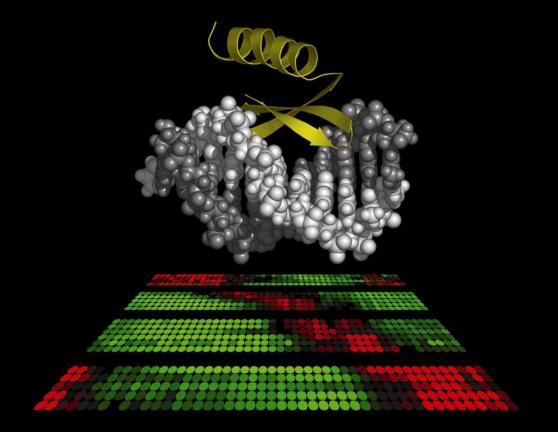
Structure, evolution and dynamics of gene regulatory networks



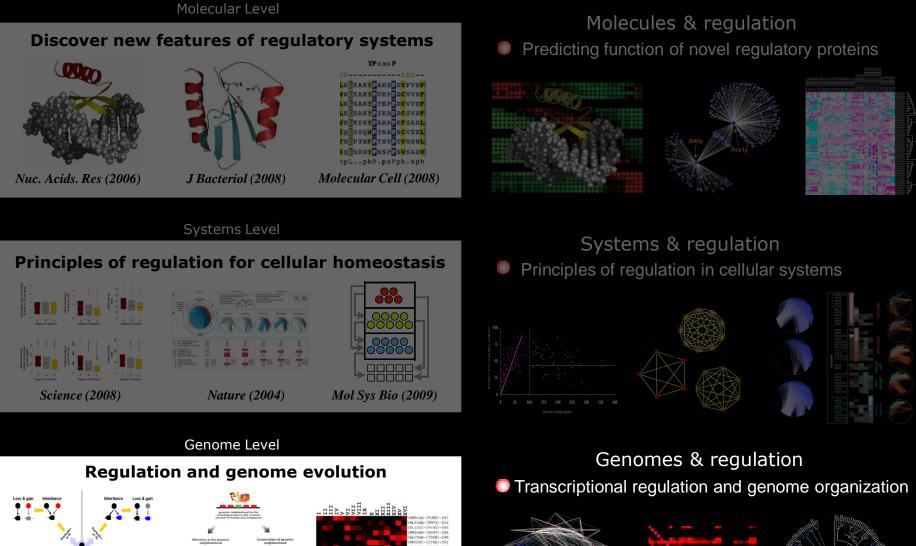
M. Madan Babu

Group Leader MRC Laboratory of Molecular Biology, Cambridge

Networks in Biology

Network	Protein Interaction	Metabolic	Transcriptional	
Nodes 🔵	Proteins	Metabolites	Transcription factor Target genes	
Links 🗕 🕘	Physical Interaction	Enzymatic conversion	Transcriptional Interaction	
Interaction	Protein-Protein	Protein-Metabolite	Protein-DNA	
A B B	A I B B		A ↓ B	

Overview: How is regulation achieved in cellular systems?



Nature Genetics (2004)



Genome research (2009)

PNAS (2008)



Outline

Structure of the transcriptional regulatory network

Evolution of the transcriptional regulatory network

Dynamic nature of the transcriptional regulatory network

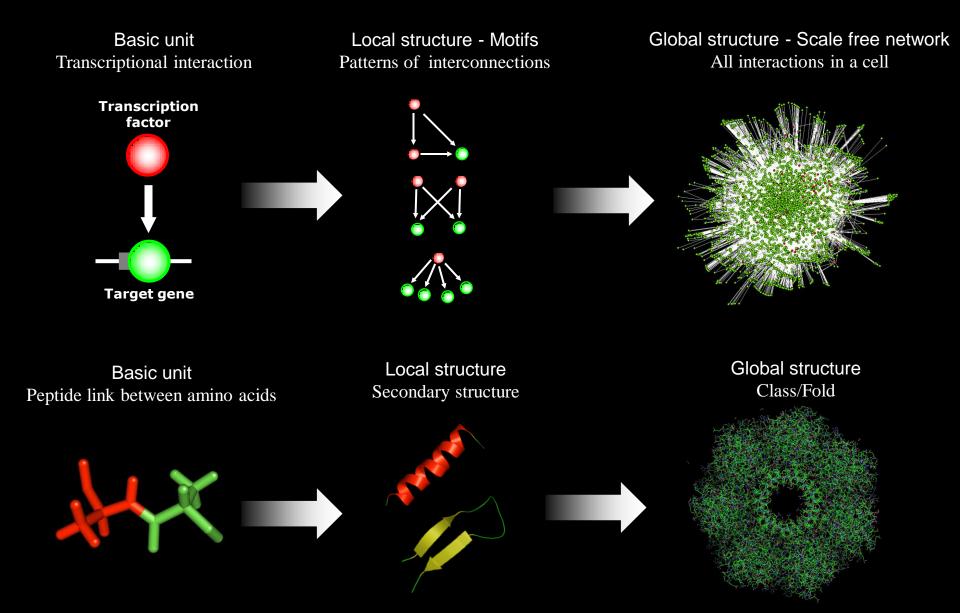
Hierarchy and node-dynamics of regulatory networks

Detailed Outline

Structure of the transcriptional regulatory network

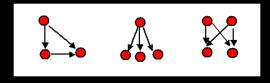
- Local network structure: network motifs
- Global network structure: scale-free structure
- Evolution of the transcriptional regulatory network
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Organization of the transcriptional regulatory network analogy to the organization of the protein structures



Properties of transcriptional networks

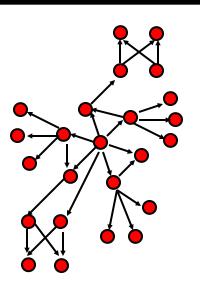
Local level: Transcriptional networks are made up of motifs which perform information processing task



Global level: Transcriptional networks are scale-free conferring robustness to the system



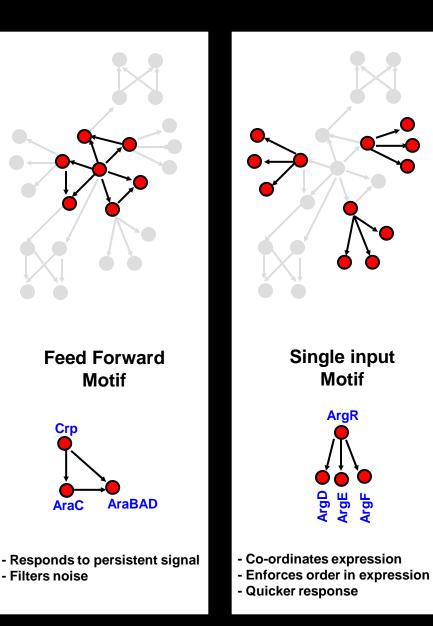
Transcriptional networks are made up of motifs

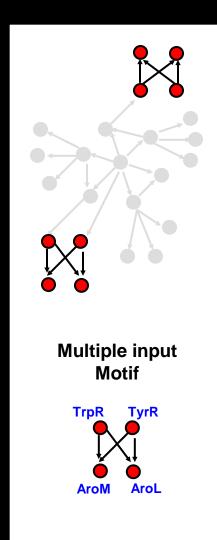


Network Motif

"Patterns of interconnections that recur at different parts and with specific information processing task"

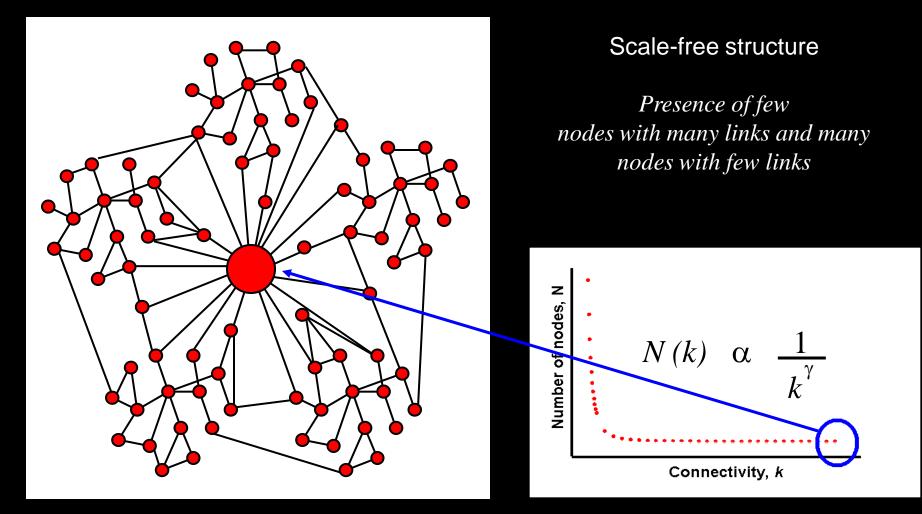
Function





Integrates different signals
Quicker response

Transcriptional networks are scale-free

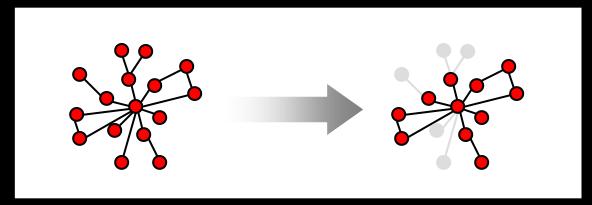


Scale free structure provides robustness to the system

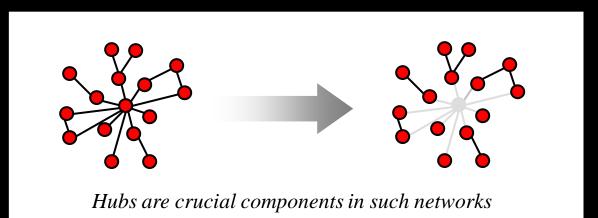
Scale-free networks exhibit robustness

Robustness – The ability of complex systems to maintain their function even when the structure of the system changes significantly

Tolerant to random removal of nodes (mutations)

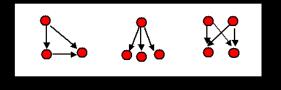


Vulnerable to targeted attack of hubs (mutations) – Drug targets

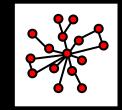


Summary I – Structure of transcriptional networks

Transcriptional networks are made up of motifs that have specific information processing task



Transcriptional networks have a scale-free structure which confers robustnessto such systems, with hubs assuming importance

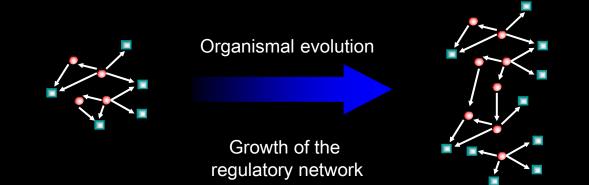


Madan Babu M, Luscombe N et. al Current Opinion in Structural Biology

Detailed Outline

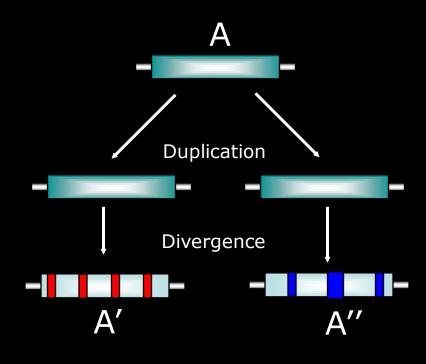
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Evolution of the transcriptional regulatory network in yeast



How did the regulatory network evolve?

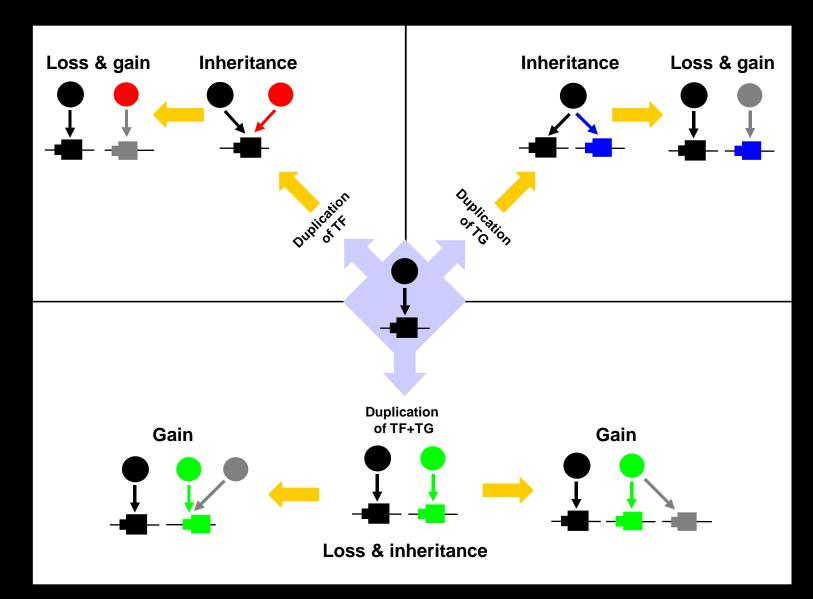
Mechanisms for the creation of new genes



Duplication & Divergence

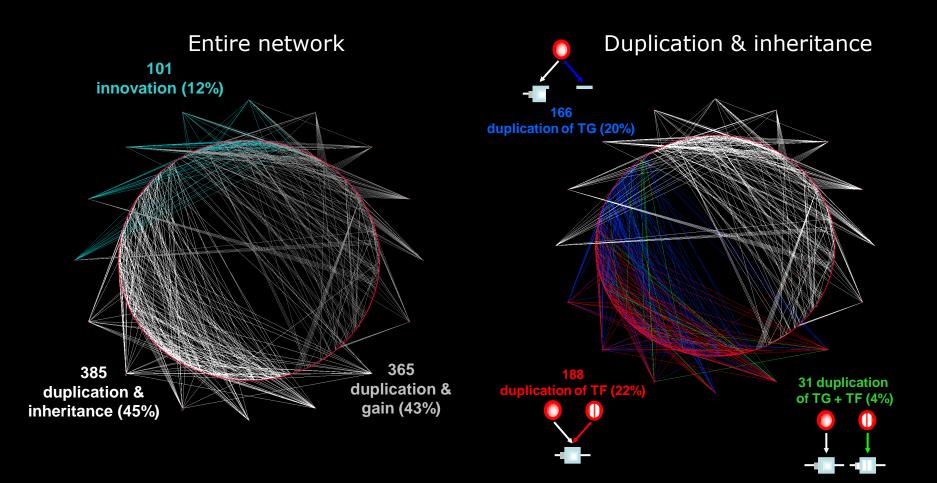
OR Recombination/Innovation

Gene duplication and network growth



Inheritance, loss and gain of interaction

Evolution of the transcriptional regulatory network

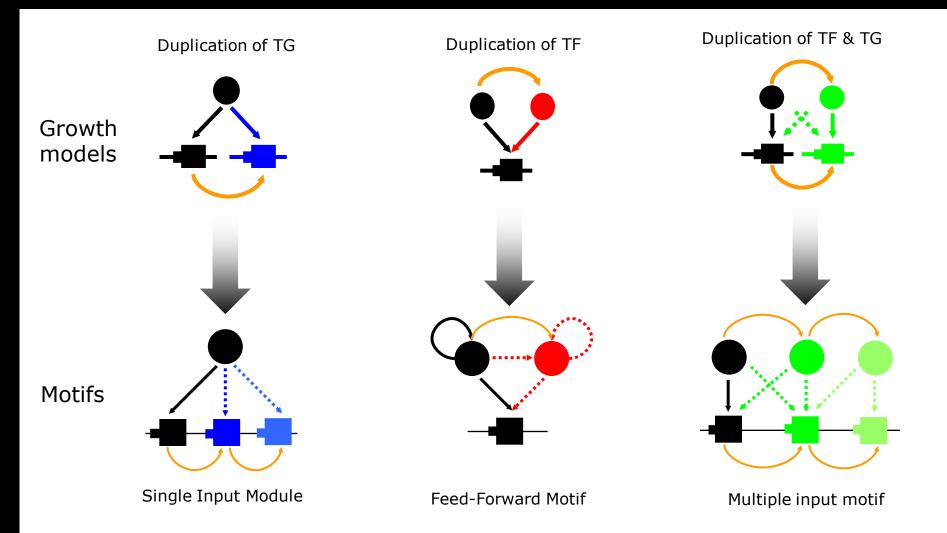


~90% of the network has evolved by duplication followed by Inheritance, loss and gain of interaction How can such events affect local and global network structure?

Are the motifs and scale free structure products of duplication events ?

Are the motifs and scale free structure selected for in evolution ?

Duplication models and evolution of network motifs



Evolution of local network structure

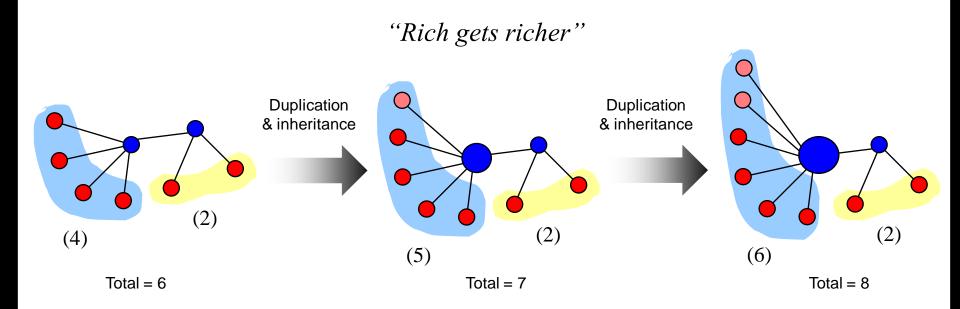
Very rarely we find instances where duplication events have resulted in the formation of network motifs



Network motifs have evolved independently (convergent evolution) multiple times because they confer specific properties to the network

Duplication and evolution of scale free structure

Growth by gene duplication and inheritance of interaction can explain evolution of scale-free structure



Regulatory hubs should control more duplicate genes

Evolution of global network structure

Regulatory hubs do not regulate duplicate genes more often than any other normal transcription factor

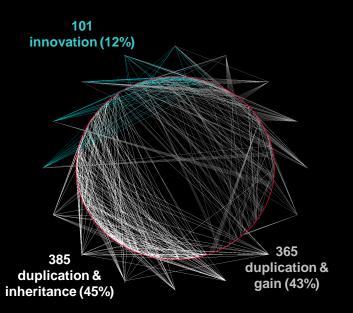


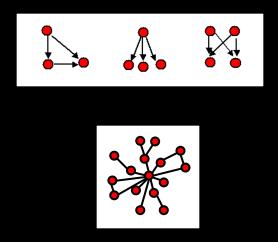
Scale free structure has been selected for in evolution and is not a product of duplication events

Summary II - Evolution

Gene duplication followed by inheritance of interaction and gain of new interactions have contributed to 90% of the network

Network motifs and the scale free structure are not products of duplication events, but have been selected for in evolution



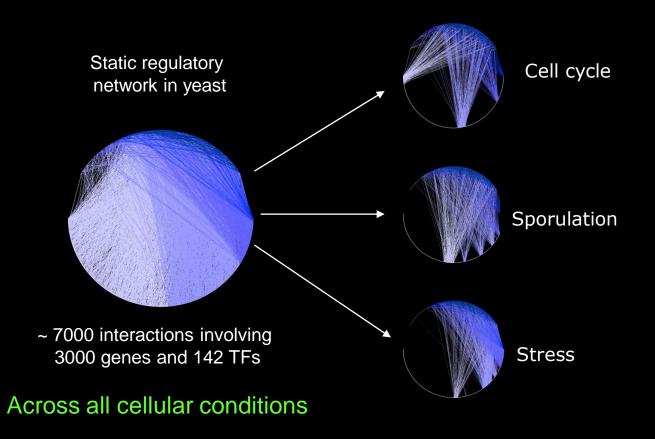


Teichmann SA & Madan Babu M Nature Genetics

Detailed Outline

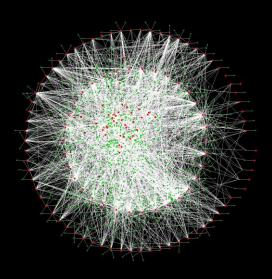
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 - Dynamics of global network structure
- Hierarchy and node-dynamics of regulatory networks

Temporal dynamics of the regulatory networks



How does the local structure change in different cellular conditions? How does the global structure change in different conditions?

Integrating gene regulatory network with expression data



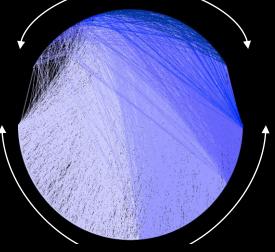
142 TFs 3,820 TGs 7,074 Interactions

Gene expression data

for 5 cellular conditions

Cell-cycle Sporulation DNA damage Diauxic shift Stress

Transcription Factors

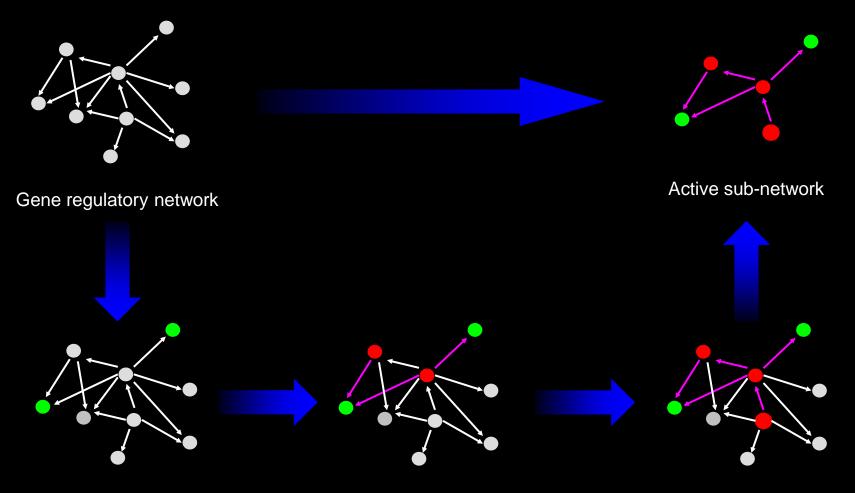


Target Genes

142 TFs 1,808 TGs 4,066 Interactions



Back-tracking method to find active sub-networks



Identify differentially regulated genes

Find TFs that regulate the genes

Find TFs that regulate these TFs

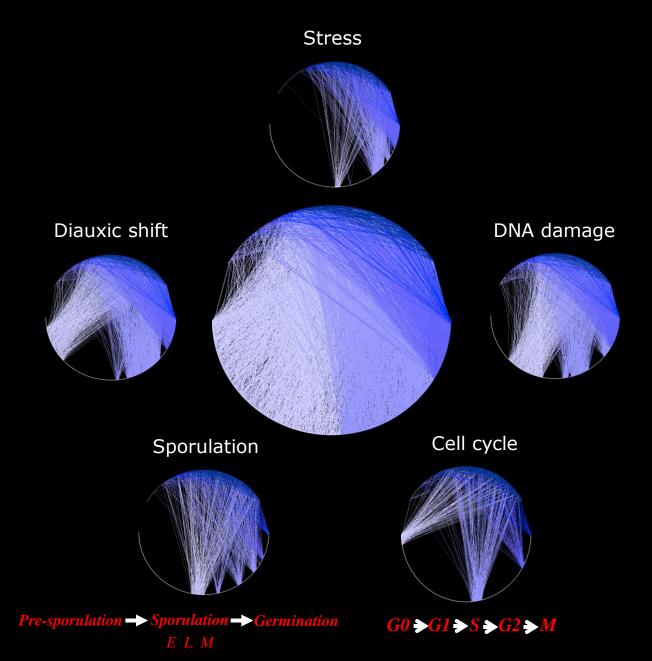
Regulatory program specific transcriptional networks

Binary Processes

Regulatory programs involved in survival

> Multi-step Processes

Regulatory programs involved in development



Temporal dynamics of local structure

Multi-step regulatory programs (development) Binary regulatory programs (survival)

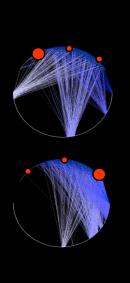
	Motifs	Cell cycle	Sporulat ion	Diauxic shift	DNA damage	Stress response
fast acting &	SIM	32.0%	38.9%	57.4%	55.7%	59.1%
direct						
fast acting & direct	MIM	23.7%	16.6%	23.6%	27.3%	20.2%
slow acting &	FFL	44.3%	44.5%	19.0%	17.0%	20.7%
indirect						

Network motifs that allow for efficient execution of regulatory steps are preferentially used in different regulatory programs

Temporal dynamics of global structure (hubs)

Condition specific hubs

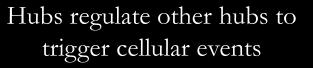
Each regulatory program is triggered by specific hubs

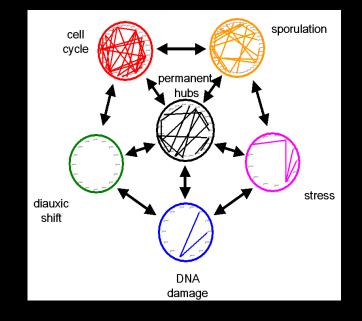




Permanent hubs

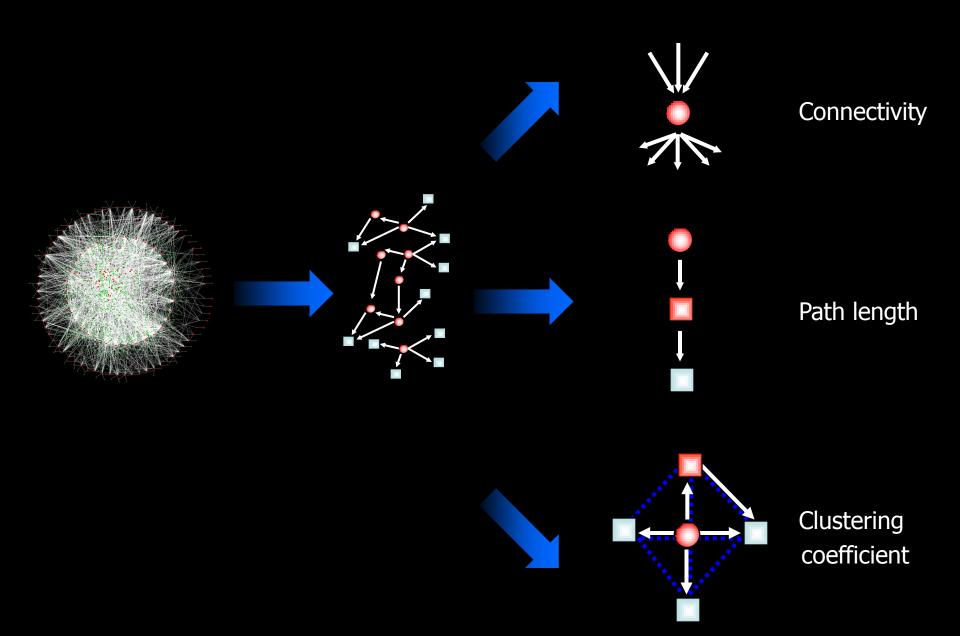
Active across all regulatory programs



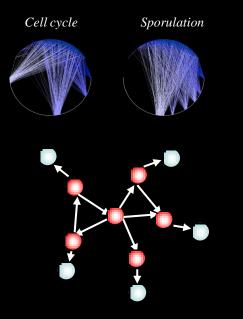


This suggests a dynamic structure which transfers 'power' between hubs to trigger distinct regulatory programs (developmental & survival)

Network Parameters



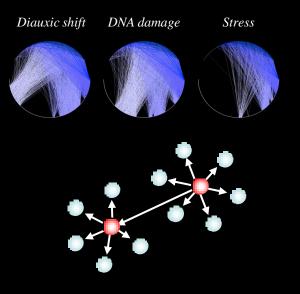
multi-stage conditions



- fewer target genes per TF
- longer path lengths
- more inter-regulation between TFs

Fidelity in response

binary conditions



- more target genes per TF
- shorter path lengths
- less inter-regulation between TFs

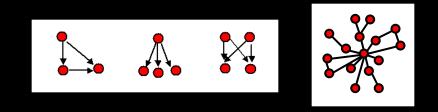
Quick response

Sub-networks re-wire both their local and global structure to respond to cellular conditions efficiently

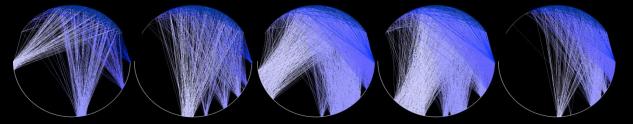
Luscombe N, Madan Babu M et. al Nature (2004)

Summary III - Dynamics

Network motifs are preferentially used under the different cellular conditions and different proteins act as regulatory hubs in different cellular conditions



Sub-networks have evolved both their local structure and global structure to respond to cellular conditions efficiently



Luscombe N, Madan Babu M et. al Nature (2004)

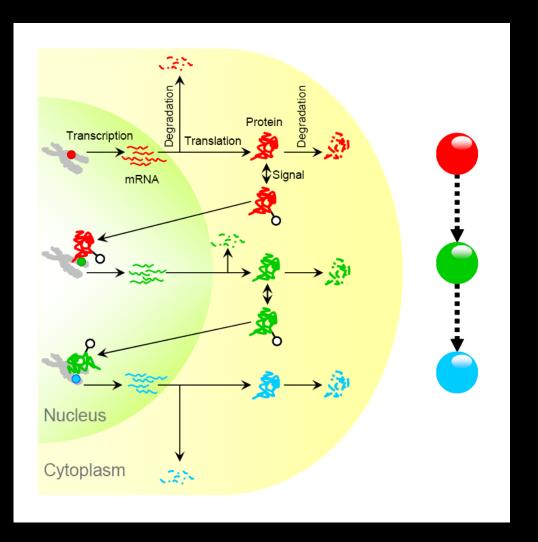
Detailed Outline

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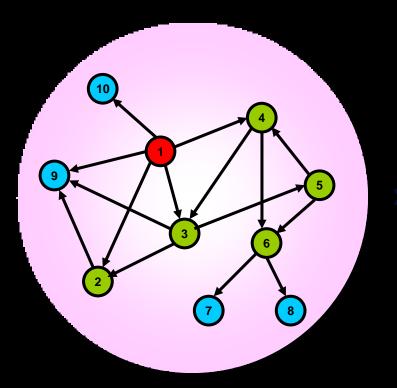
Hierarchy and node-dynamics of regulatory networks

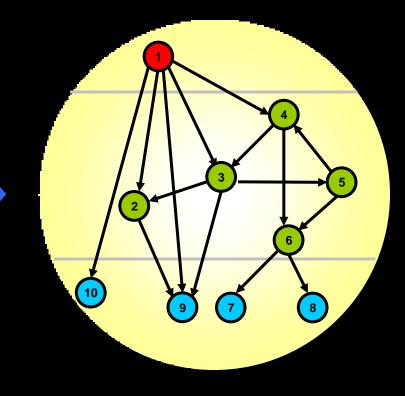
- Hierarchy in transcriptional networks
- Noise in gene expression implications for transcriptional networks

Dynamics in transcription and translation



Each node in the network represents several entities (gene, mRNA, and protein) and events (transcription, translation, degradation, etc) that are compressed in both space and time Higher-order organization of transcriptional regulatory networks





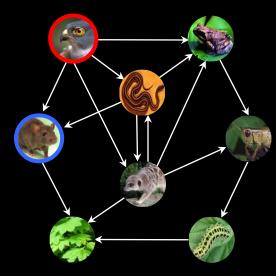
Inherent higherstoratenarganization

Hierarchical structure

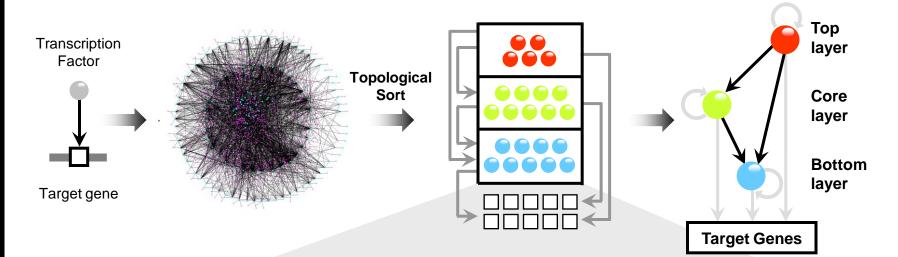
Methods to infer hierarchical organization in networks

Food web network

predator \rightarrow prey



Hierarchical organization of the yeast regulatory network

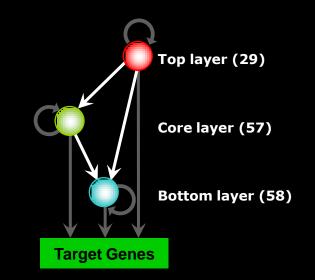


(25)	Level 7	OAF1	MAC1	MBP1*	SKN7*	ARG80 DAL82	ARO80 FLO8 *	AZF1 HAL9	CHA4 MET31						
om (59) Core (64) Top	Level 6	ABF1*◀	MAL33	MCM1 <	NRG1*	MET32	PIP2	ZAP1	IVIE 101	Т					
	Level 5	ACE2 CUP9 GAT3 HMLALPHA2 MIG1	ADR1 DAL80 GCN4* HMS2 MIG2	AFT1* DAL81 GLN3 HSF1* ◀ MSN2*	AFT2* FHL1* GTS1 INO2 MSN4*	ARG81 FKH1 GZF3 INO4 NDT80	ASH1 FKH2* HAP1 IXR1 PHD1*	CBF1* GAL4 HAP4 LEU3 PLM2*	CIN5* GAT1 HCM1* MGA1* PUT3	PDC2 <	ARR1 DAT1 RTG1	OTU1 RPH1 PHO2	PHO2		RDR1 RLM1 THI2 UPC2 YBL054W YDR266C YJL206C
		RAP1* ◀ STE12* XBP1	REB1* SUT1 YAP1	SWI4* YAP5*	ROX1 SWI5 YAP6*	RPN4 TEC1* YAP7*	TOS4* YHP1	SMP1 TOS8* YOX1*	SOK2* TYE7 UME6*					EDS1	
	Level 4	RTG3		CAT8	HMS1	FZF1	HAC1	MET4◀	ACA1					YKR064W	
	Level 3	HMRA1	HMRA2	SPT23	STP1	PDR1	PHO4	CRZ1	CAD1	CUP2					YRM1
	Level 2	RME1	SIP4 STP2	BAS1	RGT1 YAP3	SUM1 YDR026C	UGA3	CST6 GCR1◀	ECM22 MOT3	LYS14 MSN1	MAL13 MSS11				
Bottom	Level 1	IME2 SFL1 SRD1	SFL1		MET28 STB4	MIG3 STB5	PDR3 YDR049W	PDR8 YRR1	PPR1 STP4	RDS1 WAR1 YER184C	RGM1 URC2 YML081W	SFP1 USV1 YPR196W			
		SKUT							TER 130C	1EK 1040		IPRI	9000		

* Regulatory hubs

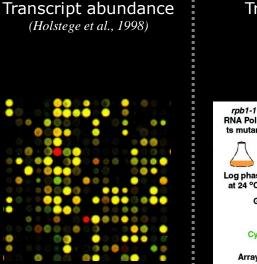
Hierarchical organization of regulatory proteins

Regulatory proteins are hierarchically organized into three basic layers

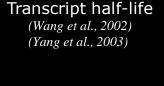


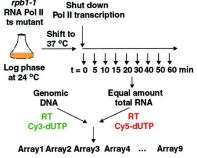
Do TFs in the different hierarchical levels have distinct dynamic properties?

Datasets characterizing dynamics of transcription and translation



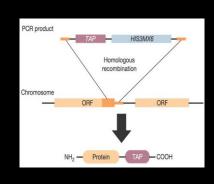
Transcript abundances for yeast grown in YPD (S. cerevisiae) and Edinburgh minimal medium (S. pombe) were determined by using an Affymetrix high density oligonucleotide array.



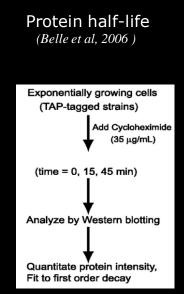


Transcript half-lives were determined by obtaining transcript levels over several minutes after inhibiting transcription. This was done using the temperature sensitive RNA polymerase rpb1-1 mutant S. cerevisiae strain.





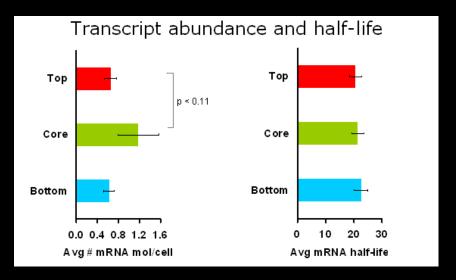
Estimates of the endogenous protein expression levels during log-phase were obtained by TAPtagging every yeast protein for S. cerevisiae.



Protein half-lives were determined by first inhibiting protein synthesis via the addition of cyclohexamide and by monitoring the abundance of each TAP-tagged protein in the yeast genome as a function of time.

Translational dynamics

Regulation of regulatory proteins within the hierarchical framework



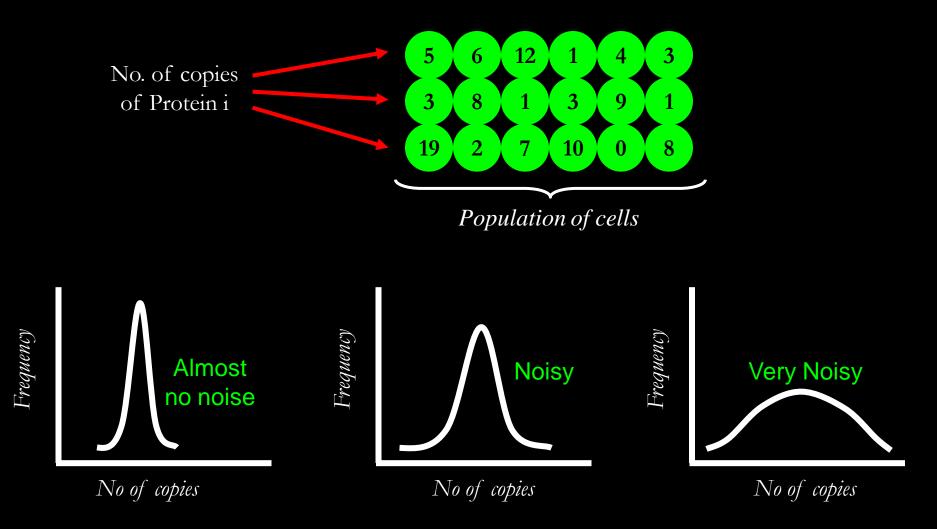
Protein abundance and half-life Тор Тор p < 0.066 p < 0.033 p < 0.087 p < 0.016 Core Core Bottom Bottom 2.000 4.000 40 20 60 Avg # protein mol/cell Avg protein half-life

Regulation of transcript abundance or degradation does not appear to be a major control mechanism by which the steady state levels of TFs are controlled Regulation of protein abundance and degradation appears to be a major control mechanism by which the steady state levels of TFs are controlled

post-transcriptional regulation plays an important role in ensuring the availability of right amounts of each TF within the cell

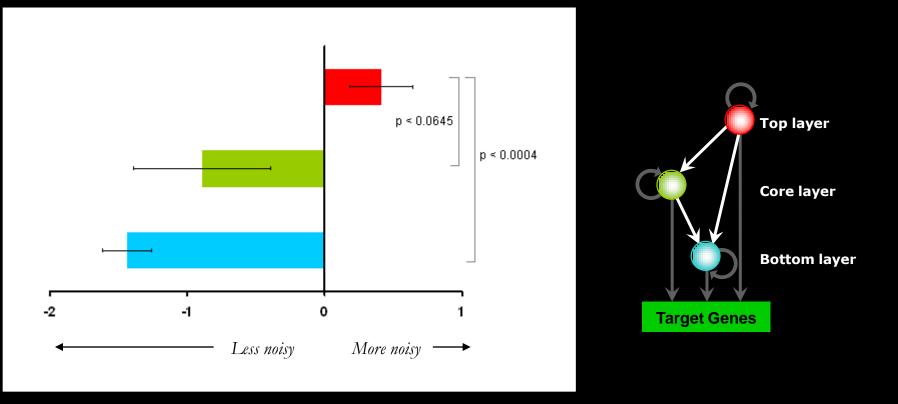
Noise in protein levels in a population of cells

Noise in a population of cells can be beneficial where phenotypic diversity could be advantageous but detrimental if homogeneity and fidelity in cellular behaviour is required



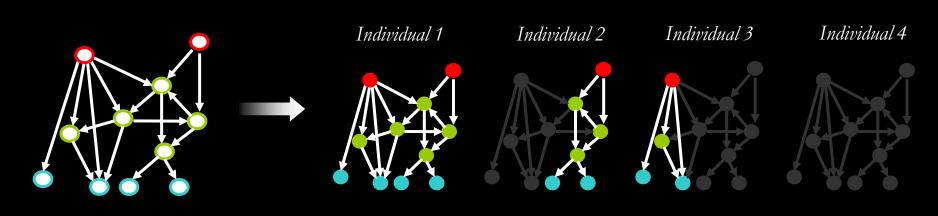
Lopez-Maury L, Marguaret S and Bahler J, Nat Rev Genet 2008

Noise levels of regulatory proteins



Regulatory proteins in the top layer are more noisy than the ones in the core or bottom layer

Implication

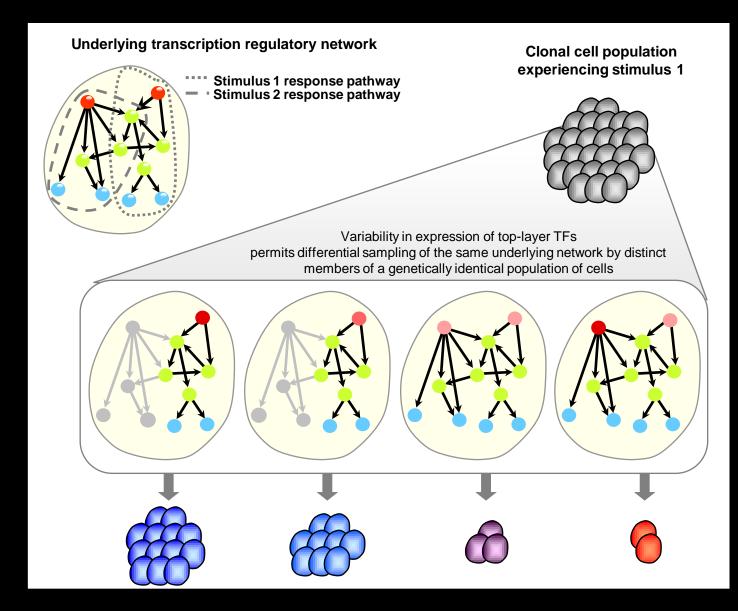


Underlying network

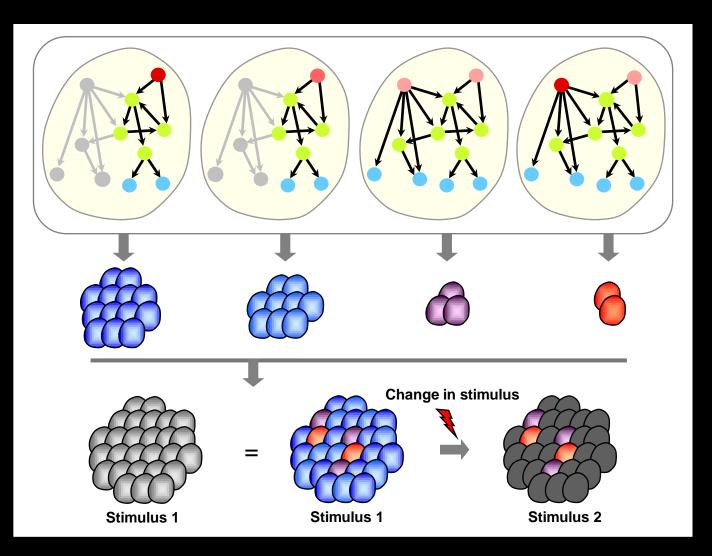
Differential utilization of the same underlying network by different individuals in a population of cells

Noise in TF expression may permit differential utilization of the same underlying regulatory network in different individuals of a population

Non-genetic variation due to differential transcription network utilization

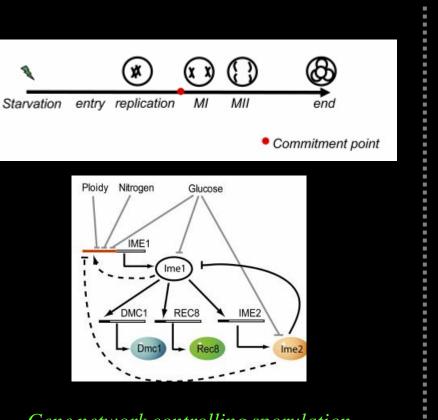


Non-genetic variation due to differential transcription network utilization

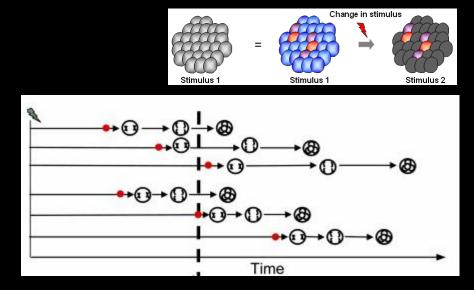


Non-genetic variability and dynamics in expression of key TFs might confer selective advantage as this permits at least some members in a clonal population to respond efficiently to (or survive in) changing conditions

Phenotypic variability in fluctuating environments



Gene network controlling sporulation

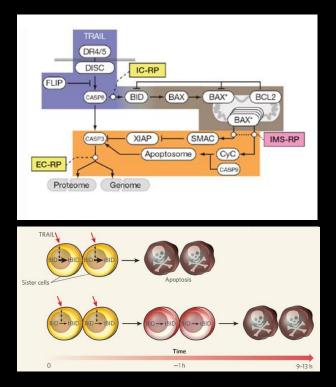


High variability in the expression of top-level TFs in a population of cells may confer a selective advantage as this permits at least some members in a population to respond quickly to changing conditions

Noise in expression of a master regulator of sporulation in yeast

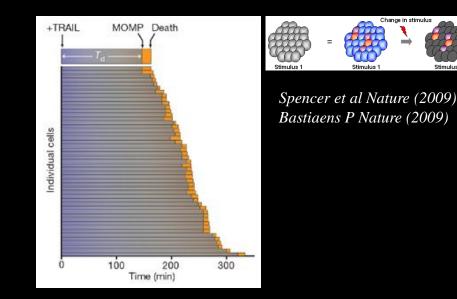
Nachman et al Cell (2008)

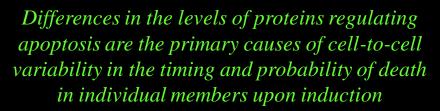
Differential cell-fate outcome in response to uniform stimulus



Network controlling apoptosis

Noise in expression of a key regulator of apoptosis in human cancer cells





Dynamics of the regulatory proteins, which either dictate cell death or survival, varied widely between individual cancer cells upon addition of a drug

Cohen et al Science (2008)

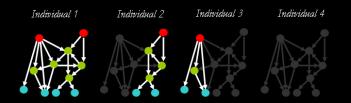
Change in stimulu

Summary IV – local dynamics of regulatory networks

Our results suggest that the core- and bottom-level TFs are more tightly regulated at the post-transcriptional level rather than at the transcriptional level itself



Our findings suggest that the interplay between the inherent hierarchy of the network and the dynamics of the TFs permits differential utilization of the same underlying network in distinct members of a population



Jothi R, Balaji S et al., Mol Sys Biol (2009)

Conclusions

"Transcriptional networks are made up of network motifs at the local level and have a scale-free structure at the global level"

"Even though close to 90% of the regulatory network in yeast has evolved by duplication, network motifs and scale free structure are not products of duplication events – instead they have been selected for in evolution"

"This has resulted in a network structure that can efficiently re-wire interactions to meet the biological demand placed by the process"

"The interplay between the hierarchy and node dynamics makes the network both robust and adaptable to changing conditions"

"All these aspects of the regulatory network has constrained the way genes are organized across the different linear chromosomes in yeast"

Acknowledgements

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Arthur Wuster Joerg Gsponer MRC-LMB, UK

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MRC - Laboratory of Molecular Biology, UK National Institutes of Health, USA Schlumberger and Darwin College, Cambridge, UK

Regulatory Genomics and Systems Biology Group



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O Pradeep Kota, Visiting student 2007

Members of the **Theoretical and Computational Biology** at the LMB group for helpful discussions



Nitish Mittal Visiting PhD Student 2008 2009

Rekin's

Janky

Post

Doctoral

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2007 2010

Sarath

Janga

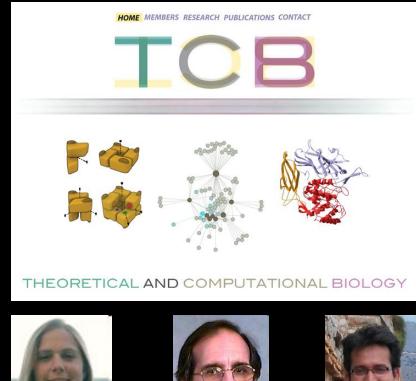
PhD

Student

2007

2010

Interested in our research?





Teichmann



Chothia



Babu

post-doctoral applications invited madanm@mrc-lmb.cam.ac.uk