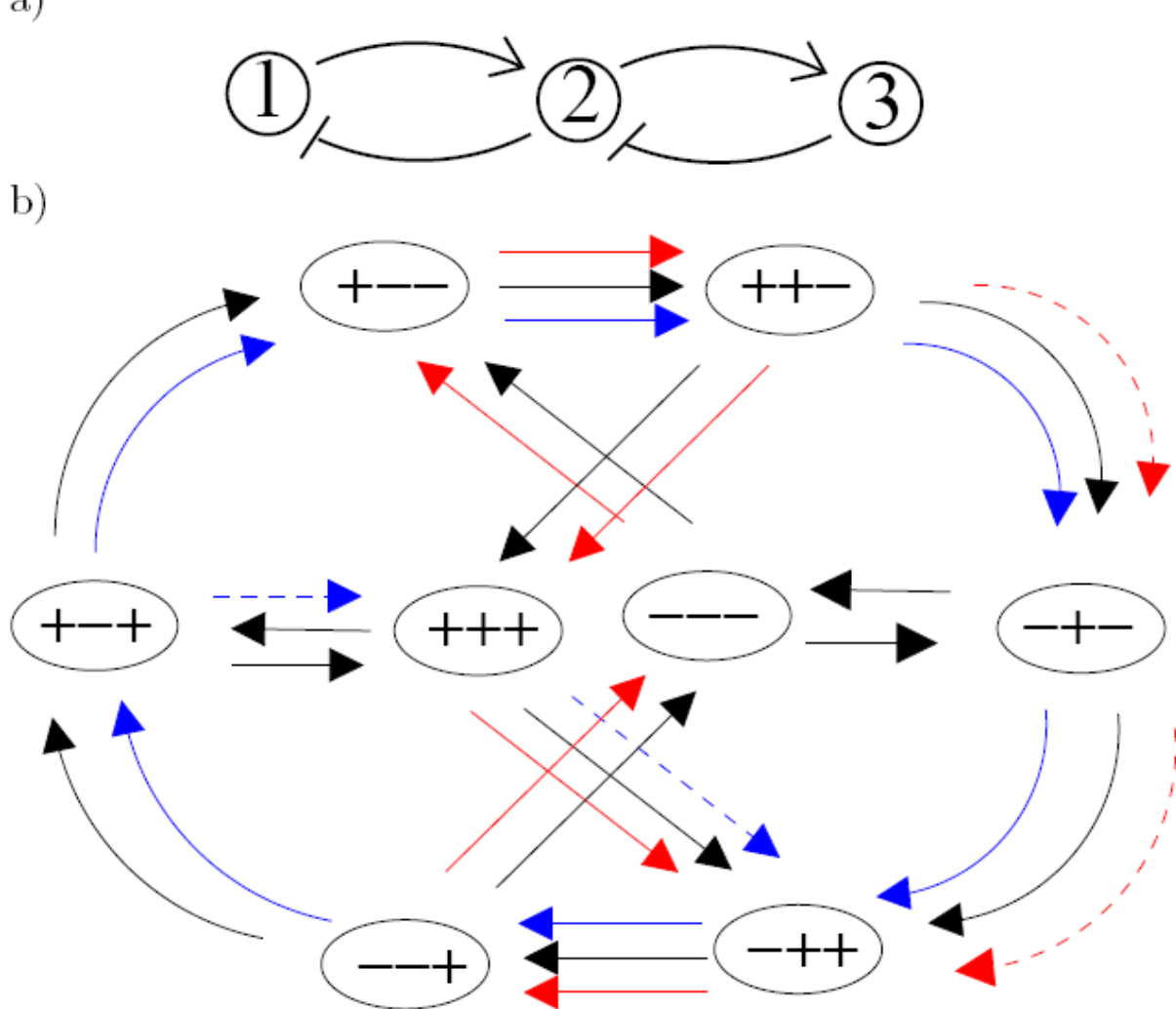


FIG. 2: Network of two coupled two-species oscillators, or a three trophic level ecosystem. (a) Structure of the network. (b) The transition network for this 3-node system. Black arrows indicate all the allowed transitions. Blue arrows are the transitions actually observed in the HP system and red arrows are the transitions observed in the BHS model (see text). In both cases, dashed arrows indicate “kicks”, i.e., transitions

Two possible dynamical systems for system

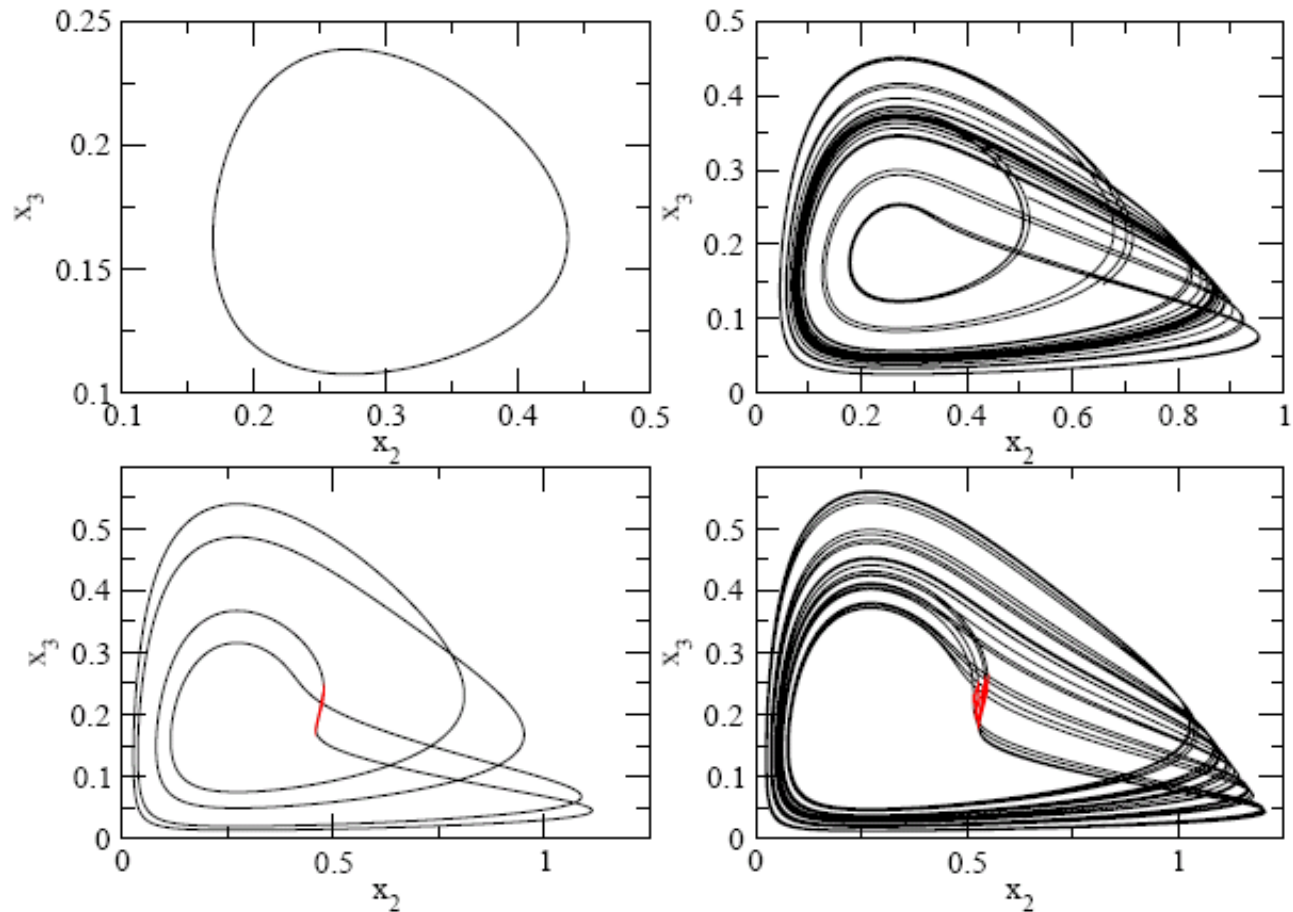
Hastings-Powell

$$\begin{aligned}\dot{x}_1 &= rx_1(1 - kx_1) - \alpha_1 \frac{x_1x_2}{1 + b_1x_1} \\ \dot{x}_2 &= -d_1x_2 + \alpha_1 \frac{x_1x_2}{1 + b_1x_1} - \alpha_2 \frac{x_2x_3}{1 + b_2x_2} \\ \dot{x}_3 &= -d_2x_3 + \alpha_2 \frac{x_2x_3}{1 + b_2x_2}\end{aligned}$$

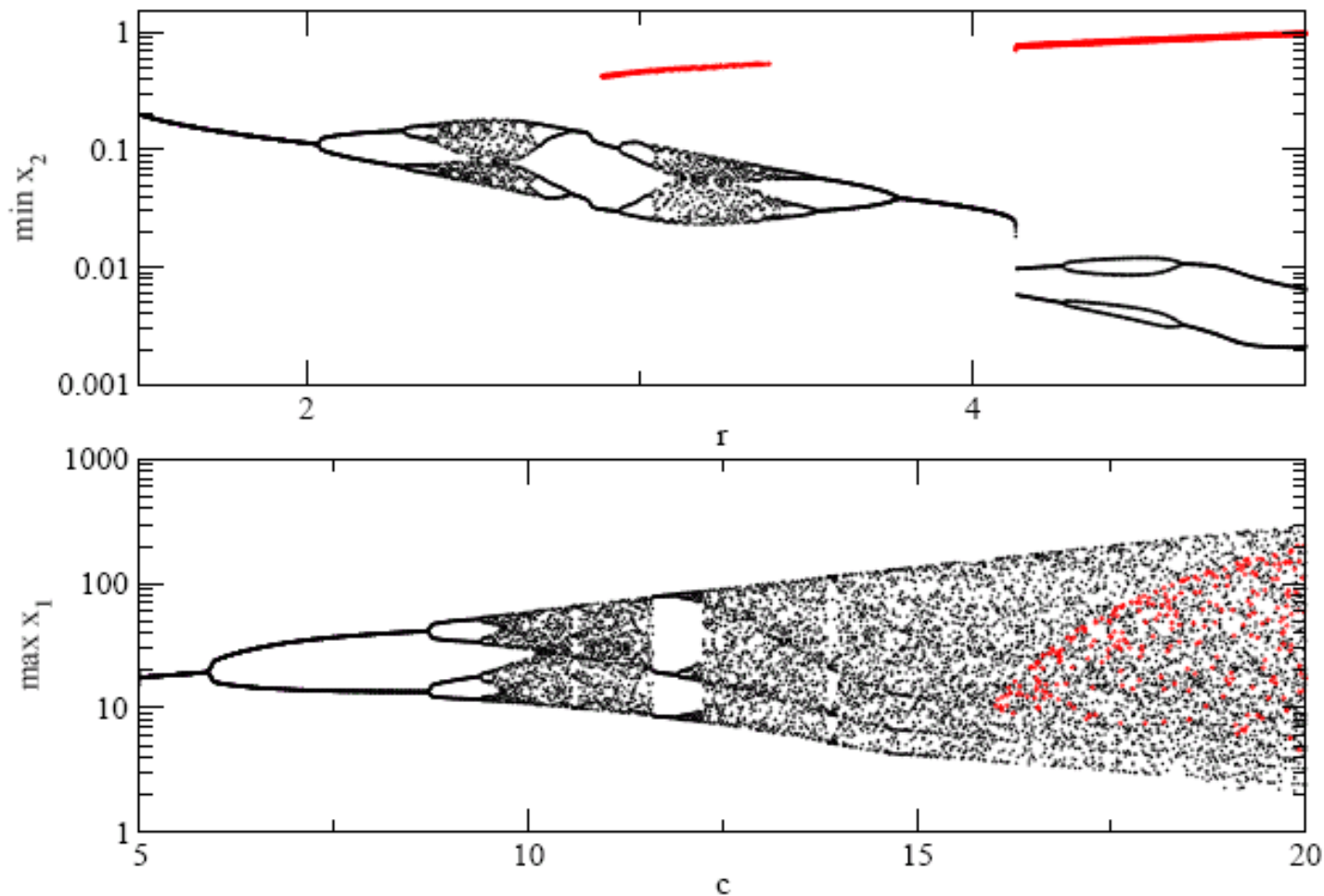
Blasius et al

$$\begin{aligned}\dot{x}_1 &= x_1 - \alpha_1 \frac{x_1x_2}{1 + kx_1} \\ \dot{x}_2 &= -dx_2 + \alpha_1 \frac{x_1x_2}{1 + kx_1} - \alpha_2x_2x_3 \\ \dot{x}_3 &= c(x_3^* - x_3) + \alpha_2x_2x_3\end{aligned}$$

Attractors of the dynamics of HP model

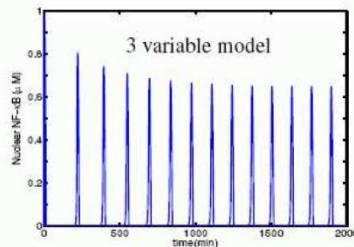
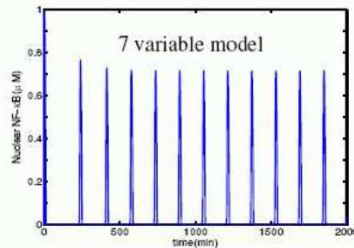
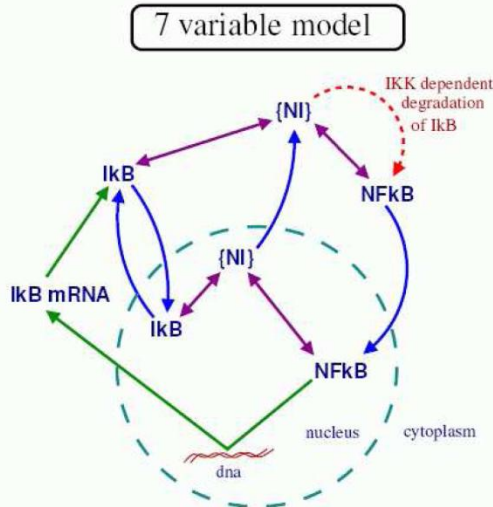


Bifurcation diagrams for HP and Blasius



Reduction of the NF- κ B System

Sandeep Krishna



$A = 0.007$
 $B = 954.5$
 $C = 0.035$
 $\epsilon = 2 \times 10^{-5}$
 $\delta = 0.029$

4 variable model

$$(NI) = (N_{tot} - N_n) \frac{I}{K_I + I}$$

$$N = (N_{tot} - N_n) \frac{K_I}{K_I + I} \quad \text{Equilibrium of complexes}$$

$$(NI)_n = I_n^{tot} \frac{N_n}{K_N + N_n}$$

$$I_n = I_n^{tot} \frac{K_N}{K_N + N_n}$$

$$\frac{dN_n}{dt} = k_{Nin} K_I \frac{(N_{tot} - N_n)}{K_I + I} - k_{NIout} \frac{I_n^{tot} N_n}{K_N + N_n}$$

$$\frac{dI_m}{dt} = k_t N_n^2 - \gamma_m I_m \quad \text{Small terms deleted}$$

$$\frac{dI}{dt} = k_{tl} I_m - \alpha \frac{(N_{tot} - N_n) I}{K_I + I} - k_{Iin} I + k_{Iout} K_N \frac{I_n^{tot}}{K_N + N_n}$$

$$\frac{dI_n^{tot}}{dt} = k_{Iin} I - k_{Iout} K_N \frac{I_n^{tot}}{K_N + N_n} - k_{NIout} \frac{I_n^{tot} N_n}{K_N + N_n}$$

$$t \rightarrow (1/\gamma_m)t$$

$$N_n \rightarrow N_{tot} N_n$$

$$I_m \rightarrow (k_t N_{tot}^2 / \gamma_m) I_m$$

$$I \rightarrow (k_t k_{tl} N_{tot}^2 / \gamma_m^2) I$$

$I_n^{tot} N_n / (K_N + N_n)$ is significant only when $N_n \gg K_N$. At this point I_n^{tot} reaches its minimum $\Rightarrow I_n^{tot} \approx \frac{k_{Iin}}{k_{NIout}} I$ ($dI_n^{tot}/dt = 0$ and $N_n \gg K_N$)

$$\frac{dN_n}{dt} = A \frac{(1 - N_n)}{\epsilon + I} - B \frac{IN_n}{\delta + N_n}$$

$$\frac{dI_m}{dt} = N_n^2 - I_m$$

$$\frac{dI}{dt} = I_m - C \frac{(1 - N_n) I}{\epsilon + I}$$

3 variable model

where

$$A = \frac{k_{Nin} K_I \gamma_m}{k_t k_{tl} N_{tot}^2}; \quad B = \frac{k_{Iin} k_t k_{tl} N_{tot}}{\gamma_m^3}; \quad C = \frac{\alpha \gamma_m}{k_t k_{tl} N_{tot}}$$

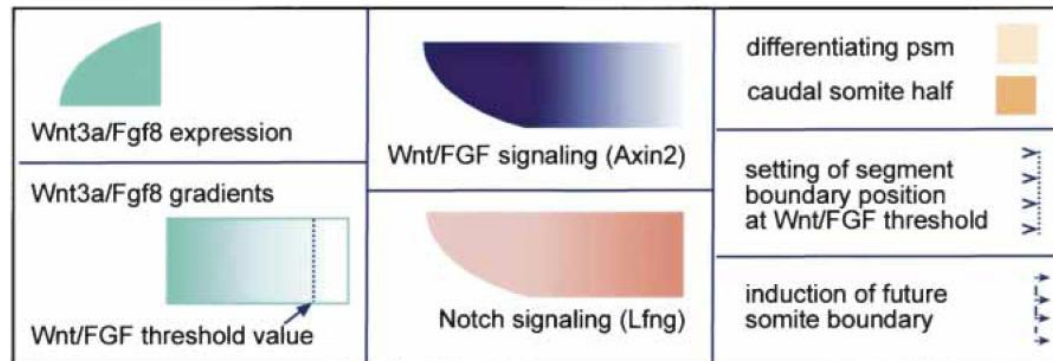
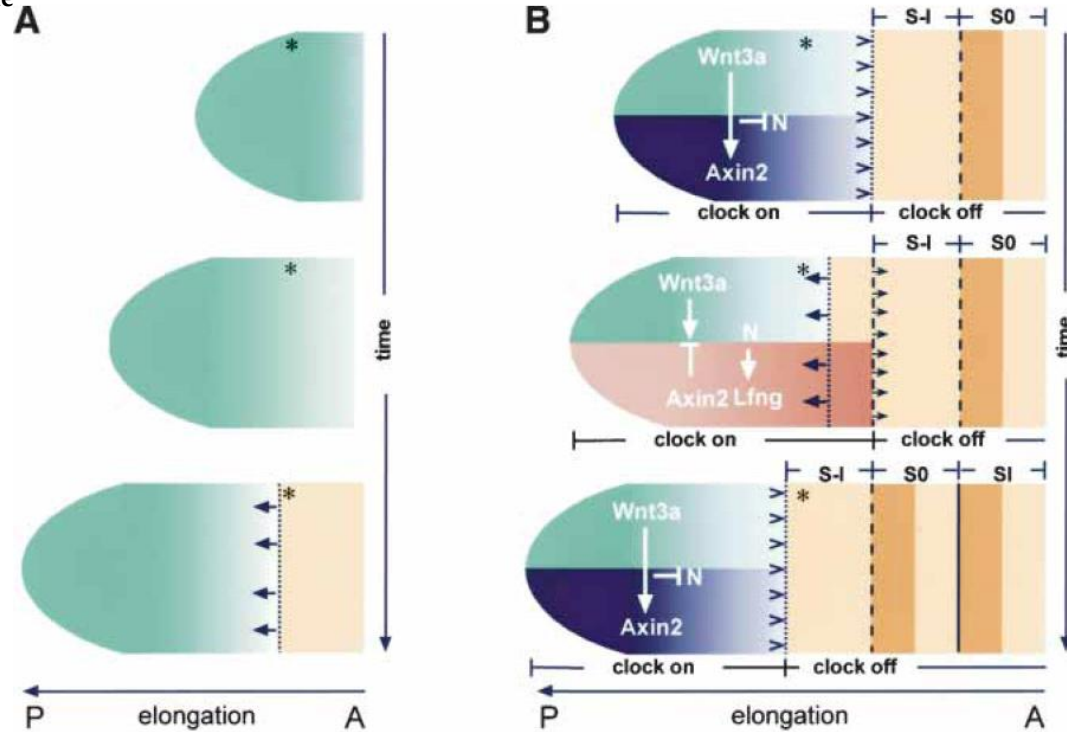
$$\epsilon = \frac{K_I \gamma_m^2}{k_t k_{tl} N_{tot}^2}; \quad \delta = K_N / N_{tot}$$

Two oscillators: Wnt and Notch: out of phase

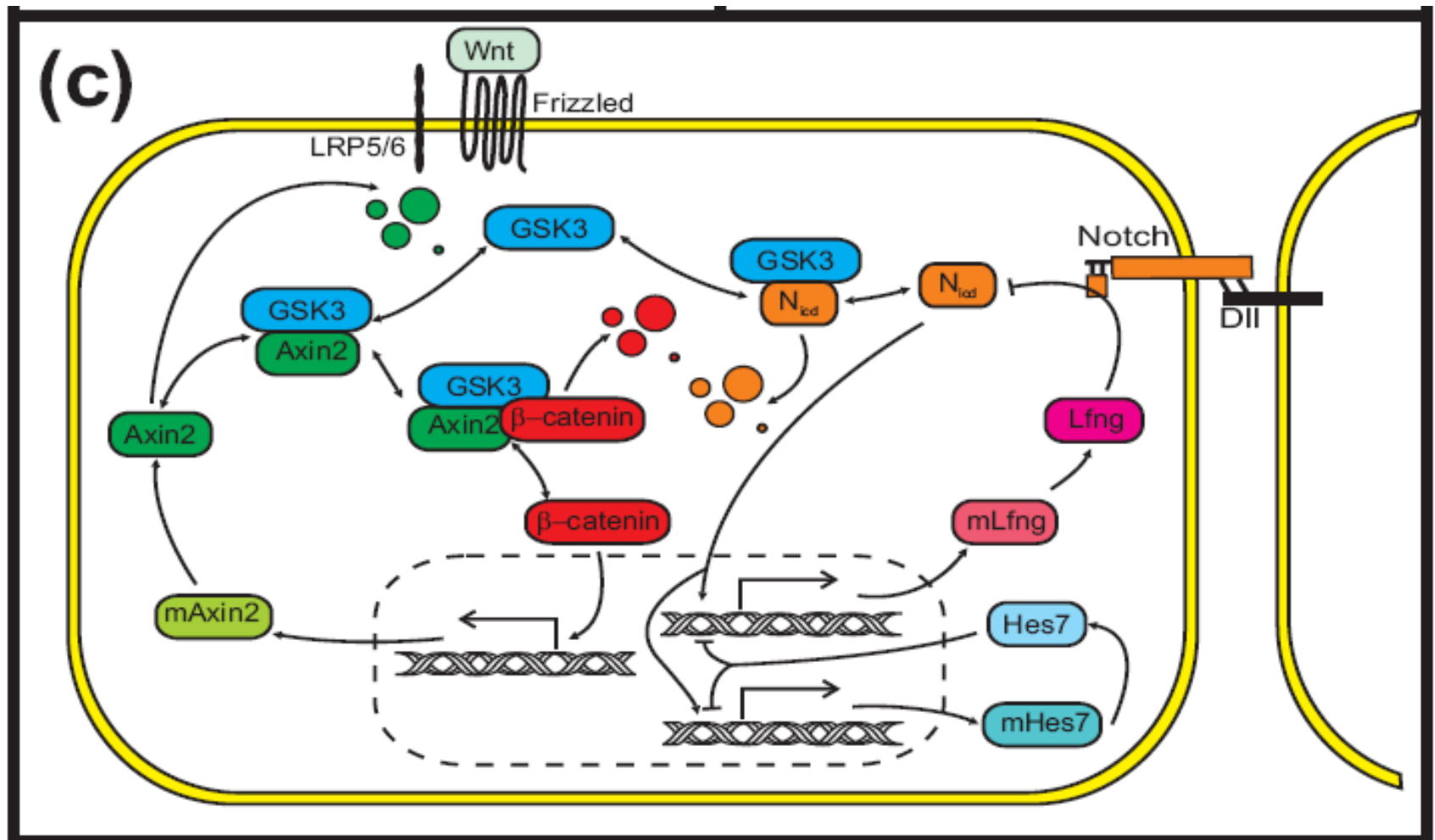
Aulehla and Herrmann

Goldbeter, Pourquie

Wnt gradient and Clock are coupled



The full systems with Wnt and Noctch and cross-talk



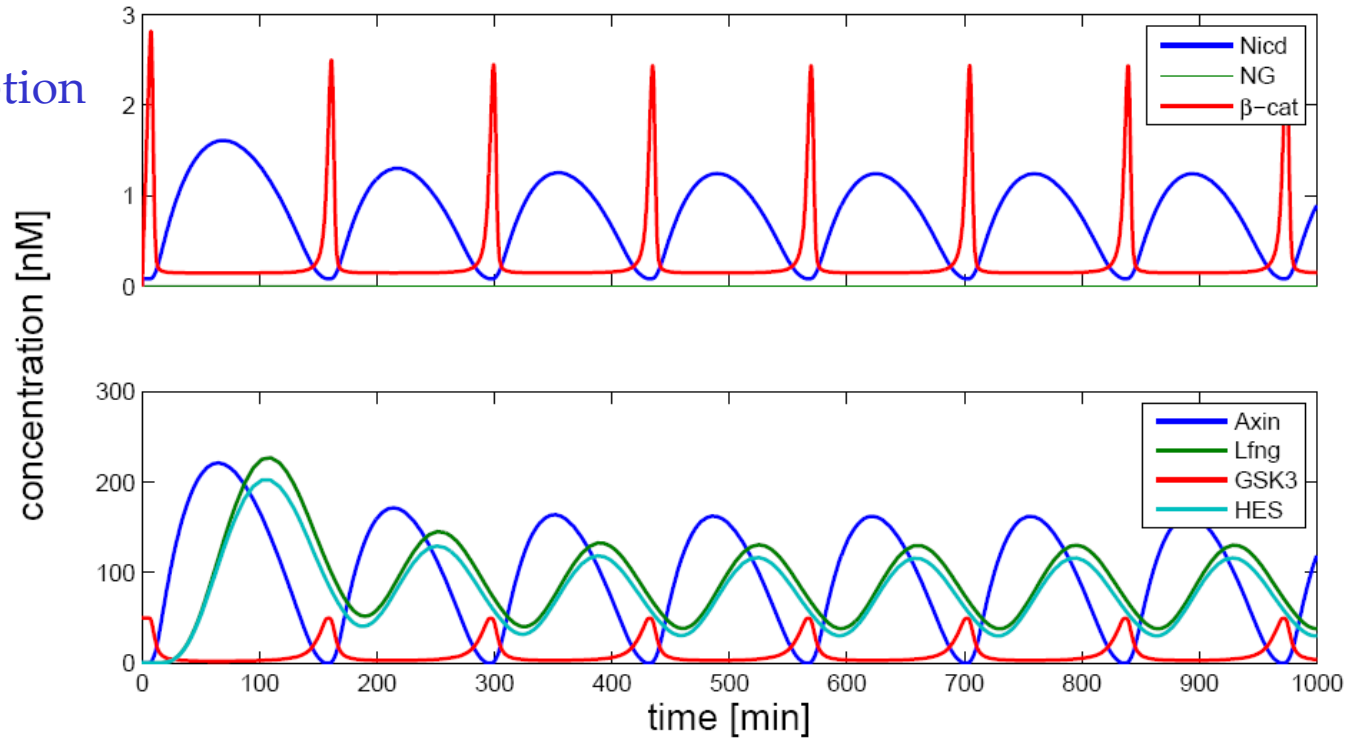
Equations of the Wnt-Notch network

Wnt	{	$\frac{dC}{dt} = c_{fC}B[GA] - c_{bC}C - \alpha_1C \quad (1)$
		$\frac{dG}{dt} = -c_{f[GA]}AG + c_{b[GA]}[GA] - c_{f[GN]}GN + c_{b[GN]}[GN] + \alpha_2[GN] \quad (2)$
		$\frac{dB}{dt} = S - c_{fC}B[GA] + c_{bC}C - \frac{B}{\tau_B} \quad (3)$
		$\frac{dA}{dt} = -c_{f[GA]}AG + c_{b[GA]}[GA] + c_{tlA}A_m - c_A \frac{A}{k_A + A} \quad (4)$
		$\frac{dA_m}{dt} = c_{tsA}B^h - \frac{A_m}{\tau_{Am}} \quad (5)$
		$\frac{d[GA]}{dt} = c_{f[GA]}AG - c_{b[GA]}[GA] - c_{fC}B[GA] + c_{bC}C + \alpha_1C \quad (6)$
		$\frac{dN}{dt} = c_s \frac{k_s}{k_s + L} - \frac{N}{\tau_N} - c_{f[GN]}GN + c_{b[GN]}[GN] \quad (7)$
Notch	{	$\frac{dH_m}{dt} = c_{tsH} \left(\frac{N^{h_{NH}}}{k_{NH} + N^{h_{NH}}} \cdot \frac{k_{HH}}{k_{HH} + H^{h_{HH}}} \right) - \frac{H}{\tau_{Hm}} \quad (8)$
		$\frac{dH}{dt} = c_{tlH}H_m - \frac{H}{\tau_H} \quad (9)$
		$\frac{dL_m}{dt} = c_{tsL} \left(\frac{N^{h_{NL}}}{k_{NL} + N^{h_{NL}}} \cdot \frac{k_{HL}}{k_{HL} + H^{h_{HL}}} \right) - \frac{L_m}{\tau_{Lm}} \quad (10)$
		$\frac{dL}{dt} = c_{tlL}L_m - \frac{L}{\tau_L} \quad (11)$
(Cross talk)	{	$\frac{d[NG]}{dt} = c_{f[NG]}NG - c_{b[NG]}[NG] - \alpha_2[NG] \quad (12)$

A total of approx. 27 constants – Some are poorly defined

The full Wnt-Notch system

Transcription factors



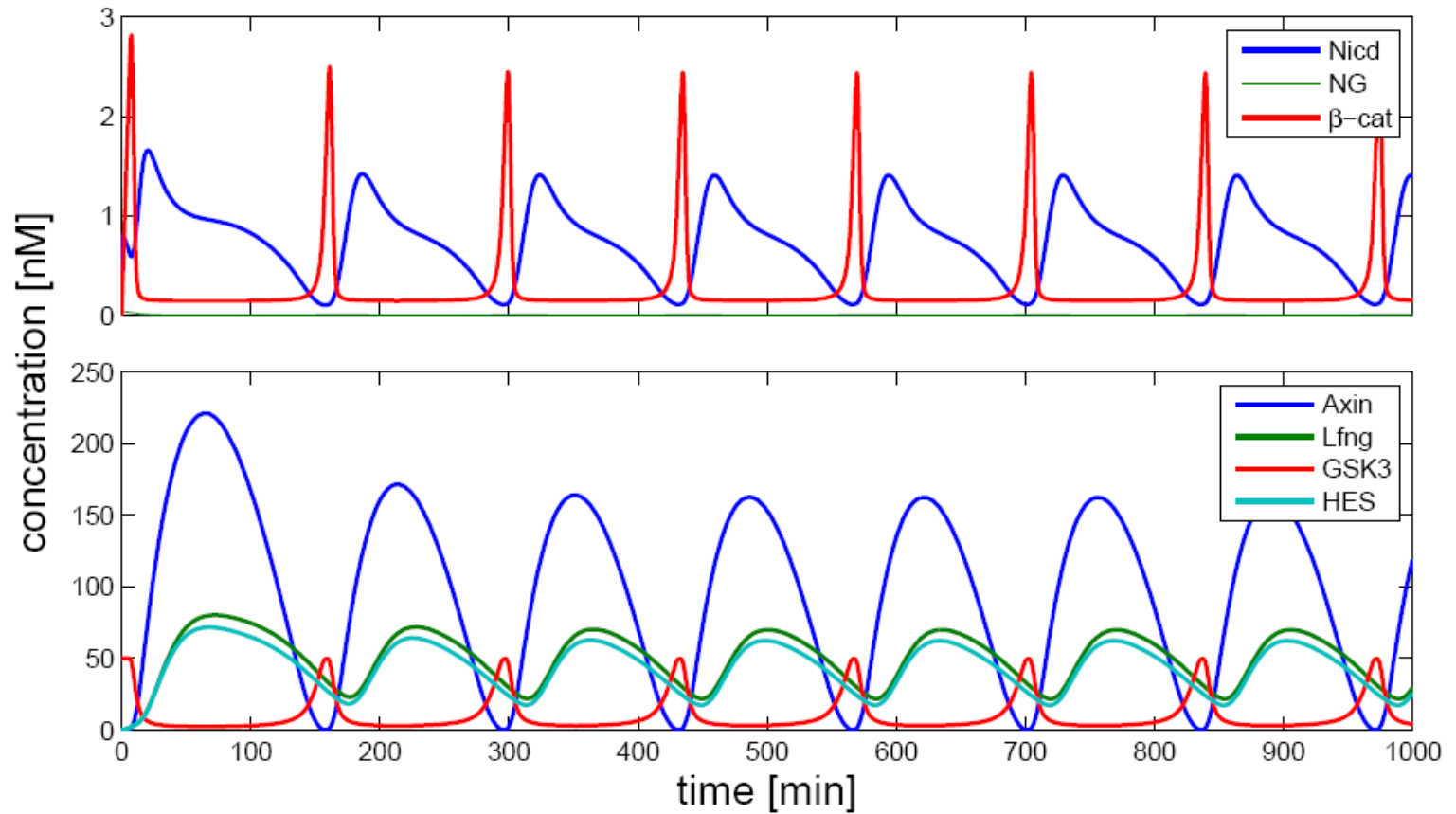
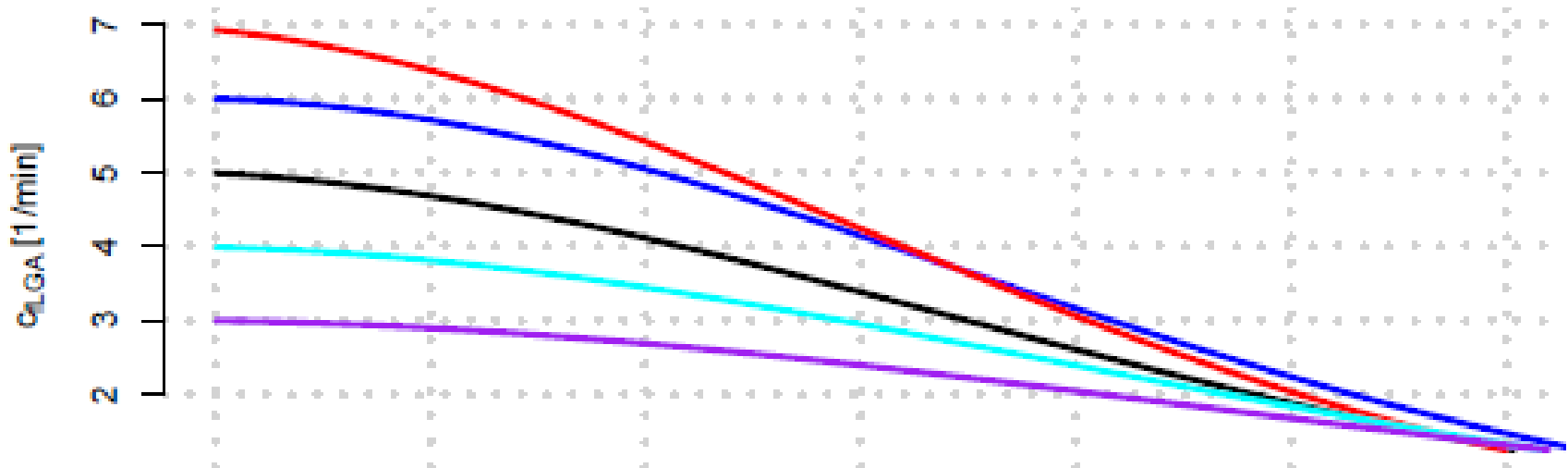


Figure 7.2: Timeseries of the 12 equation system with Lfng negative feedback

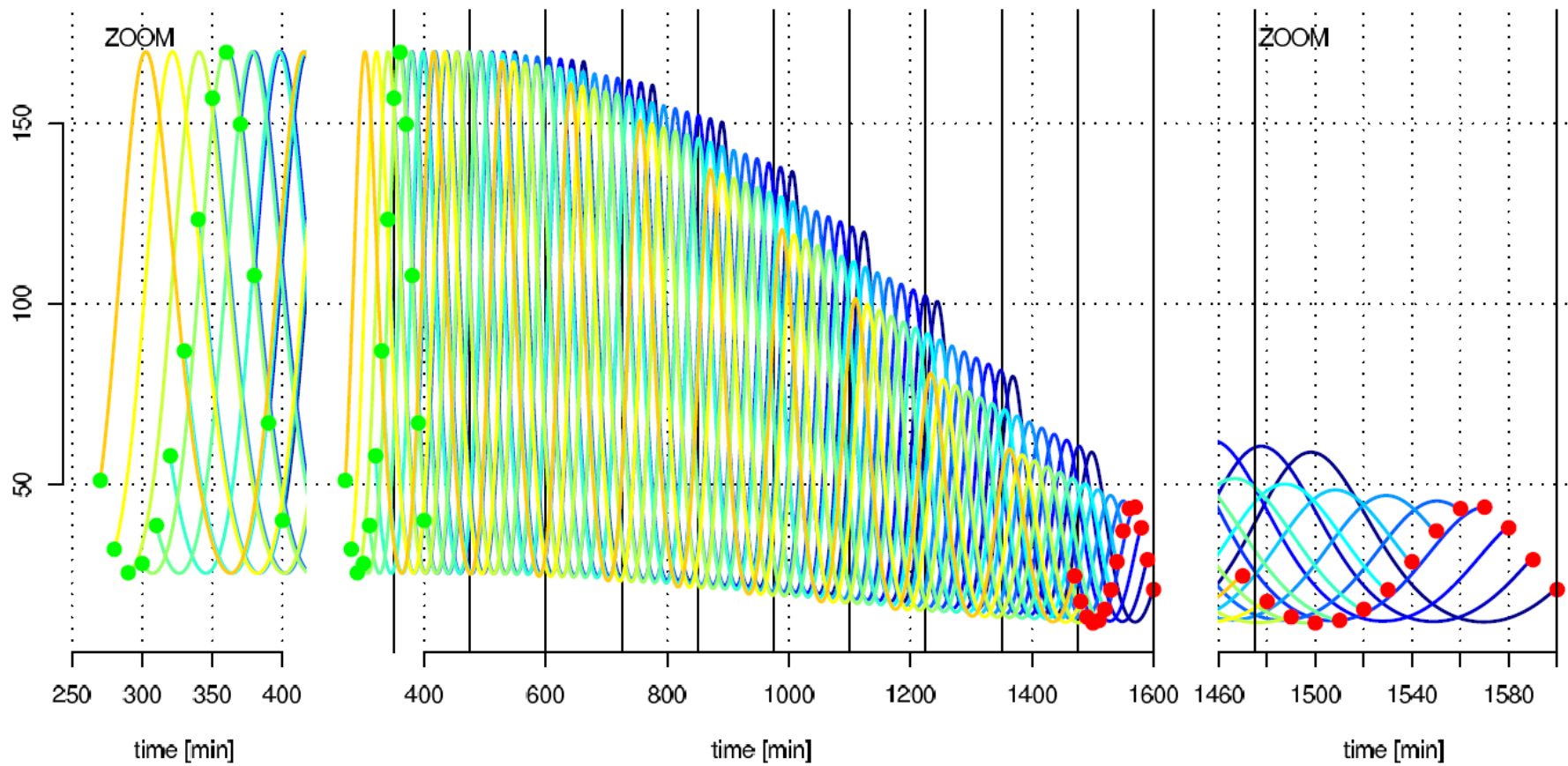
Spatial gradient of Wnt:

Gaussian profiles



Oscillations stop when Wnt under a threshold

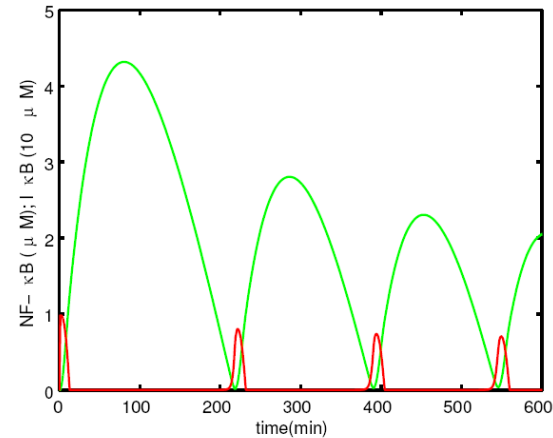
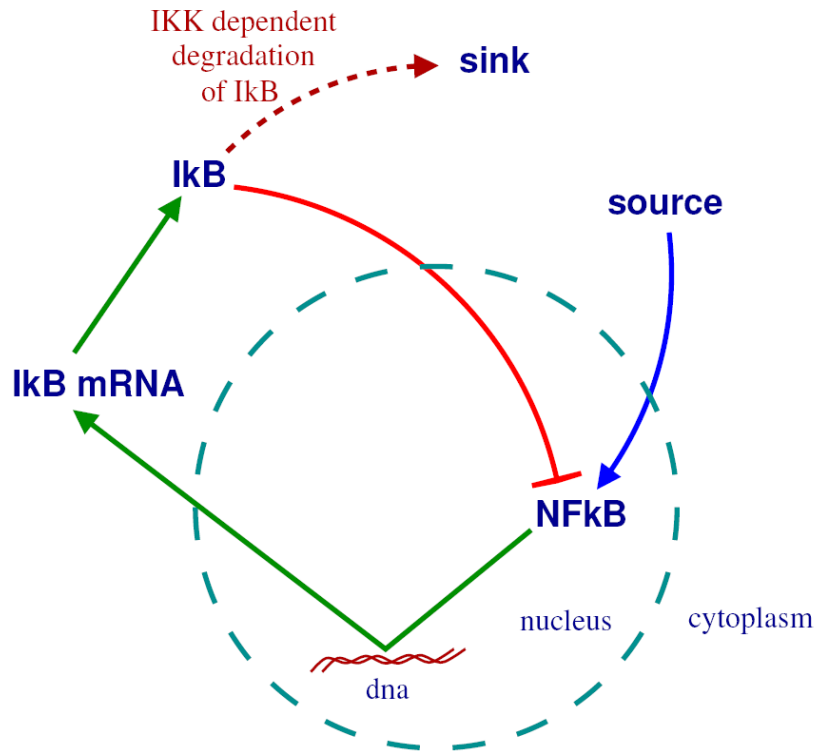
Axin2 nM



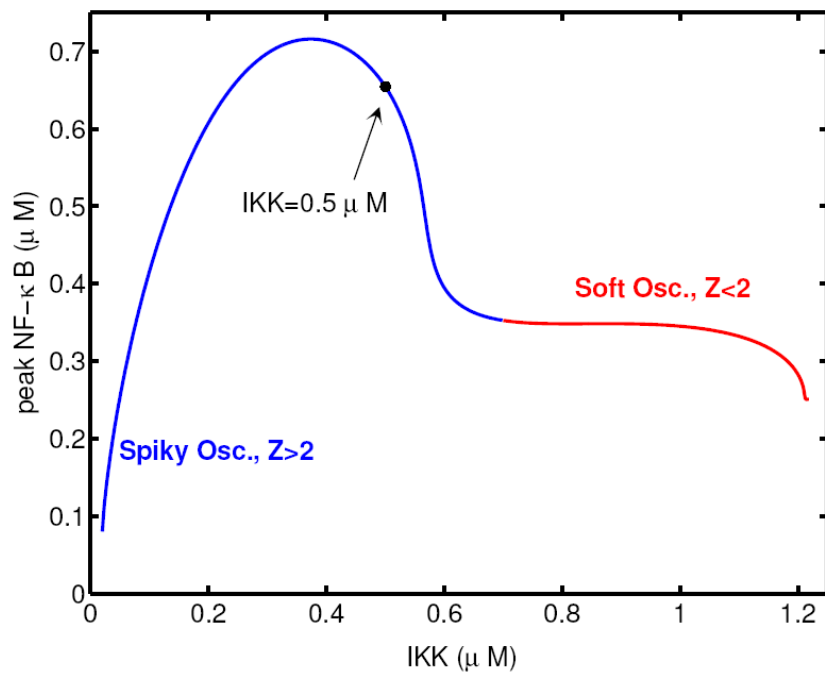
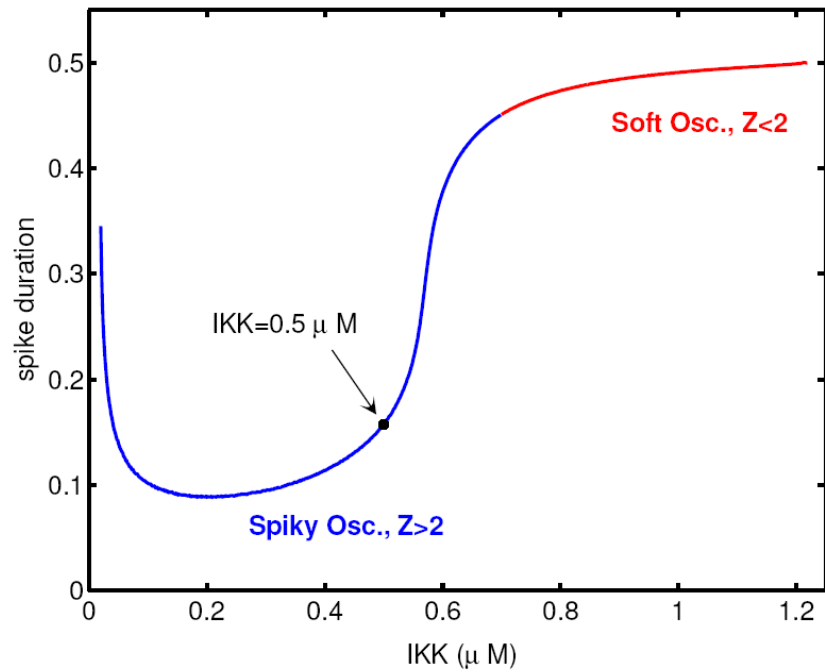
Simple Model for Protein Oscillations

Simple model qualitatively reproduces several features of NF- κ B oscillations:

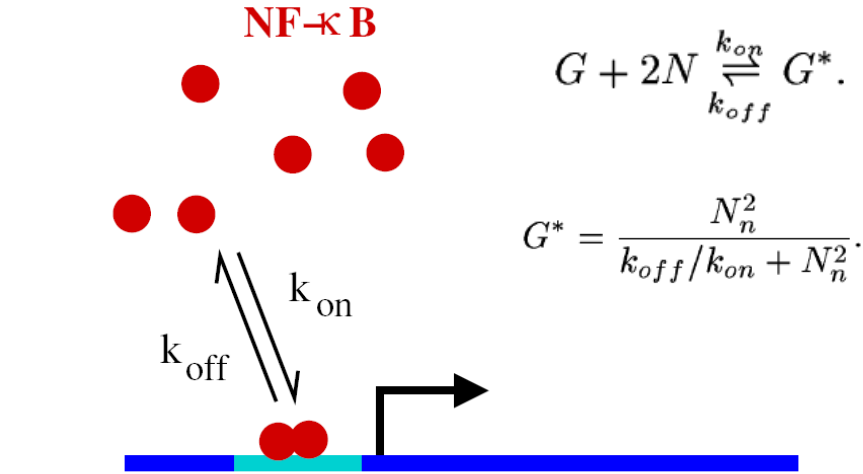
- fact of oscillations
- time period
- shape of oscillations
- phase relationships
- no osc. in the absence of feedback
- increased transcription \rightarrow lower freq.



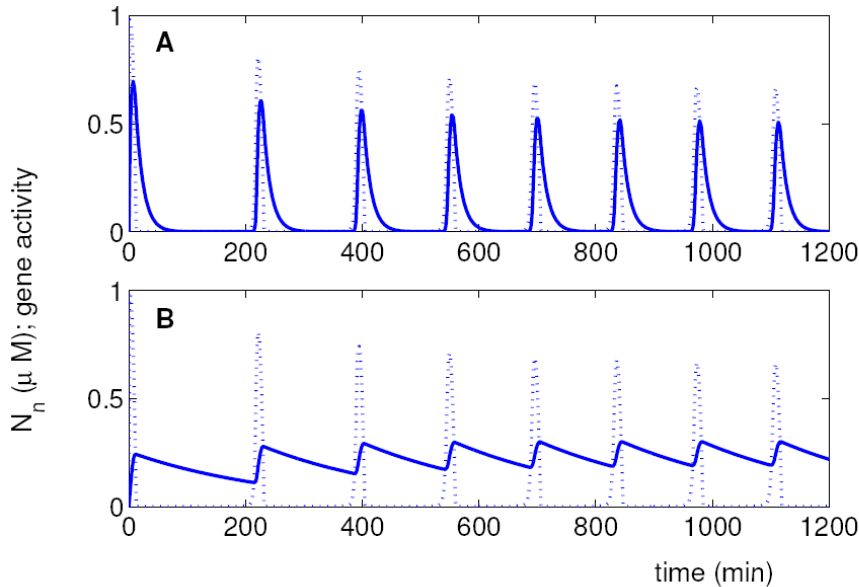
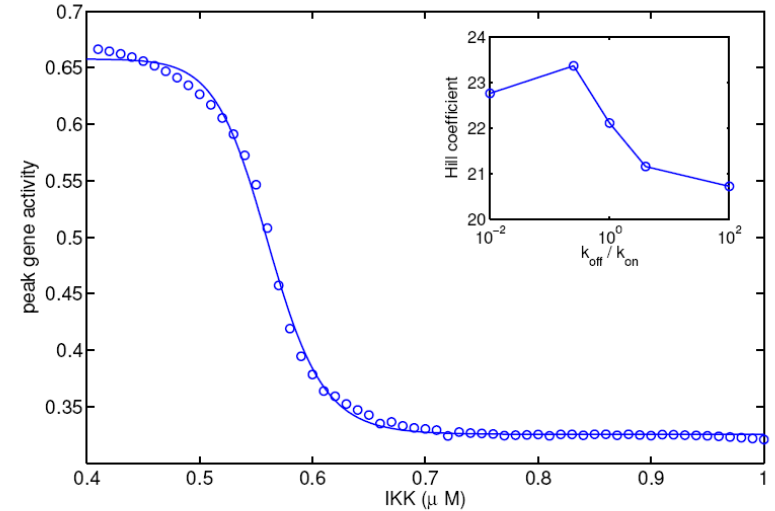
$$A = 0.007, B = 954.5, C = 0.035, \\ \delta = 0.029, \epsilon = 2 \times 10^{-5}$$



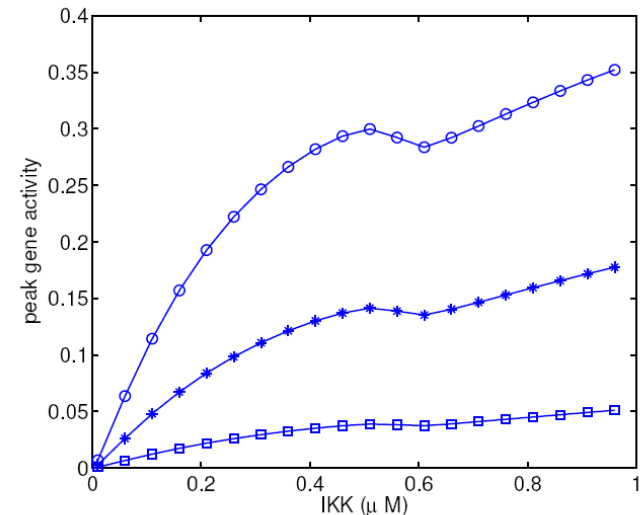
Coupling NF- κ B to a Downstream Gene



Equilibrium binding, $1/k_{off} \ll$ time period



Non-equilibrium binding, $1/k_{off} >$ time period



Variable	Description	
N_n	nuclear NF- κ B	
$I_{\alpha/\varepsilon}$	free I κ B	
$I_{m\alpha/\varepsilon}$	I κ B mRNA	
A_m	A20 mRNA	
K	active IKK	
K_i	inactive IKK	
Parameter	Description	Value
T	TNF stimulation	1
B	proportionality factor of the export of nuclear NF- κ B	102.6
A	proportionality factor of the import of NF- κ B	0.004
η		0.092
K_I	I κ B - NF- κ B complex dissociation factor	$1.26 \cdot 10^{-5} \mu M$
δ	concentration at which half of the I κ B α/ε is bound in complex with NF- κ B	$0.0414 \mu M$
p	NF- κ B in-dependent transcription rate of I κ B α mRNA	$3.36 \cdot 10^{-5} \text{ min}^{-1}$
t_a	NF- κ B dependent transcription rate of I κ B α mRNA	$0.0042 \mu M \text{ min}^{-1}$
t_e	NF- κ B dependent transcription rate of I κ B ε mRNA	$0.084 \mu M \text{ min}^{-1}$
t_A	NF- κ B dependent A20 transcription rate	$0.0168 \mu M^{-1} \text{ min}^{-1}$
tl_a	translation rate of I κ B α	0.0672 min^{-1}
tl_e	translation rate of I κ B ε	$1.2 \cdot 10^{-5} \text{ min}^{-1}$
tl_A	translation rate of A20	0.3024 min^{-1}
$\gamma_{Im\alpha}$	half-life of I κ B α mRNA	0.0168 min^{-1}
$\gamma_{Im\varepsilon}$	half-life of I κ B ε mRNA	0.00168 min^{-1}
$\gamma_{I\alpha/\varepsilon}$	half-life of the I κ B's	0.005 min^{-1}
γ_{A20m}	half-life of the A20 mRNA	0.0168 min^{-1}
γ_{A20}	half-life of the A20	0.001 min^{-1}
α_α	IKK dependent degradation of I κ B α	0.00025 min^{-1}
α_ε	IKK dependent degradation of I κ B ε	$7.6 \cdot 10^{-6} \text{ min}^{-1}$
μ	rate of IKK self-inactivation	0.063 min^{-1}
σ	strength of A20 negative feedback	0.25
β	proportionality factor of A20 on IKK	1.25

When oscillations in feed-back loops?

- Process that takes a finite (minimum) time
evt. time delay τ : $dx/dt \sim P(t-\tau)$
- Many intermediate steps
binding, complex, steps on DNA, etc
several components: repressilator
- Sharp response: high Hill coefficient
 $dm/dt \sim p^n/(k^n + p^n)$
- Saturated degradation: depends on level
 $dI/dt \sim I_m - cI/(\epsilon+I)$
- Autocatalysis: $dx/dt \sim x^n/(k^n + x^n)$

k_g : dissociation constant between p53 and DNA (O-operator site)

$$[p] = [p_f] + [pm] \quad [O] = [O_f] + [pO] \quad ([O] = 1)$$

$$k_g = \frac{[p_f][O_f]}{[pO]} \Rightarrow [pO] = \frac{[p_f][O]}{k_g + [p_f]} = \frac{[p] - [pm]}{k_g + [p] - [pm]}$$

(can add a Hill coefficient)

$[pO]$: "Equilibrium" probability that p53 is bound to DNA

k : dissociation constant between p53 and mdm2

$$k = \frac{[p_f][m_f]}{[pm]} = \frac{([p] - [pm])([m] - [pm])}{[pm]} \Rightarrow$$

$$[pm]^2 - ([p] + [m] + k)[pm] + [p][m] = 0 \Rightarrow$$

$$[pm] = \frac{1}{2}([p] + [m] + k) - \sqrt{([p] + [m] + k)^2 - 4[p][m]}$$

Why oscillations ?

- Importance for apoptosis
- Essential in segmentation
- Spiky oscillations →
important for **sharp responses**,
fast regulations,
high Hill coefficients
→ hormones also come in spikes

Mathematically: The most 'simple' dynamics !