

# Antimicrobial polymer action: towards coarse graining

Vani Vemparala  
Institute of Mathematical Sciences  
Chennai, India

December 19 2009

Collaborators:

Experimental:

Kenichi Kuroda (University of Michigan)

Atomistic Simulations/FF development:

Ivaylo Ivanov (Georgia State University)

Vojislava Pophristic (University of the Sciences-Philadelphia)

Michael Klein (Temple University)

Coarse Graining:

Wataru Shinoda ( AIST, Tsukuba, Japan)

# Research Interests

Tools: Classical Molecular Dynamics, supplemented by force field development using (minimal) quantum chemistry tools, **Coarse grain modeling**

Systems: Polymers, Proteins, Lipid Membranes

## Polymers:

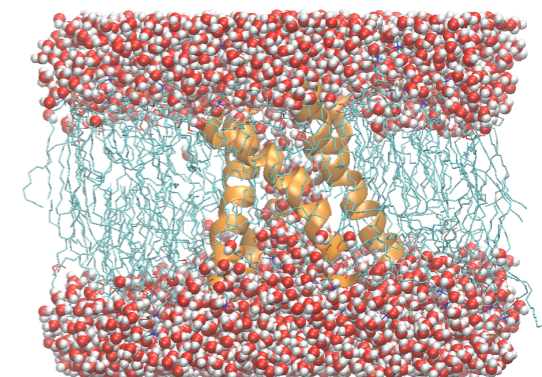
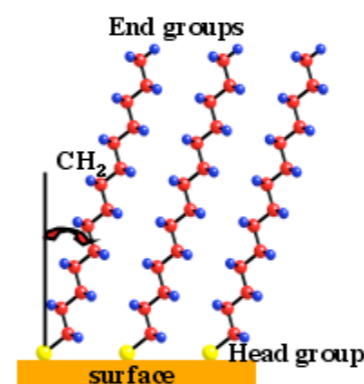
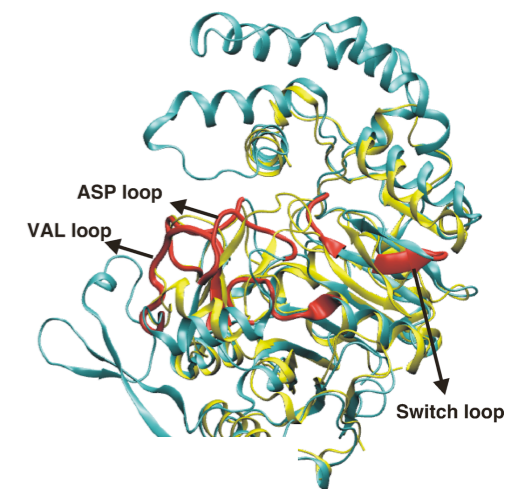
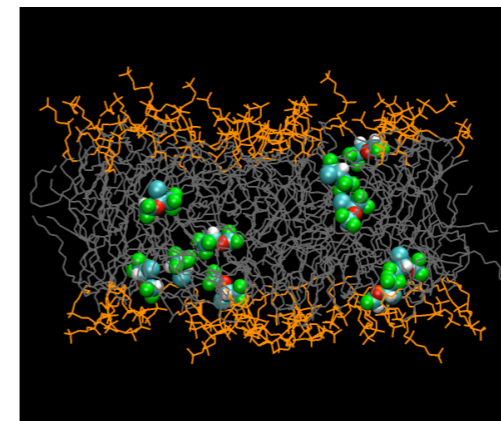
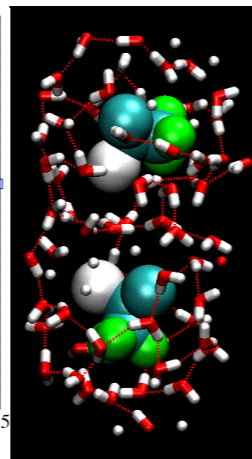
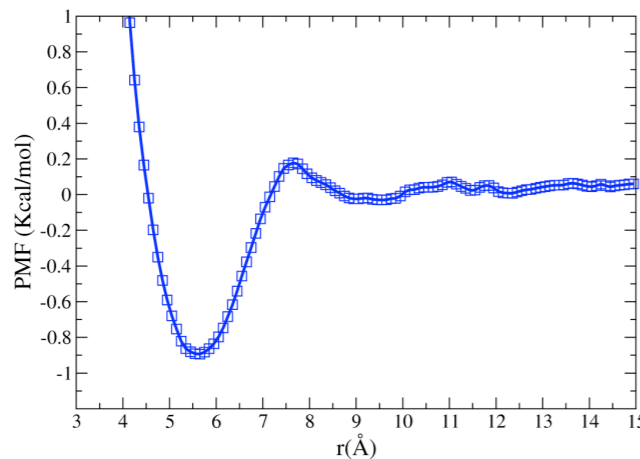
- ▶ Self assembled monolayers
- ▶ Polyelectrolytes

## Proteins:

- ▶ Ion channels
- ▶ Thermal stability
- ▶ Protein-drug interactions

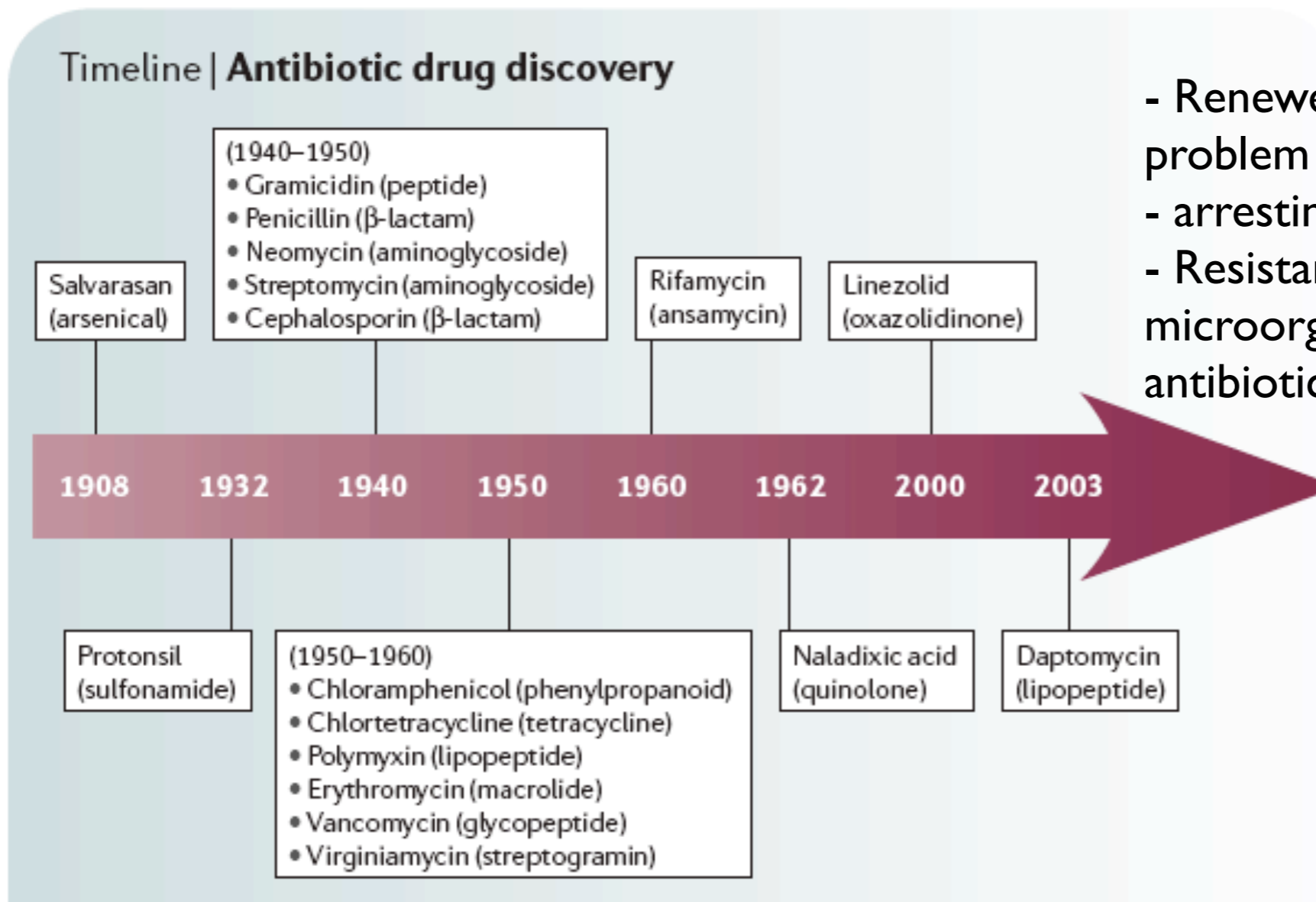
## Lipid Membranes:

- ▶ Small molecule transport
- ▶ Antimicrobial peptides/polymers
- ▶ Rafts



- Antibacterial mechanism
- All Atom simulations
- Coarse graining attempts

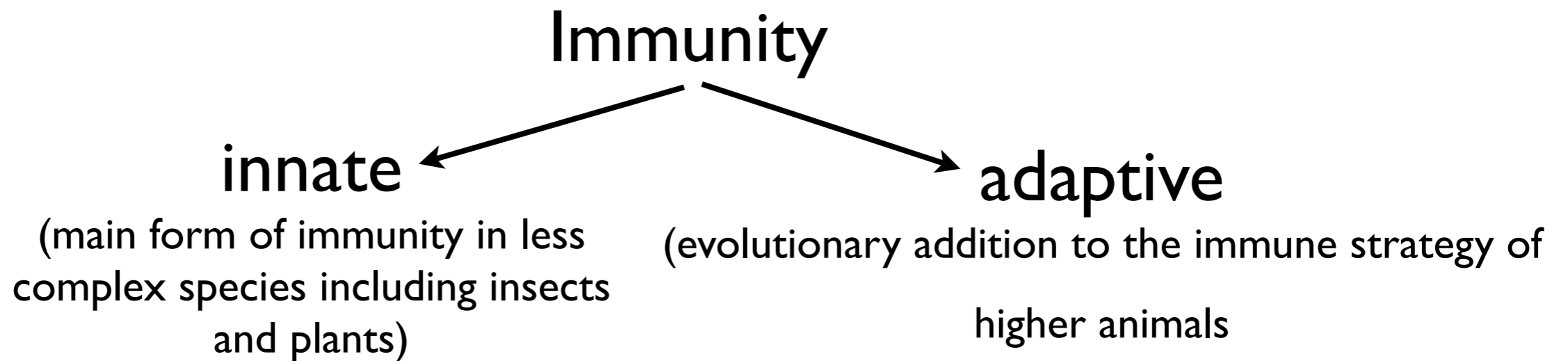
# Antibiotics



- Renewed interest in antibiotics due to the problem of growing antibiotic resistance
- arresting cell growth
- Resistance is the continuous growth of microorganisms in the presence of antibiotics

antibiotics put selective pressure on microorganisms leading to “superbugs”

# Innate Immunity



**Innate immunity:** highly effective set of mechanisms used by multicellular organisms to recognize and counter the constant threat of microbial infections

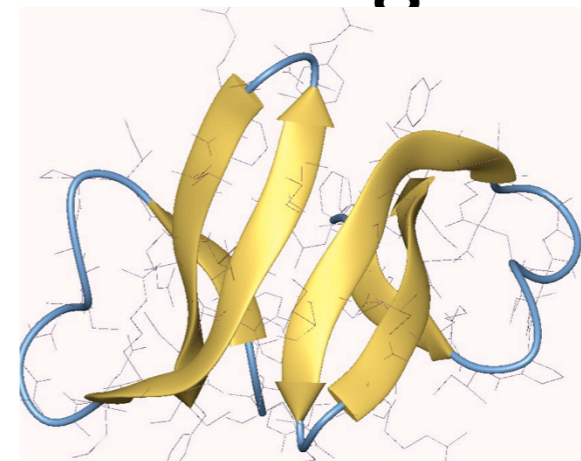
**cationic host defense peptides:**

- stimulated by bacterial signalling molecules
- an element of innate immunity in controlling infections

**LL-37 (14-18 residues)**

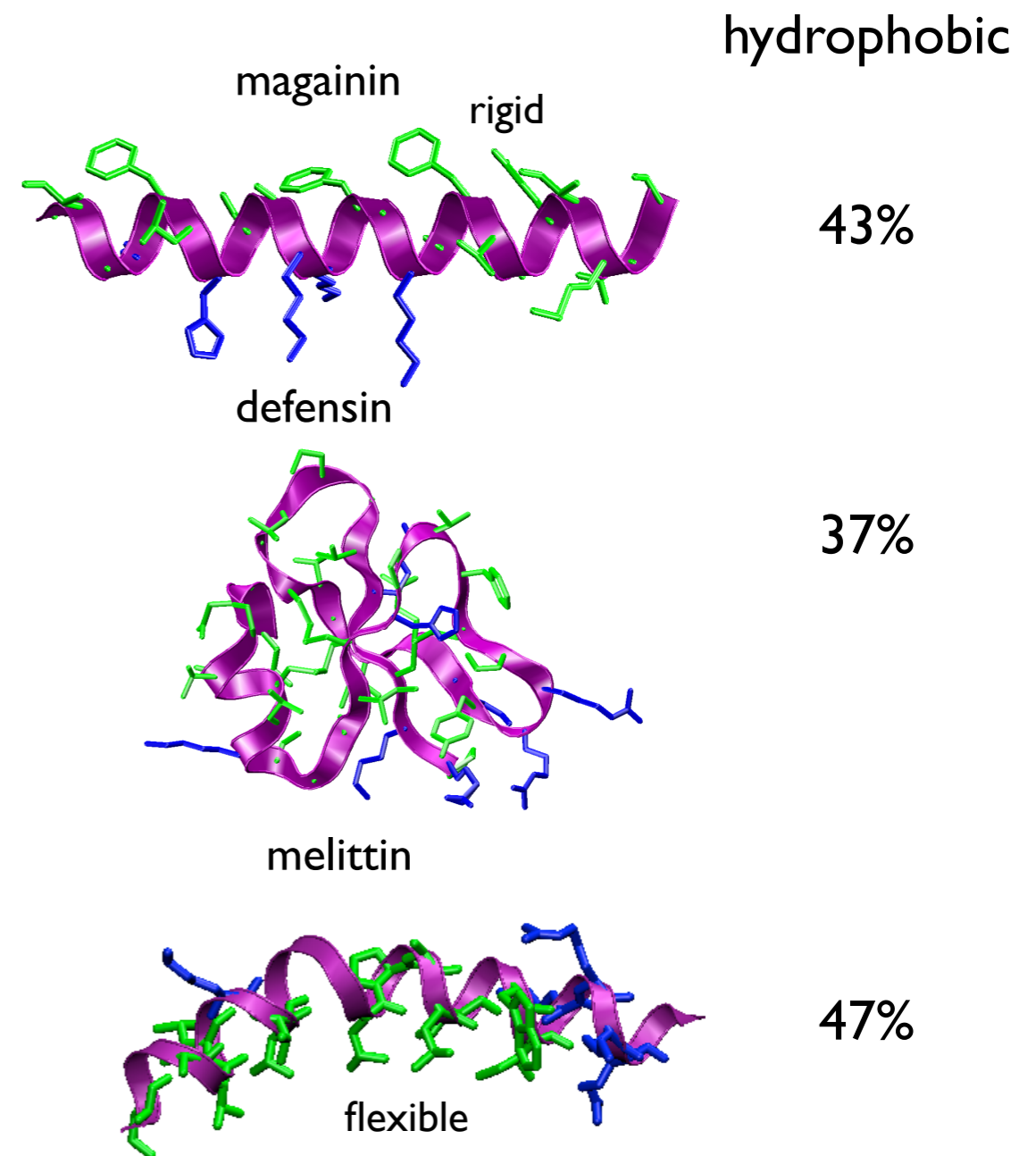
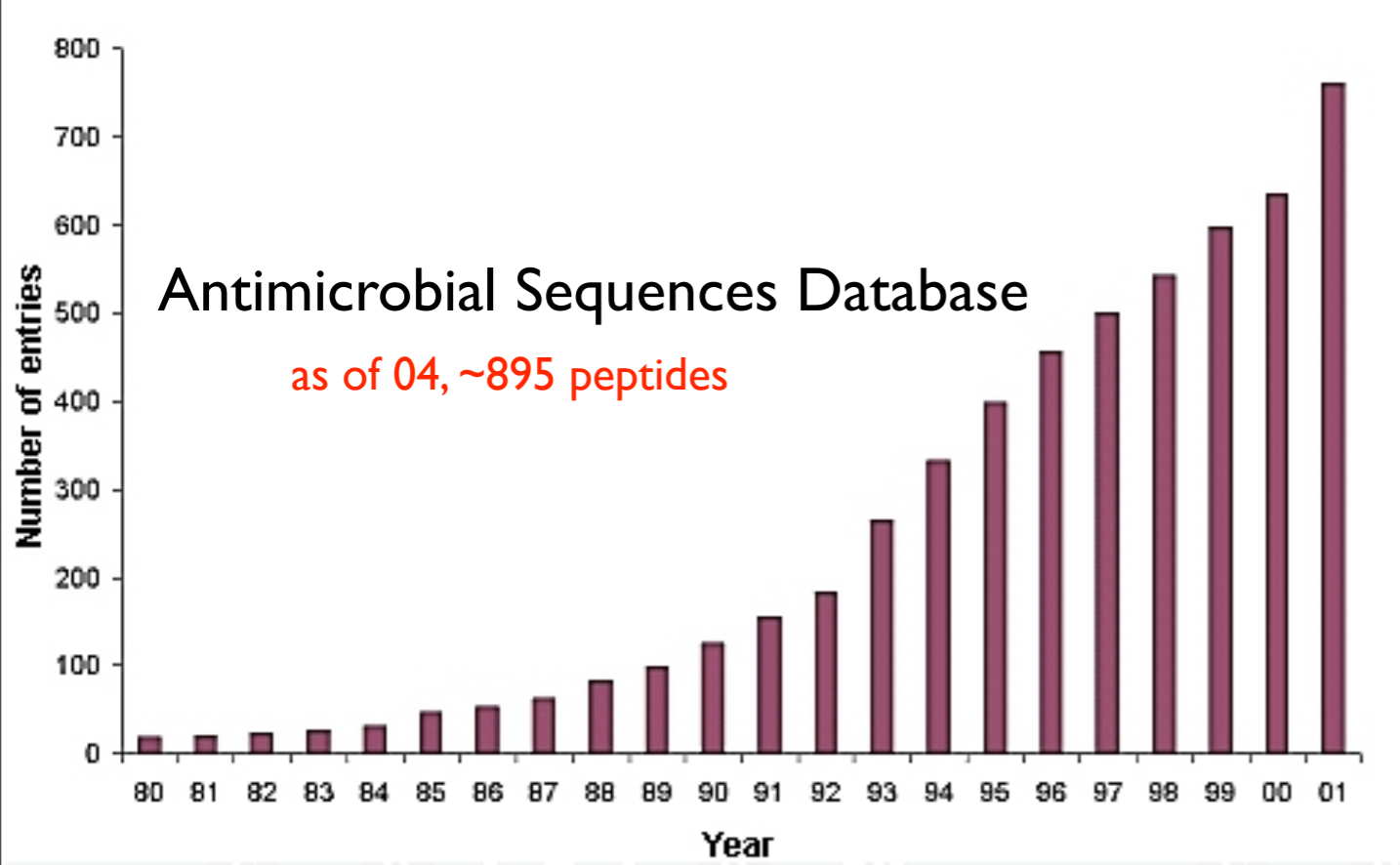


**defensin (28-42 residues)**



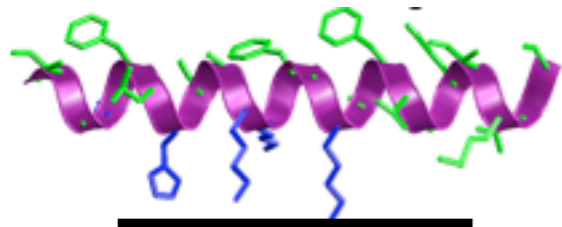
# Natural antimicrobial peptides

- size varies from 6 -60 amino acids
- come in both anionic and cationic forms
- existence of spatial separation of polar and hydrophobic residues (amphipathicity)
- disruption of membrane: mode of action on bacteria
- ratios of hydrophobic to charged residues can vary from 1:1 to 2:1
- part of innate immune system (first line of defense)



# Antimicrobial mechanism

Antimicrobial peptide



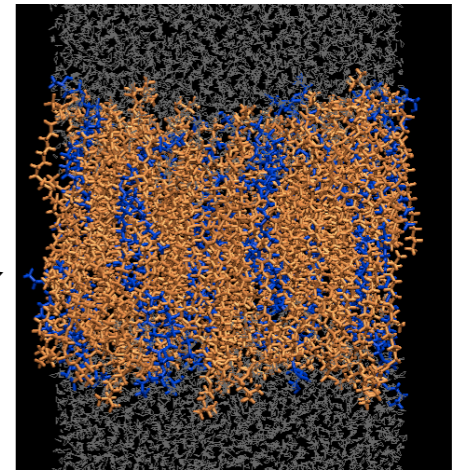
charged

hydrophobic

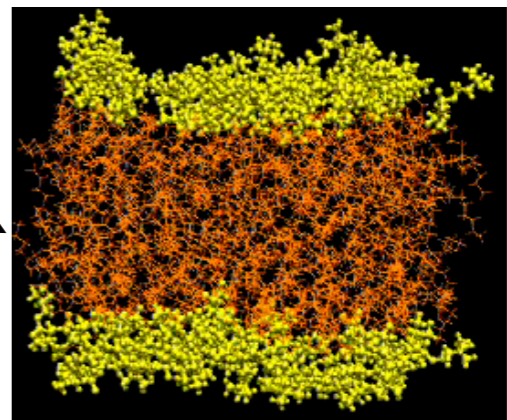
+

Selectivity

Disruption



bacterial membrane



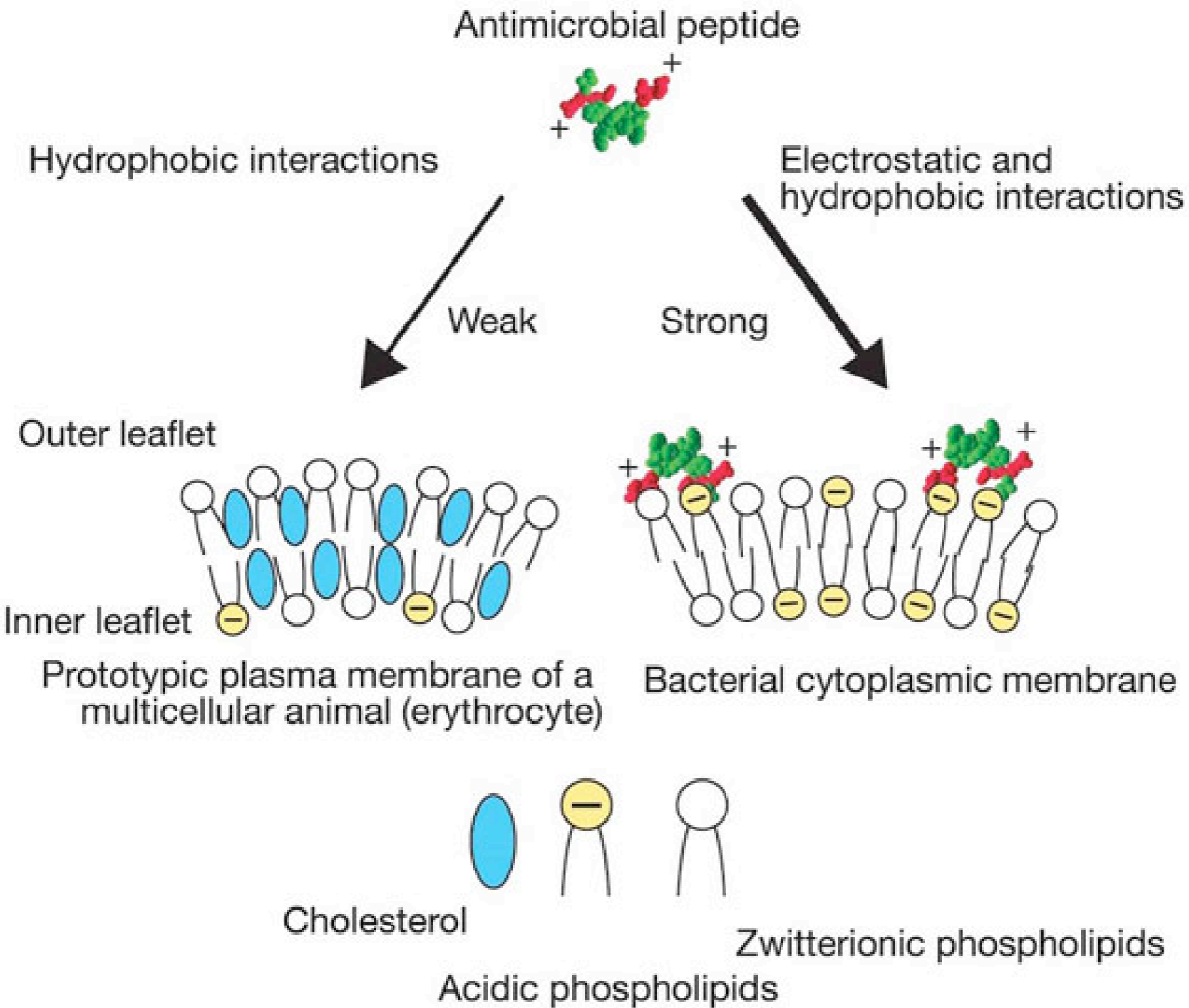
plasma membrane

Barrel-stave

Carpet

Toroidal

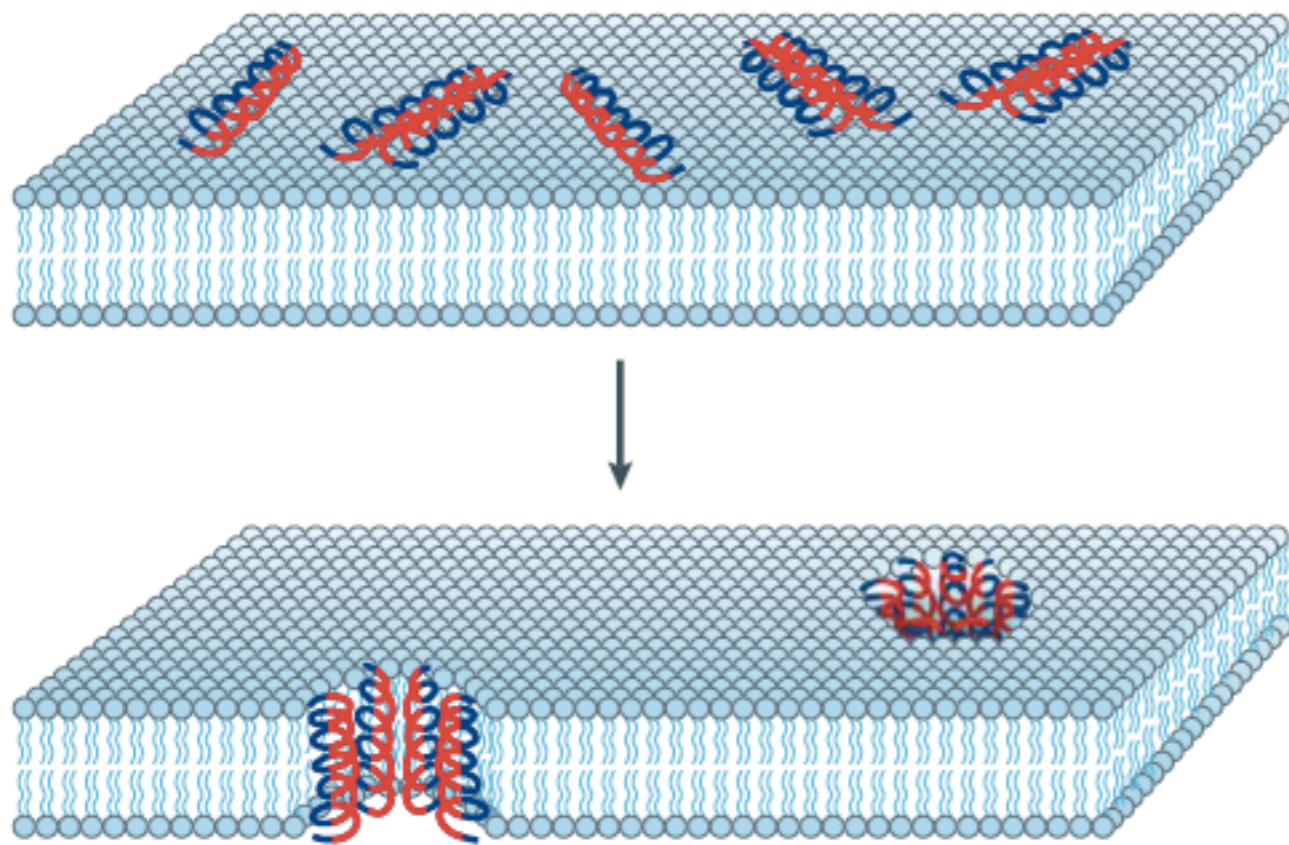
# Antimicrobial mechanism: Selectivity



Bacterial membranes have 20-30% negatively charged lipids



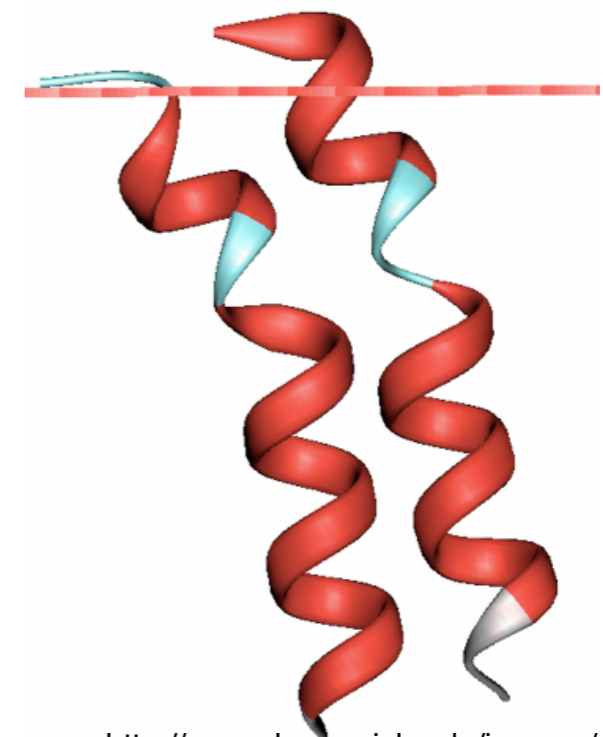
# Disruption Mechanisms: Barrel Stave model



- Peptides aggregate
- hydrophobic parts (blue) aligns with the lipid core region
- hydrophilic parts (red) forms the interior of the pore

## Example: Alamethicin

- adopts helical configurations
- 3-11 helices
- inner diameter  $\sim 1.8\text{nm}$
- changes in lipid composition can affect the number of peptides in the aggregate



<http://opm.phar.umich.edu/images/proteins/1amt.gif>

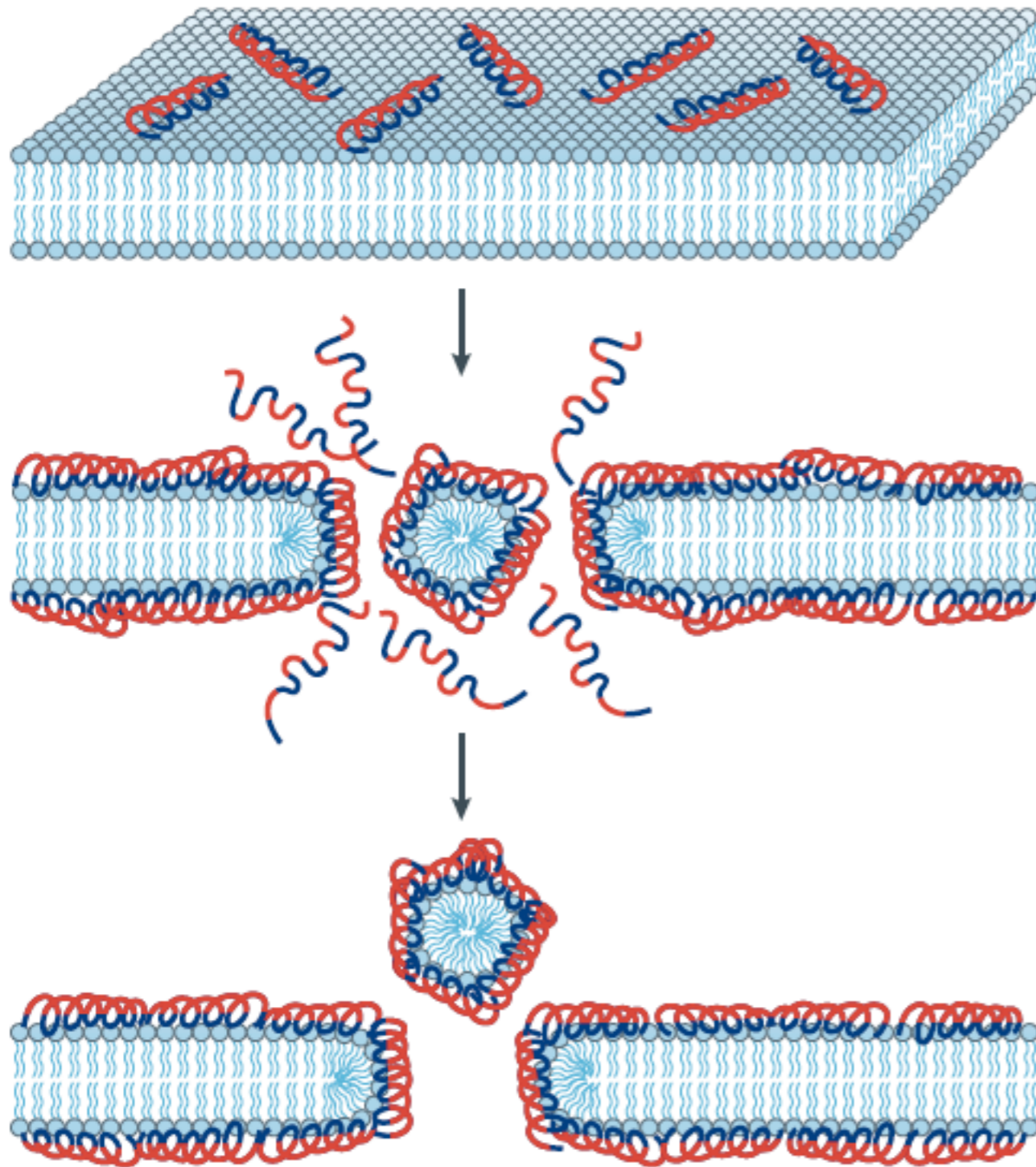
Rocca et al, BBA, 1999

Yang et al, Biophys. J, 2002

Zaslhoff, nature, 2002

Brodgen, Nature Reviews Microbiology, 2005

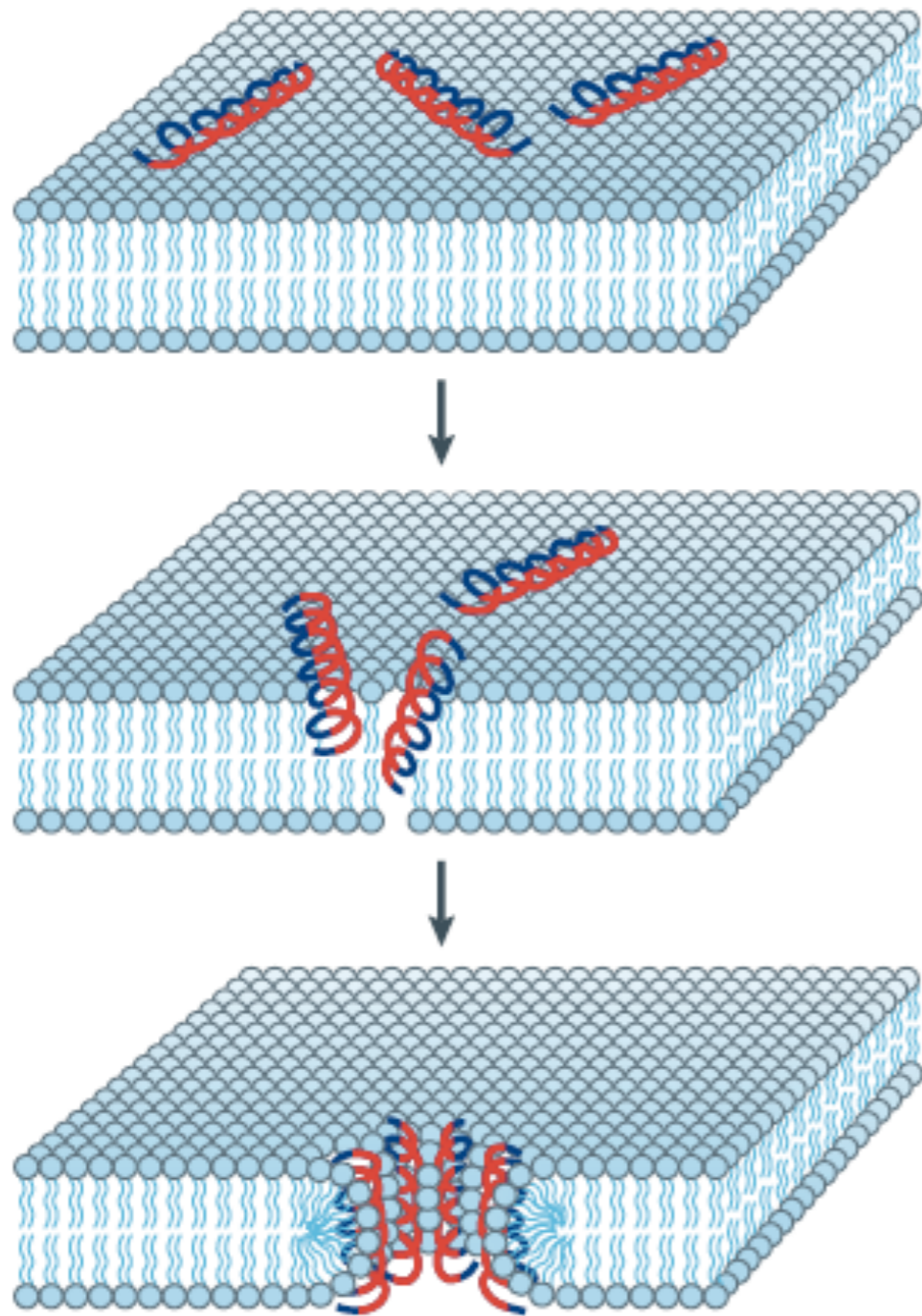
# Disruption Mechanisms: Carpet model



- Peptides accumulate parallel to the bilayer
- electrostatic interactions with anionic sites of the bilayer
- at high peptide concentration, possible formation of micelles leading to lysis
- possible transient toroid like holes for further access

Examples: Ovispirin, cecropin

# Disruption Mechanisms: Toroidal model



- attached peptides aggregate and induce lipids to bend continuously
- water core is lined with both hydrophilic parts of the peptide and lipid head groups
- peptides are always associated with lipid head groups (unlike barrel-stave model)

Examples: magainin, melittin

magainin induced toroidal pore have inner diameter of ~3-5 nm and each pore has 4-7 magainin monomers and ~90 lipid molecules

# Biomimetic polymers: desired properties

MIC: minimum inhibitory concentration

MIC was defined as the lowest polymer concentration to completely inhibit bacterial growth **lower values preferred**

HC<sub>50</sub>: HC<sub>50</sub> is the polymer concentration to achieve 50% hemolysis **higher values preferred**

ment of peptidic analogues of AMPs as intravenously administered antibiotics.<sup>1</sup> Experimental studies *in vitro* have shown that bacteria cannot easily develop resistance to this class of compounds,<sup>67</sup> suggesting they might not suffer from some of the same problems of resistance encountered in other classes of antibiotics with more specific targets. Indeed, a compound designed by these principles successfully completed the first phase of human clinical trials as an iv antibiotic for treatment of multidrug-resistant *S. aureus*.<sup>96</sup>

TABLE 1. Conformational Tuning of Properties of Antimicrobial Oligomers<sup>a</sup>

Compound	Structure	MIC (μM)		
		<i>S. aureus</i> <sup>b</sup>	<i>E. Coli</i> D31	HC <sub>50</sub> (μM)
2		50	12.5	14
3		7.8	3.9	445
4		0.80	0.80	18
5		0.87	0.87	145
6		0.66	5.3	593
7		0.70 <sup>c</sup>	0.70	5
8		0.2	2.8	63-90
9		0.20	0.1	440
10		0.20	0.10	440

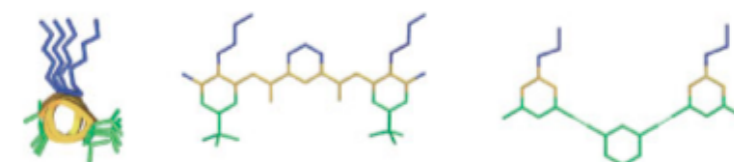


FIGURE 2 Facially amphiphilic structures of the SMAMPs;  $\beta$ -peptide 1 (left), pyrimidine arylamide oligomer 2 (middle), and phenylene ethynylene oligomer 3 (right).

## Synthetic Mimics of Antimicrobial Peptides

Abhigyan Som<sup>1</sup>, Satyavani Vemparala<sup>2</sup>, Ivaylo Ivanov<sup>3,4</sup>, Gregory N. Tew<sup>1</sup>

PeptideScience Volume 90 / Number 2 2008

SMAMP	MIC (Gram-positive) (μg/mL)	MIC (Gram-negative) (μg/mL)	HC <sub>50</sub> (μg/mL)	Selectivity <sup>a</sup> (HC <sub>50</sub> /MIC)
1 <sup>b</sup>	—	9.0	135	15
1 <sup>c</sup>	—	26.0	4680	180
2	0.8	0.8	14	17.5
3	0.2	0.1	88	880
MSI-78 <sup>d</sup>	12.0	12.0	120	10

ACCOUNTS  
of chemical research

De Novo Design of Antimicrobial Polymers, Foldamers, and Small Molecules: From Discovery to Practical Applications

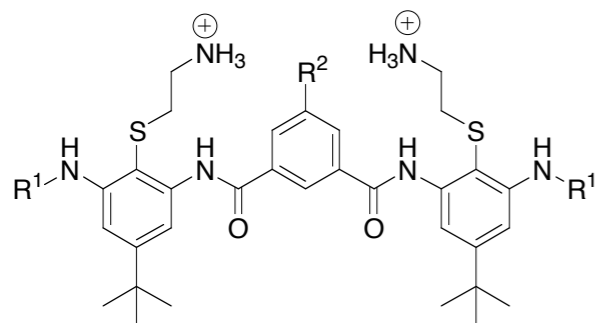
GREGORY N. TEW,<sup>†</sup> RICHARD W. SCOTT,<sup>‡</sup> MICHAEL L. KLEIN,  
AND WILLIAM F. DEGRADO\*

2009

# Biomimetic polymers

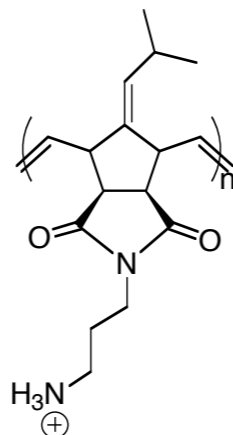
## Facial amphiphilic

- Aryl amides:



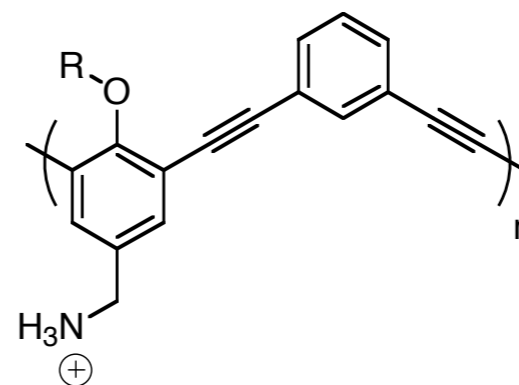
- Tew, G. N., Liu D., et al. *PNAS*, **2002**, 99, 5110
- Liu, D., et al. *Angew. Chem. Int. Ed.* **2004**, 43, 1158

- Polynorbornenes



- Ilker, M. F., et al. *J. Am. Chem. Soc.* **2004**, 126, 15870

- Poly(*m*-phenylene-ethynylene)

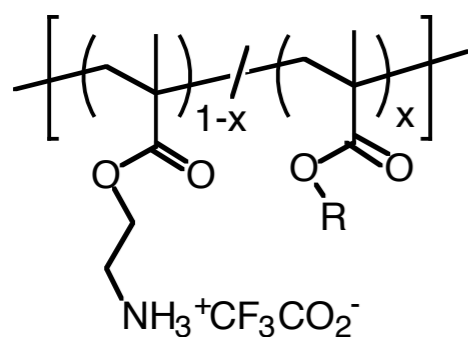


- Arnt, L., et al. *Polym. Sci., Part A: Polym. Chem.* **2004**, 42, 3860.
- Arnt, L.; Tew, G. N. *JACS*, **2002**, 124, 7664

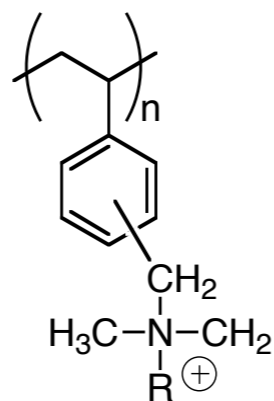
## properties

- Rigid backbones
- Synthetically difficult to make
- Non-toxic to human cells

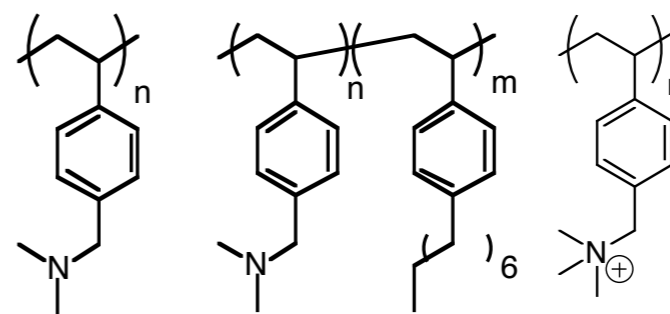
## Flexible amphiphilic



- Kuroda, K, DeGrado, W.F *JACS* **2005**, 127 4128.



- Ikeda, T., Tazuke, S., *Makromol. Chem. Rapid Commun.* **1983**, 4, 459.



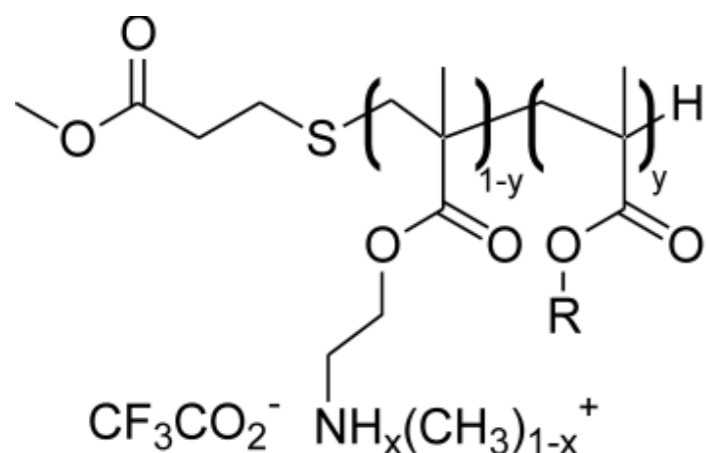
- Gelman, M. A.; Weisblum, B., Lynn, D. M.; Gellman, S. H. *Org. Lett.* **2004**, 6, 557.

- Flexible backbones
- Facile preparation
- Toxic to human cells

# Biomimetic polymers: systems of interest

## Design paradigms:

- amphiphilicity achieved through introduction of charged & hydrophobic groups
- rigid backbones allow pre-built facial amphiphilicity
- highly repetitive structures are thought beneficial and enhance activity
- aim is to get **high selectivity** and **low toxicity**



## Amphiphilic Polymethacrylate Derivatives as Antimicrobial Agents

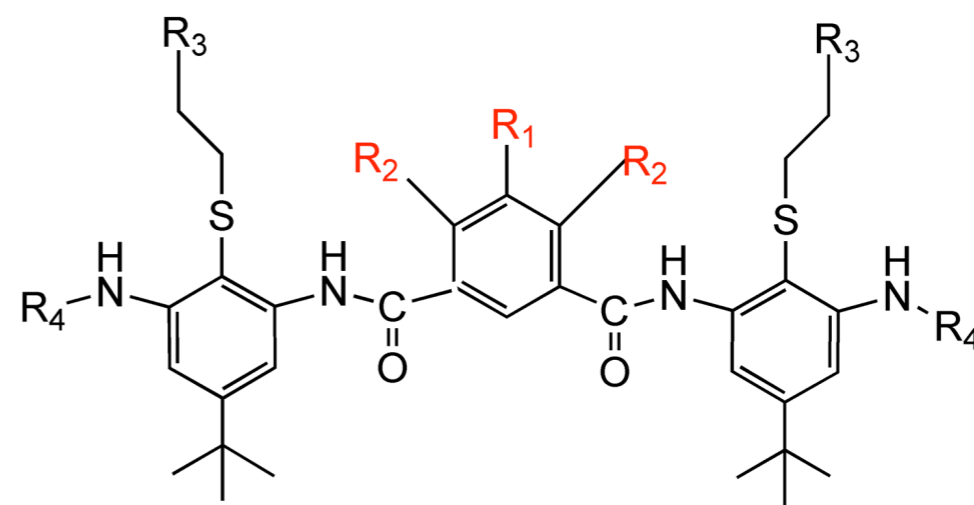
Kenichi Kuroda and William F. DeGrado\*

4128 ■ J. AM. CHEM. SOC. 2005, 127, 4128–4129

## AIMS:

- is 'inbuilt amphiphilicity' a requirement for antimicrobial action?
- understand the interplay between hydrophilic & charged groups with respect to antimicrobial action

- understand aggregation and disruption mechanism
- phase space of charge & hydrophobicity



## De novo design of biomimetic antimicrobial polymers

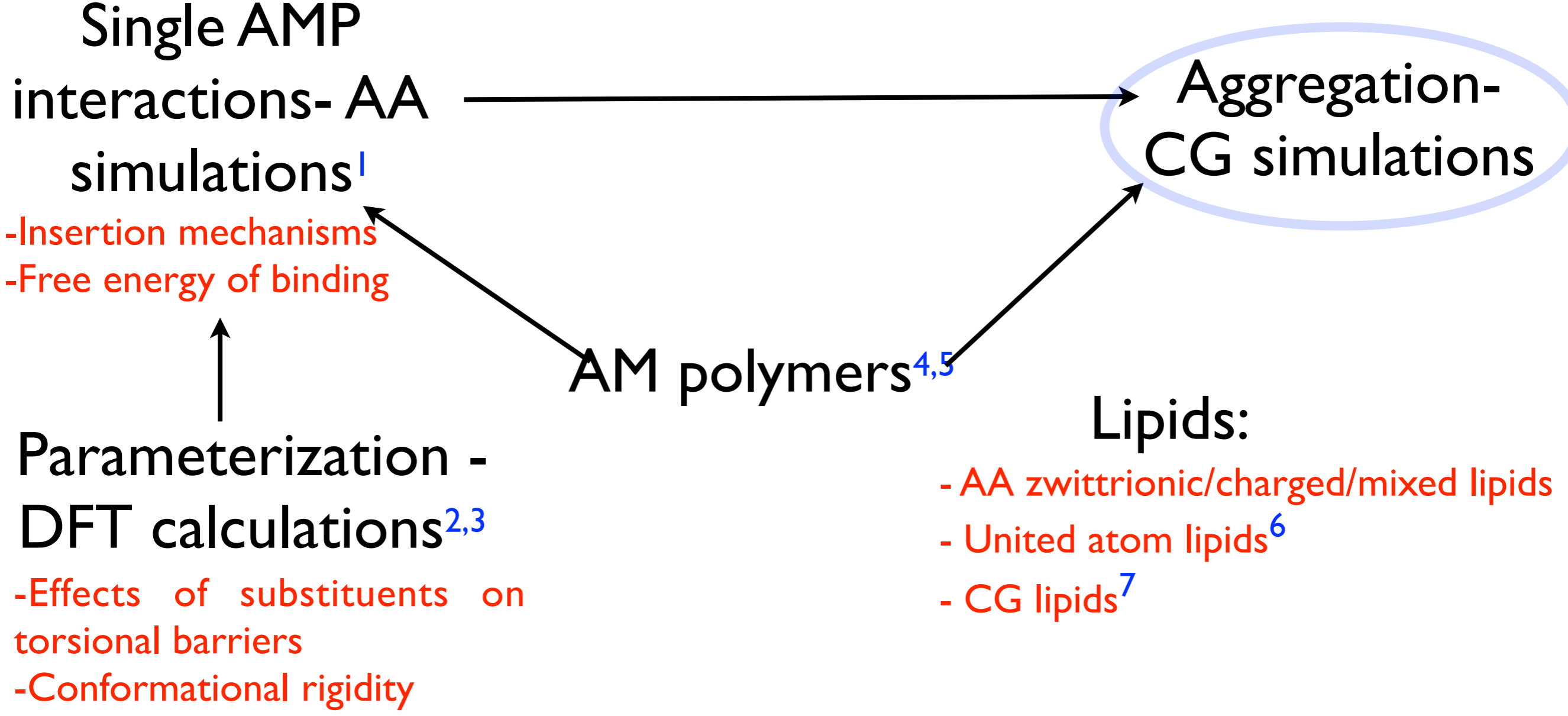
Gregory N. Tew<sup>\*,†</sup>, Dahui Liu<sup>\*,†</sup>, Bin Chen<sup>‡</sup>, Robert J. Doerksen<sup>‡</sup>, Justin Kaplan<sup>\*</sup>, Patrick J. Carroll<sup>¶</sup>, Michael L. Klein<sup>‡</sup>, and William F. DeGrado<sup>\*,†</sup>

5110–5114 | PNAS | April 16, 2002 | vol. 99 | no. 8

## AIMS:

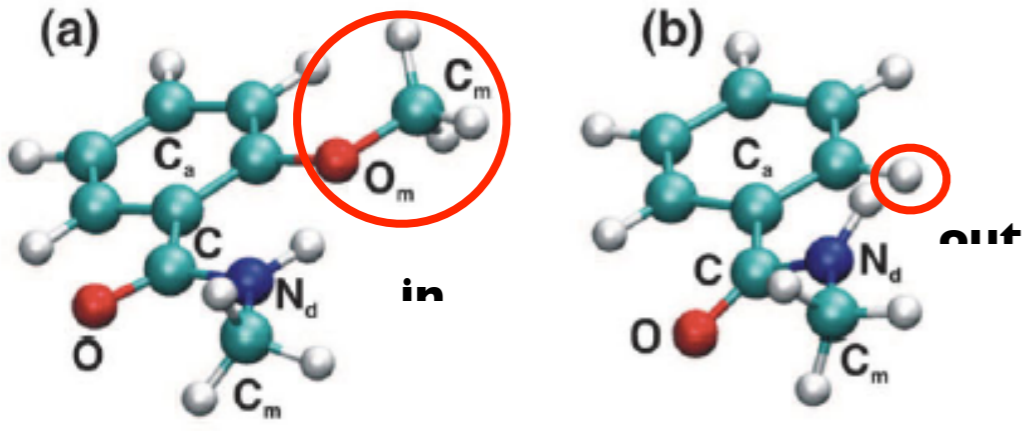
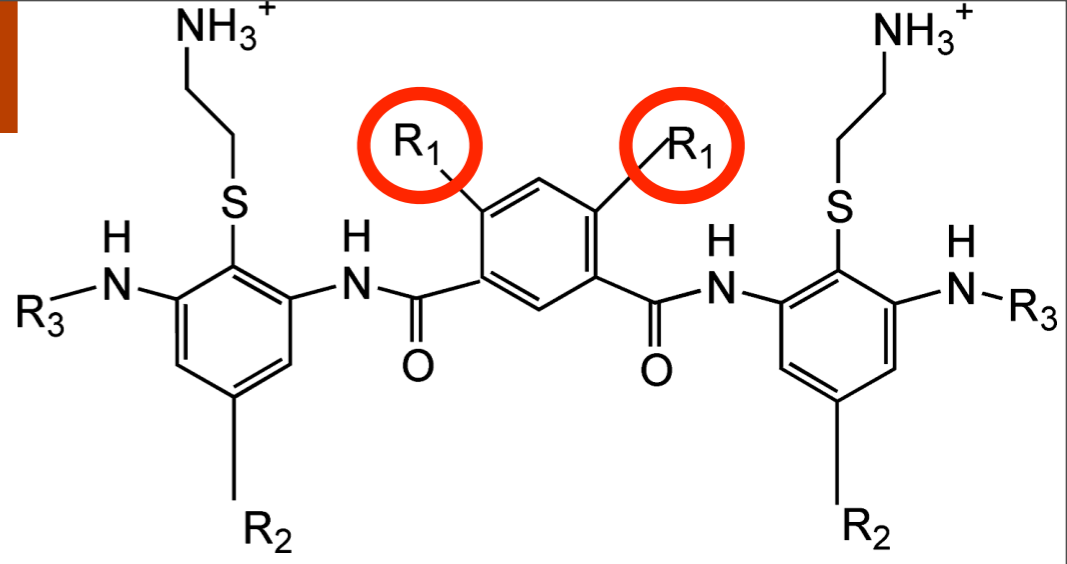
- calculate appropriate backbone torsion potentials
- effect of substitutions (like R<sup>1</sup> & R<sup>2</sup>) on the 'flexibility' of the molecule
- study interactions of arylamides with both zwitterionic & charged lipids

# Computational approach

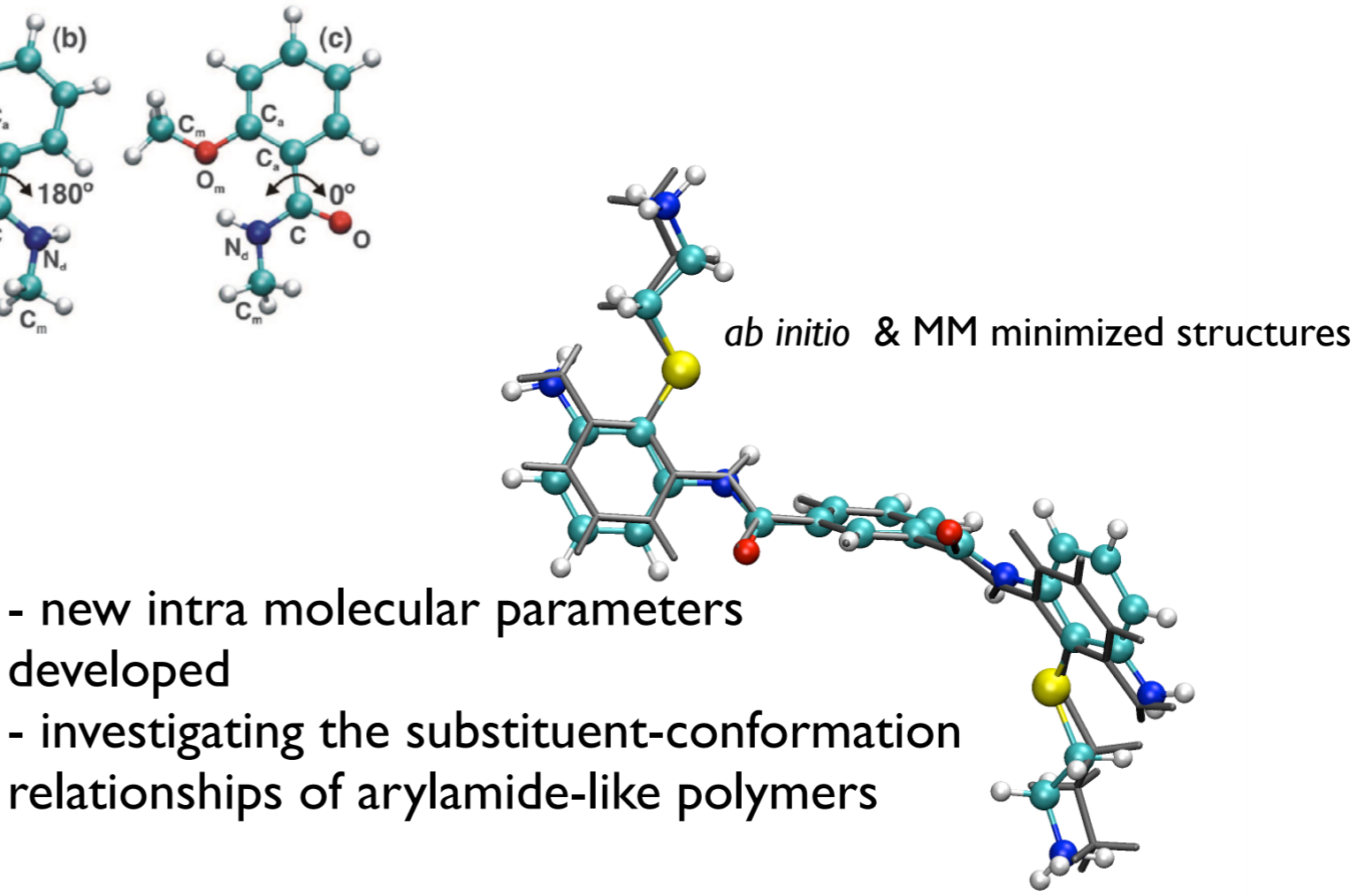
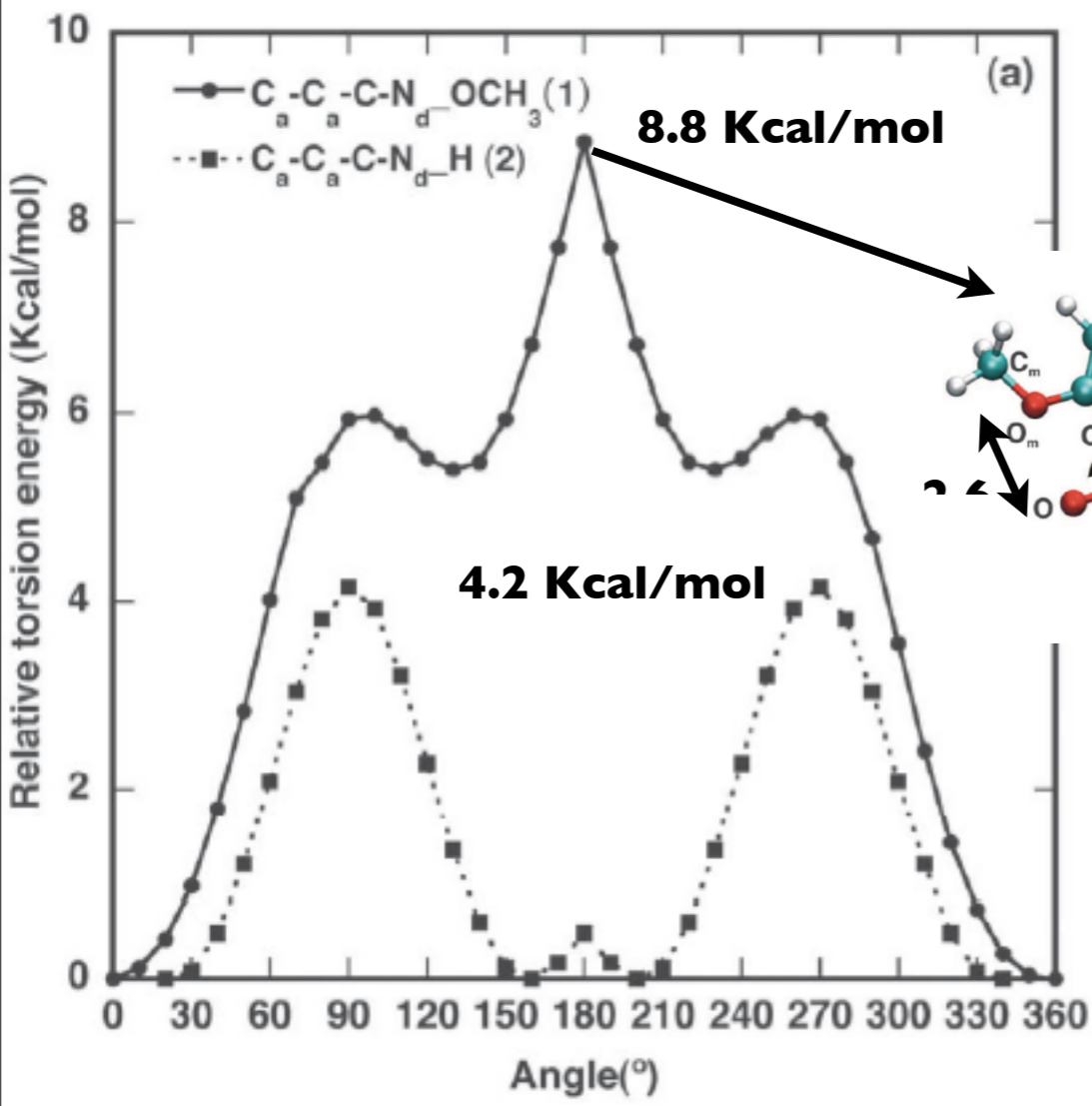


1. JACS, 2006, 128, 1778-1779  
2. J Comp Chem, 2006, 27, 693-700  
3. J Phys Chem B, 2006, 110, 3517-3526  
4. Peptide Science (Review), 2008, 90, 83-93  
5. Acc. Chemical Res, 2009, ASAP article

# Arylamide polymers: Parameters



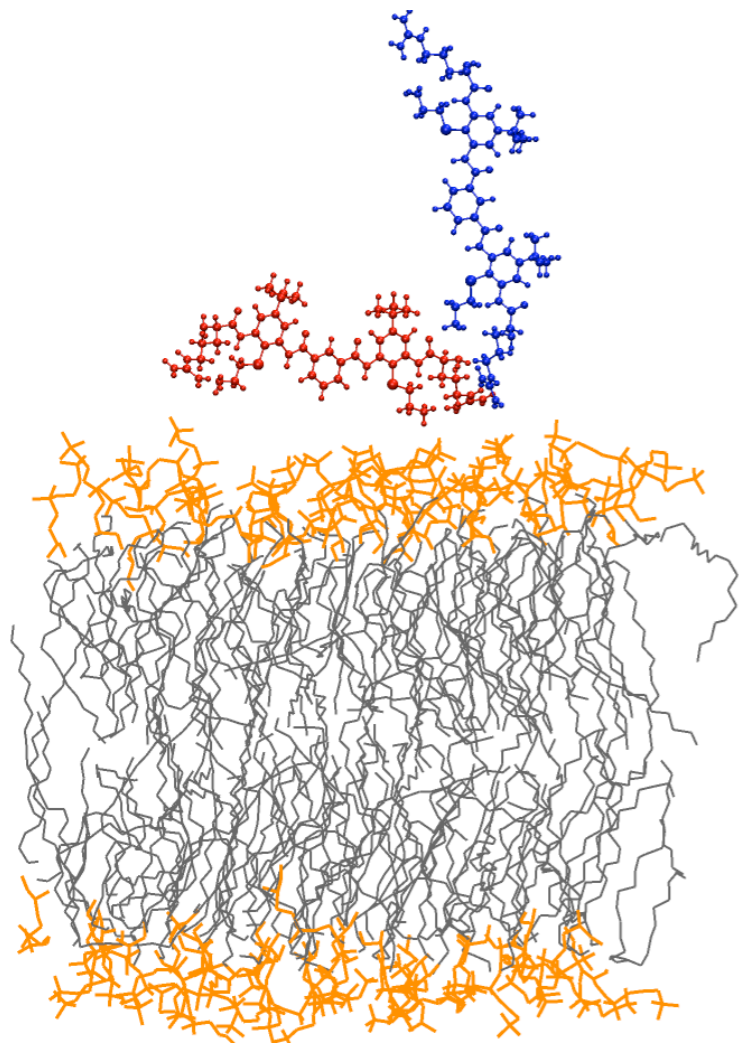
- Effect of methoxy group substitution on backbone flexibility
- intra molecular hydrogen bond (1.91 Å) between amide proton and ether oxygen



*Ab Initio* Calculations of Intramolecular Parameters for a Class of Arylamide Polymers  
 SATYAVANI VEMPARALA,<sup>1</sup> IVAYLO IVANOV,<sup>2</sup> VOJISLAVA POPHRISTIC,<sup>3</sup> KATRIN SPIEGEL,<sup>1</sup> MICHAEL L. KLEIN<sup>1</sup>  
 I. Comp. Chem. 2006.



# Arylamide polymers: MD



Simulation time: 8 ns (each of 4 arylamides)

Lipid Bilayer: DOPC

Temperature: 305K

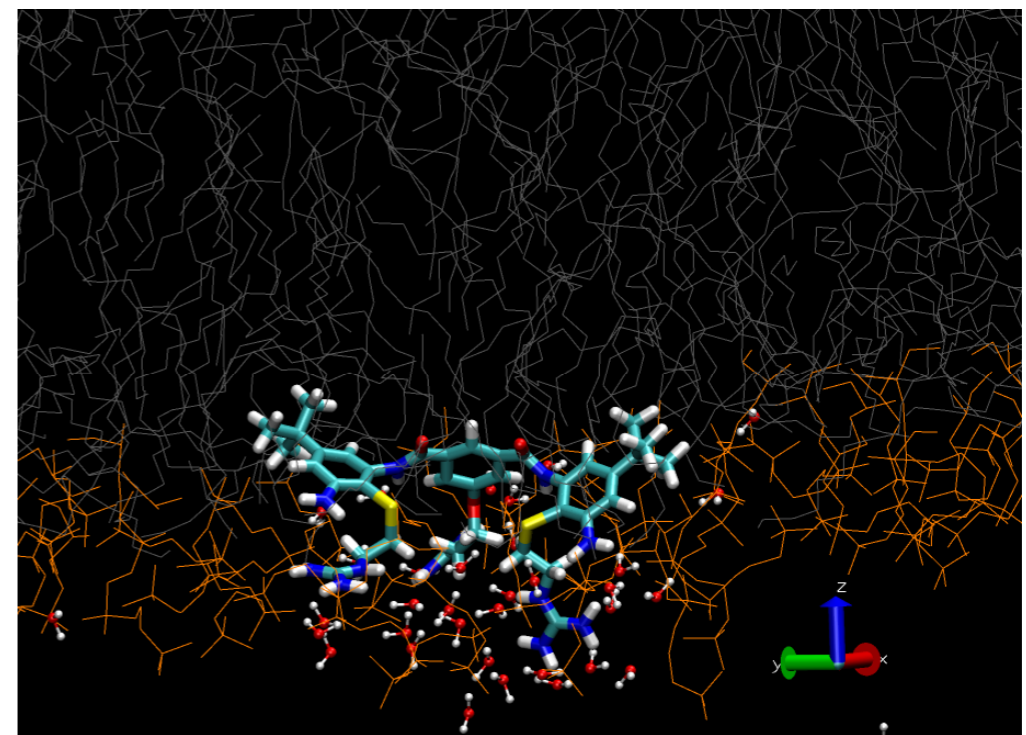
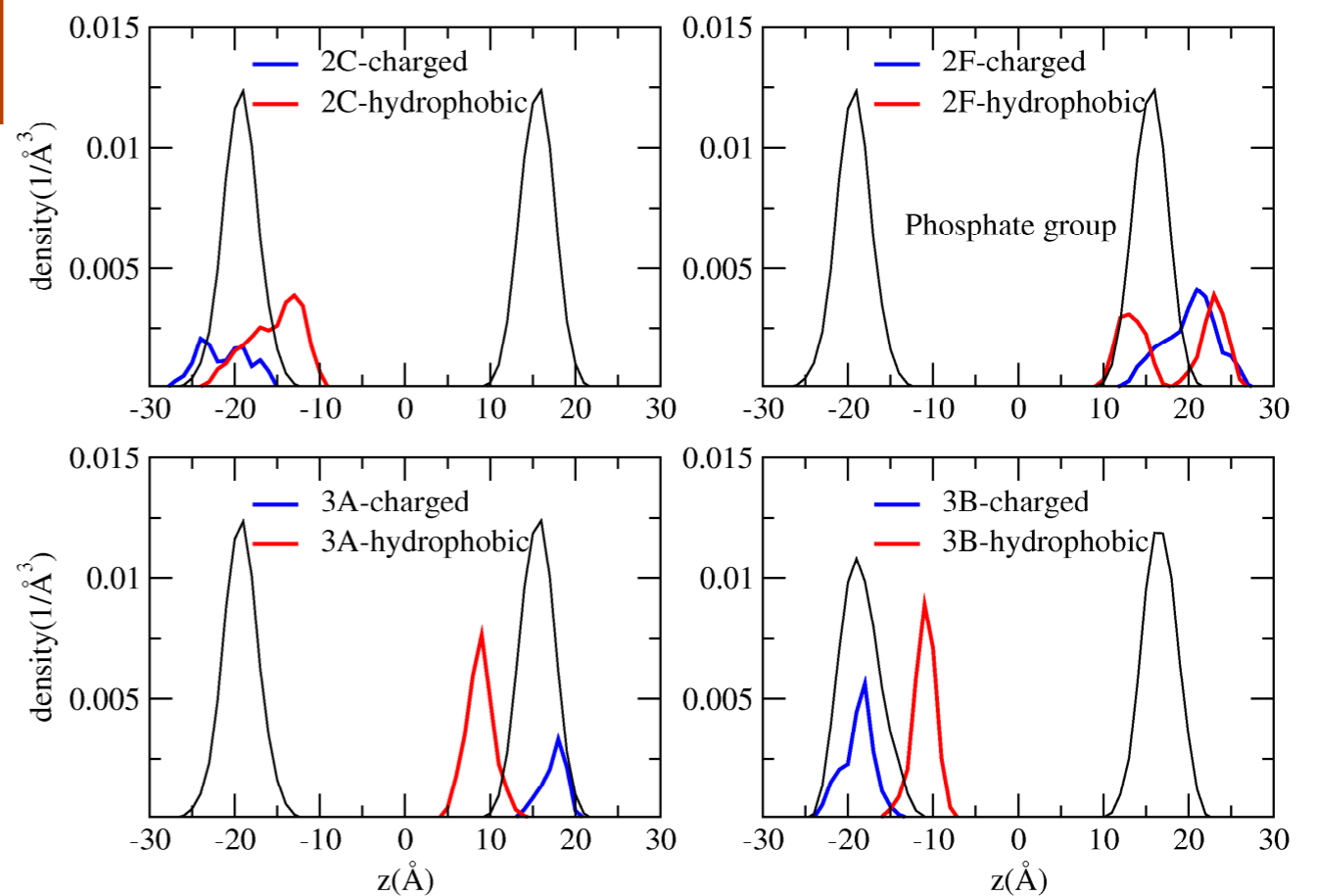
NPT ensemble

PME electrostatics

Systems:

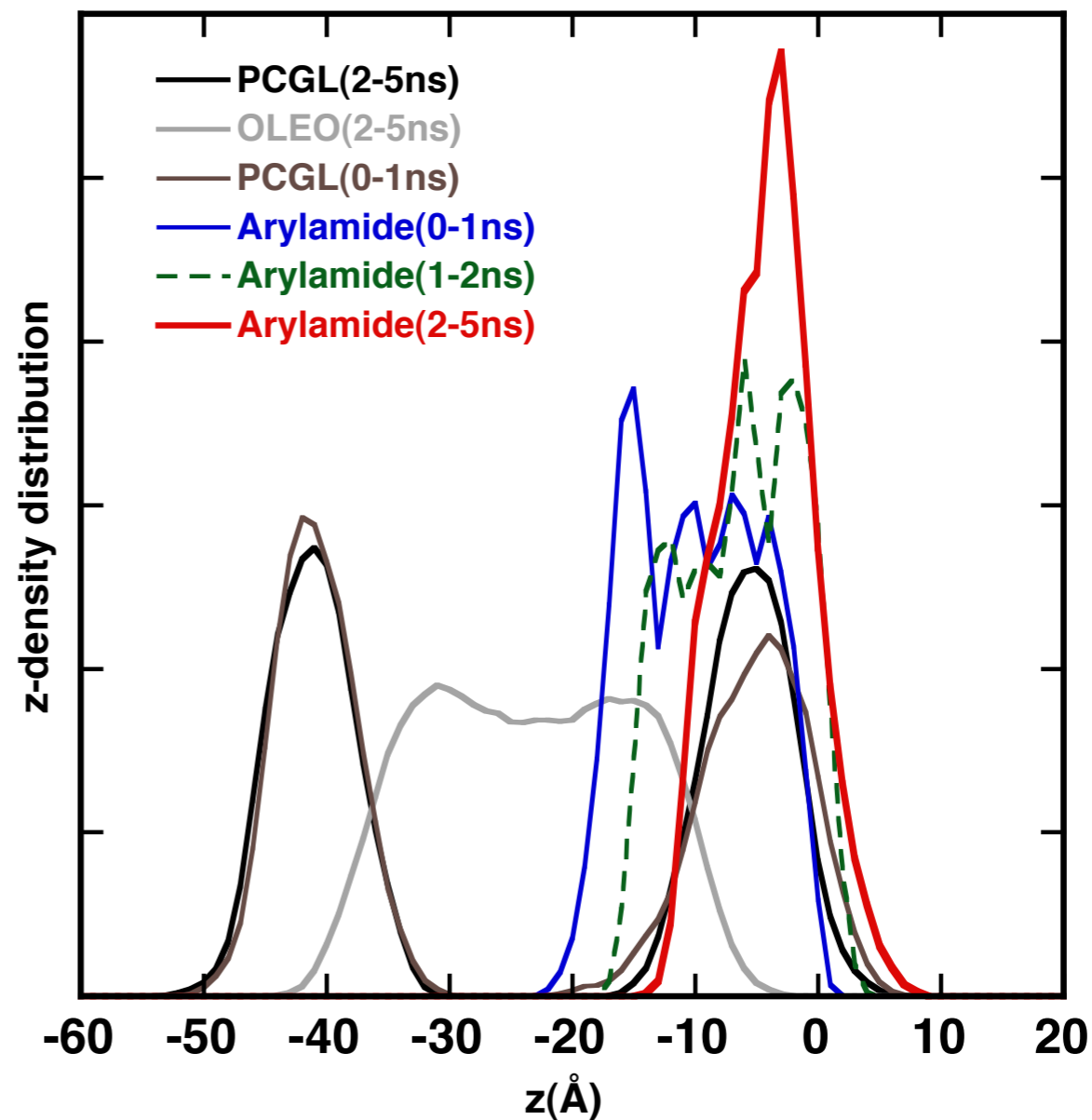
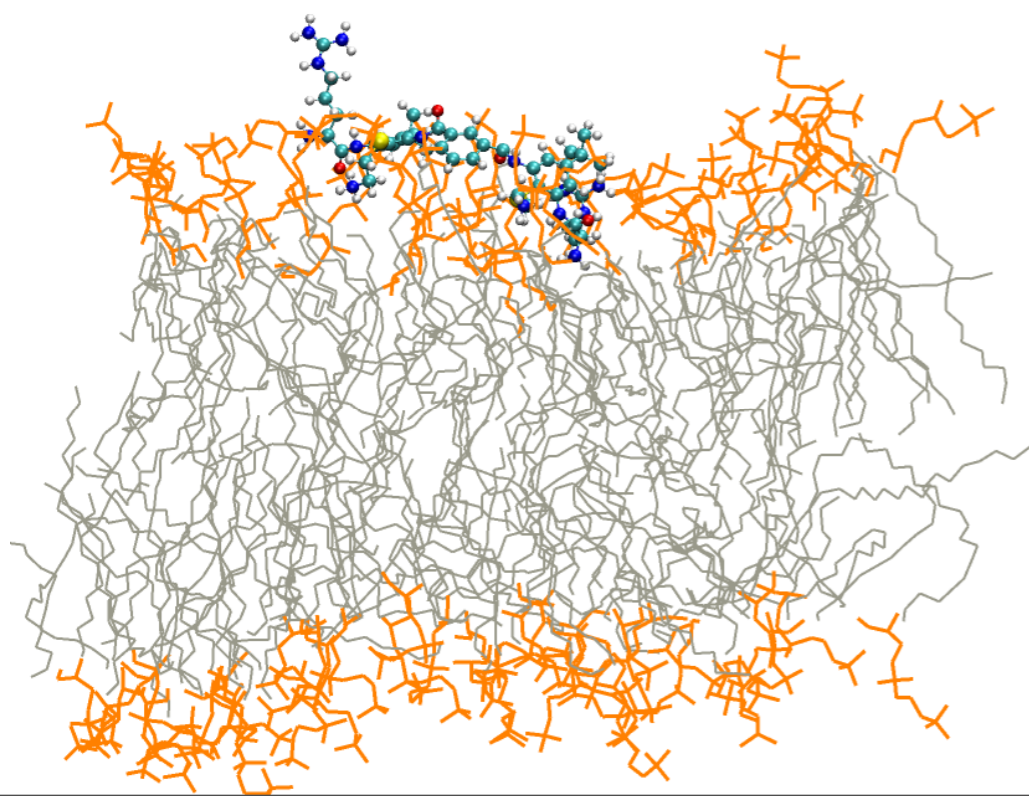
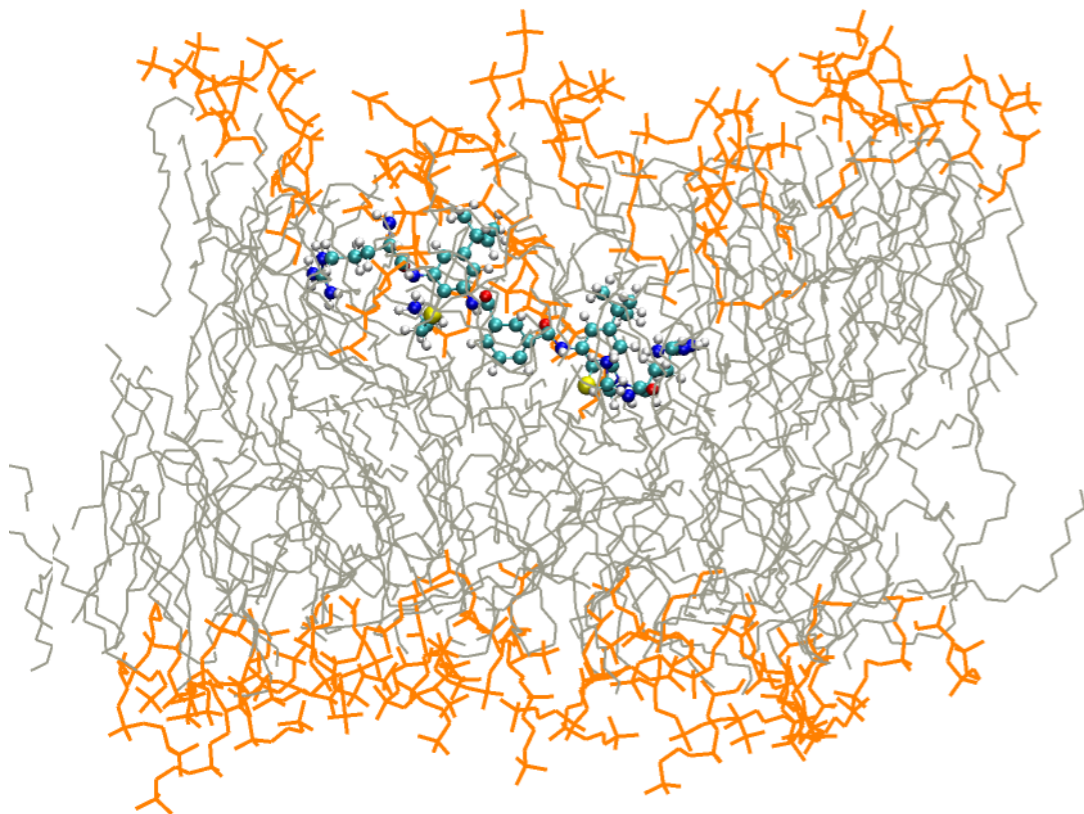
72 lipids

~18,000 atoms



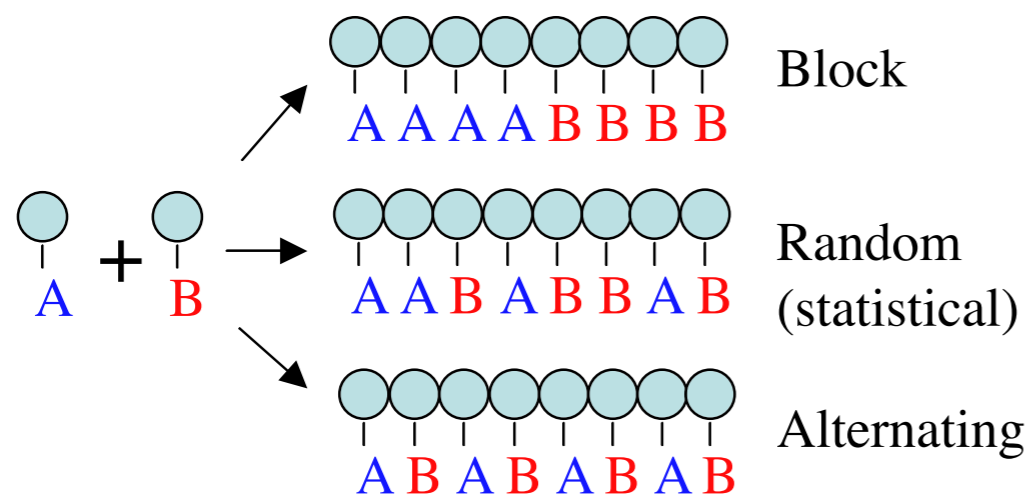
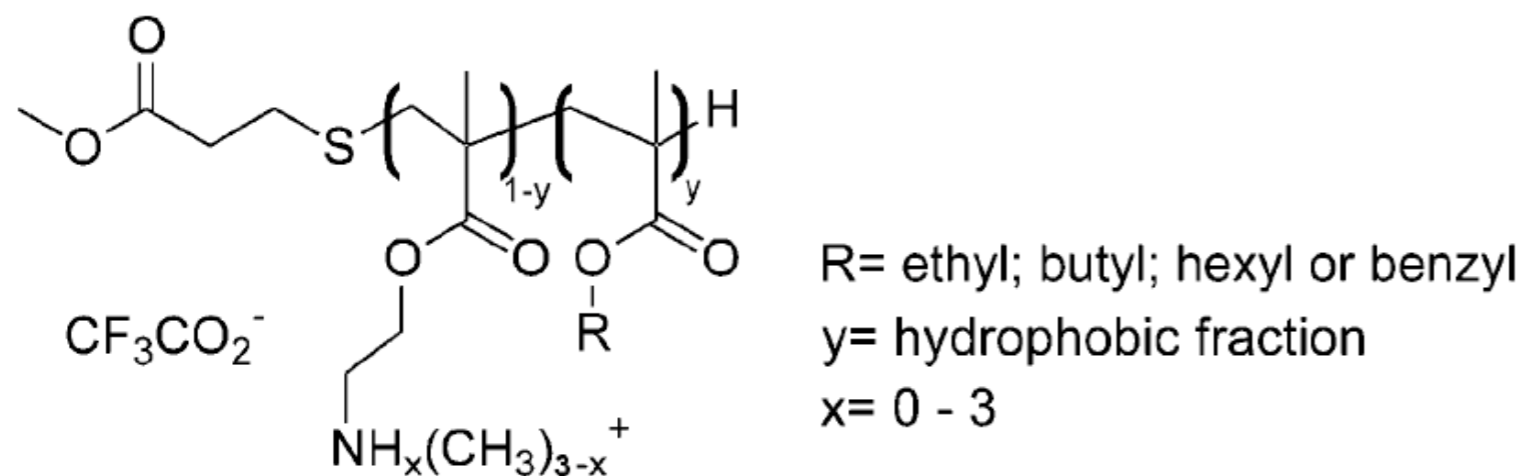
Snorkeling of guanidine/amine side chains enables them to make polar interactions with complex lipid-head-water region. The hydrophobic side chains are located well into the lipid tail region

# Arylamide polymers: MD



A coarse steered molecular dynamics simulation has been done to get different starting points for further MD. Shown in the right is a starting point where the arylamide was just below the head group region. On relaxing and running the system for 5ns more, the most optimal position for the arylamides is in the head group

# System setup: random copolymers



Simulation time: 7.5 - 9.5 ns (each of 20 polymer systems)

Lipid Bilayer: DOPC

Temperature: 303K

NPT ensemble

PME electrostatics

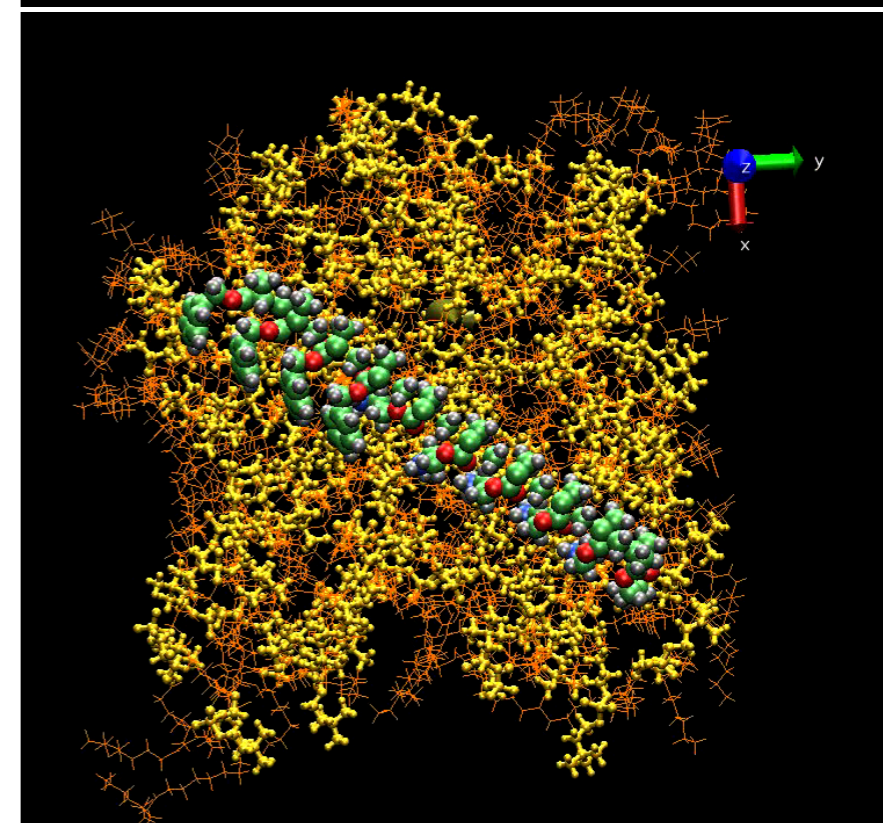
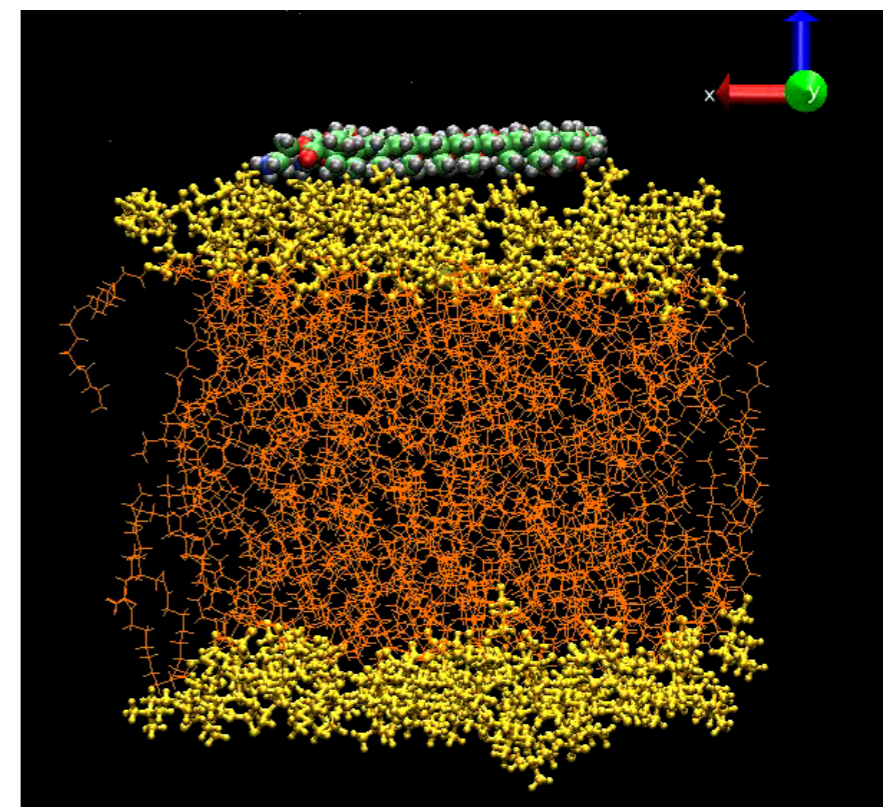
Systems:

72 or 85 lipids

19000 to 24000 atoms

20-12-8-ben-nch0-block

20-12-8-ben-nch0-alt



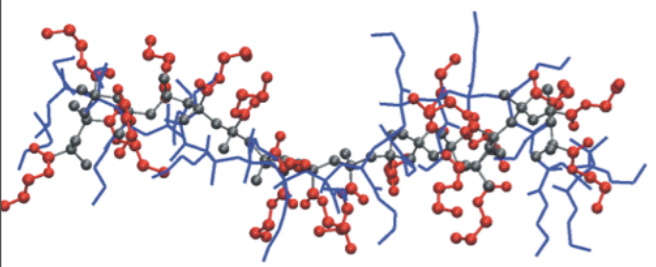
Characterization of Nonbiological Antimicrobial Polymers in Aqueous Solution and at Water–Lipid Interfaces from All-Atom Molecular Dynamics

Ivaylo Ivanov,<sup>\*,†</sup> Satyavani Vemparala,<sup>‡</sup> Vojislava Pophristic,<sup>§</sup> Kenichi Kuroda,<sup>‡,||</sup> William F. DeGrado,<sup>||</sup> J. Andrew McCammon,<sup>†</sup> and Michael L. Klein<sup>‡</sup>

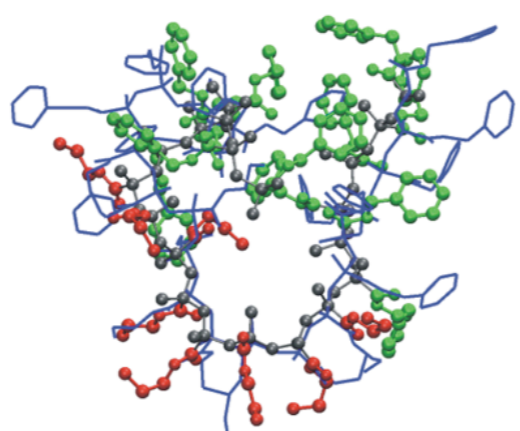
1778 ■ J. AM. CHEM. SOC. 2006, 128, 1778–1779

# Adopted structures

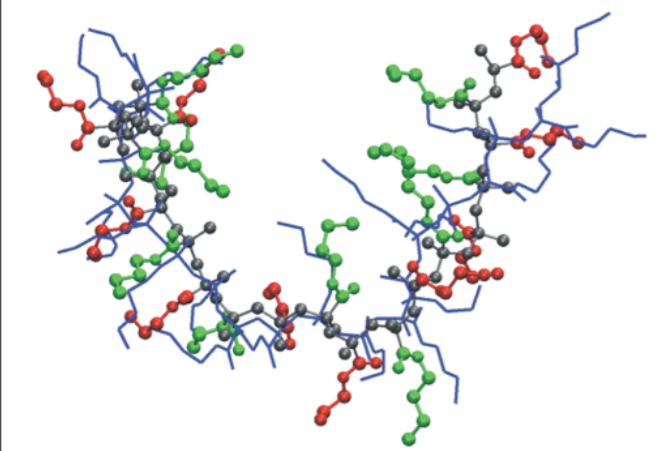
(a) 20-0-20-but-nch0



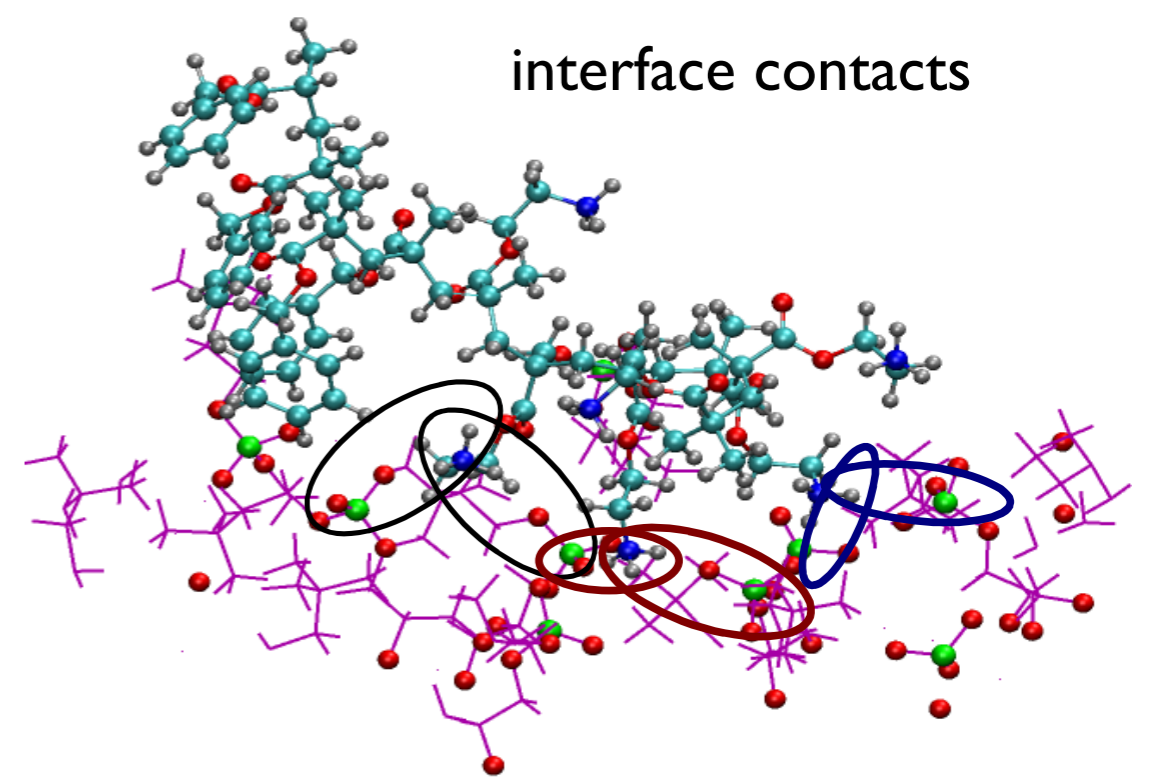
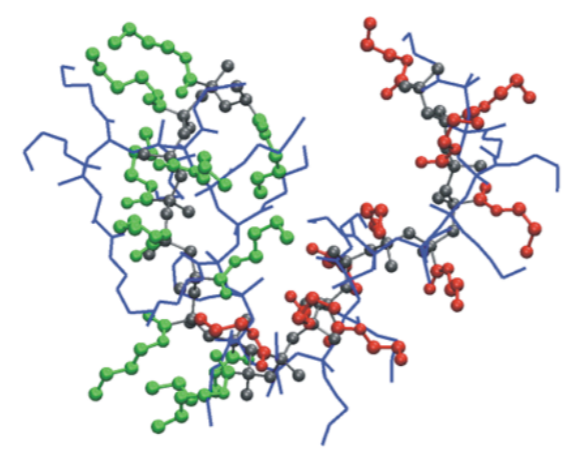
(b) 20-12-8-ben-nch0-block



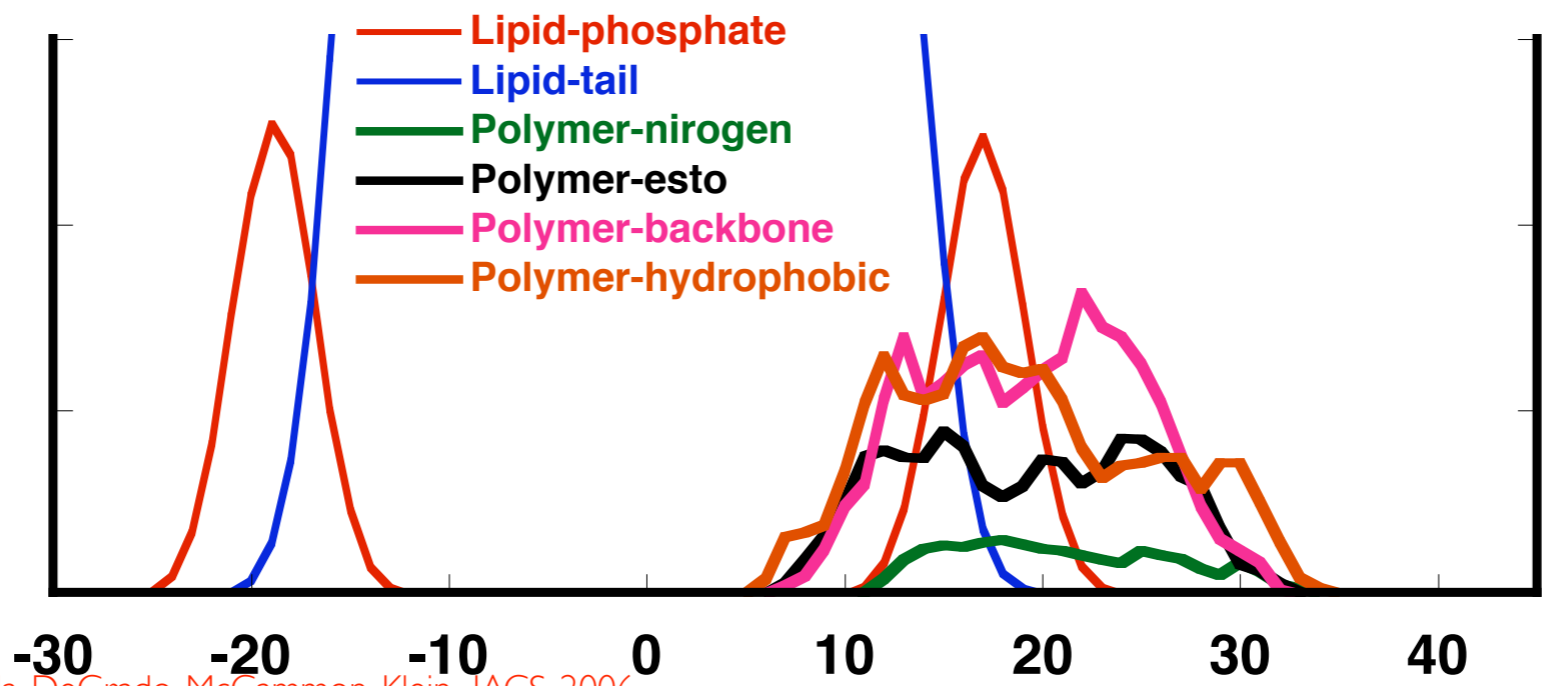
(c) 20-10-10-but-nch0-alt



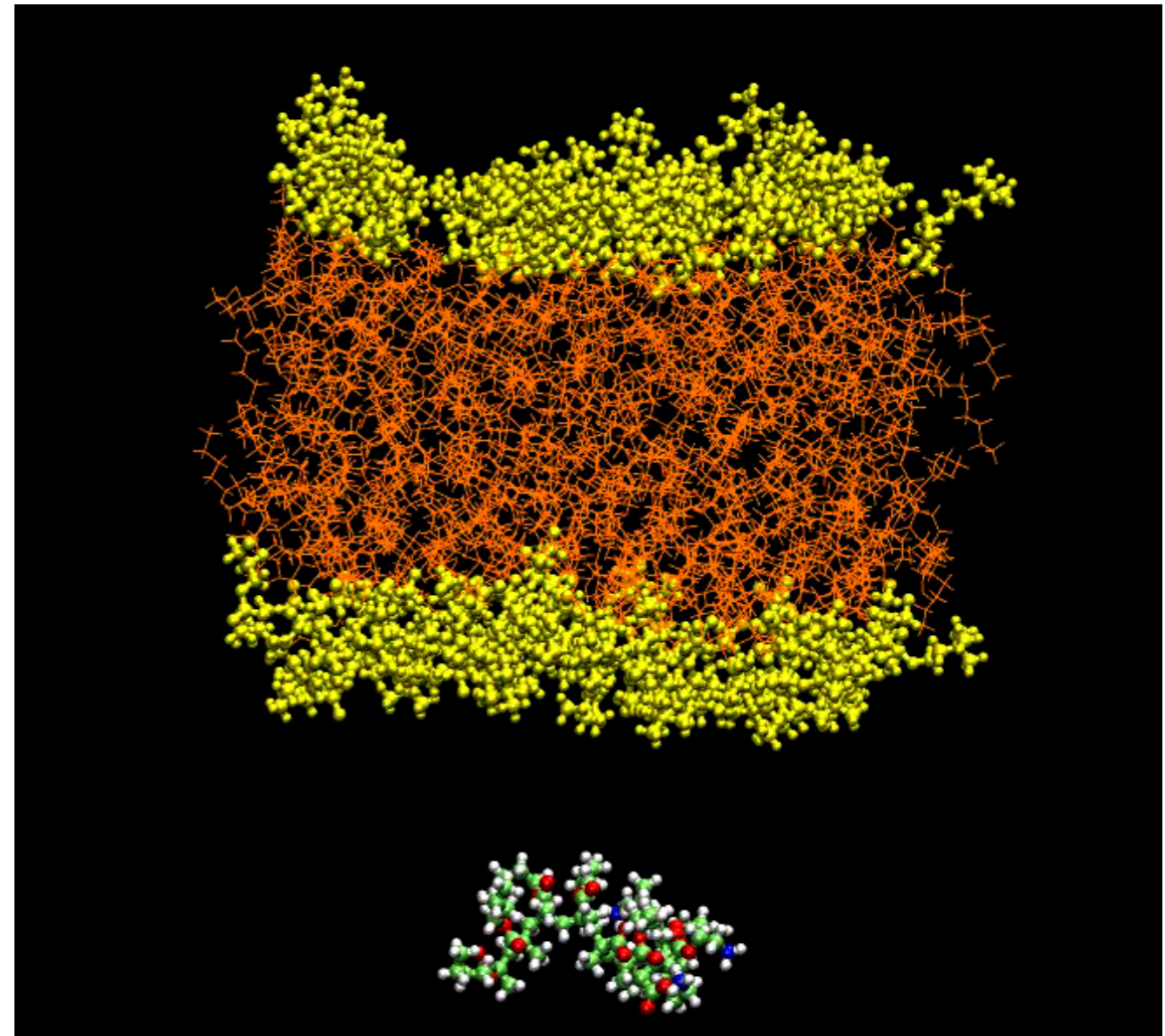
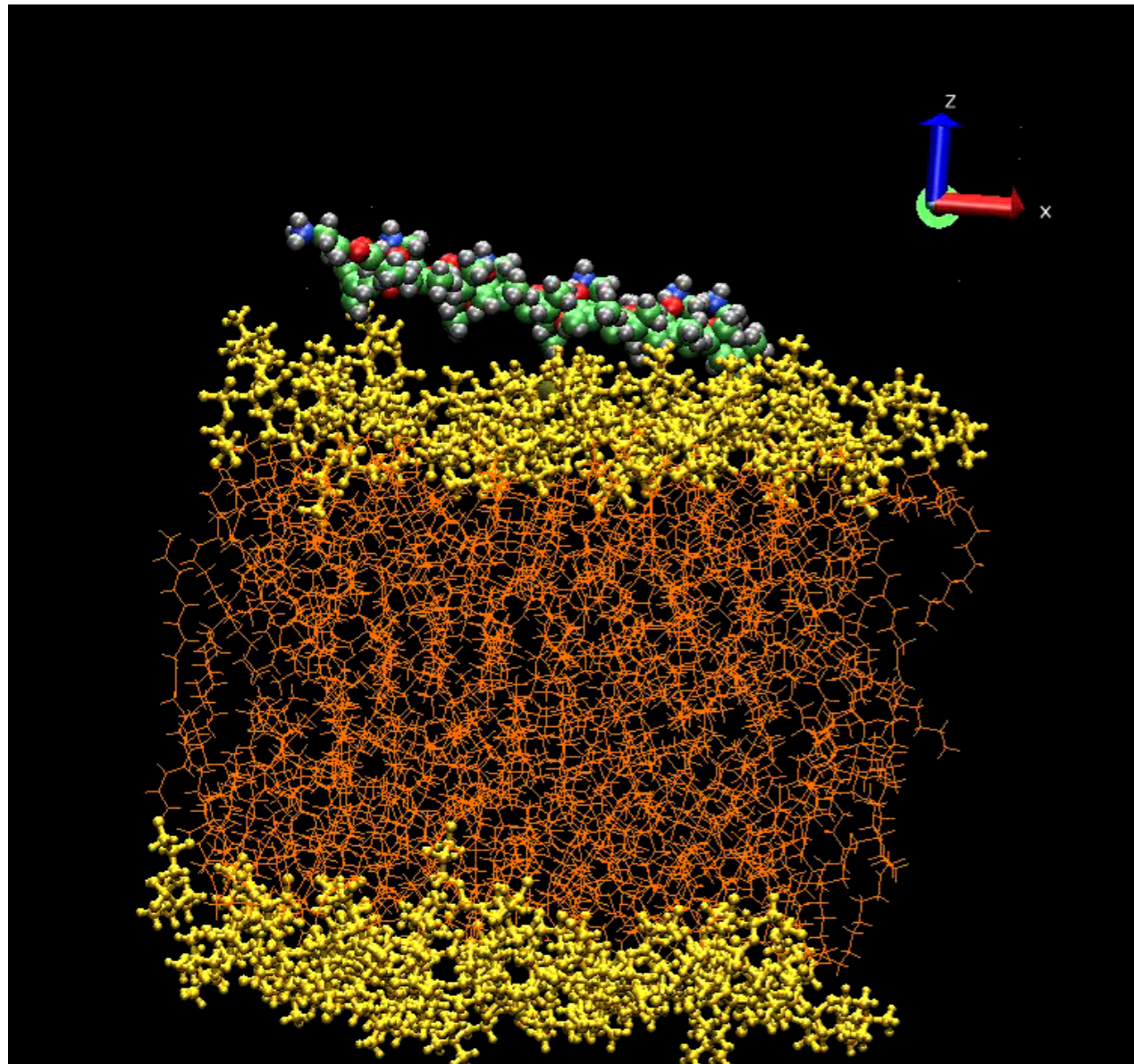
(d) 20-10-10-but-nch0-block



interface contacts

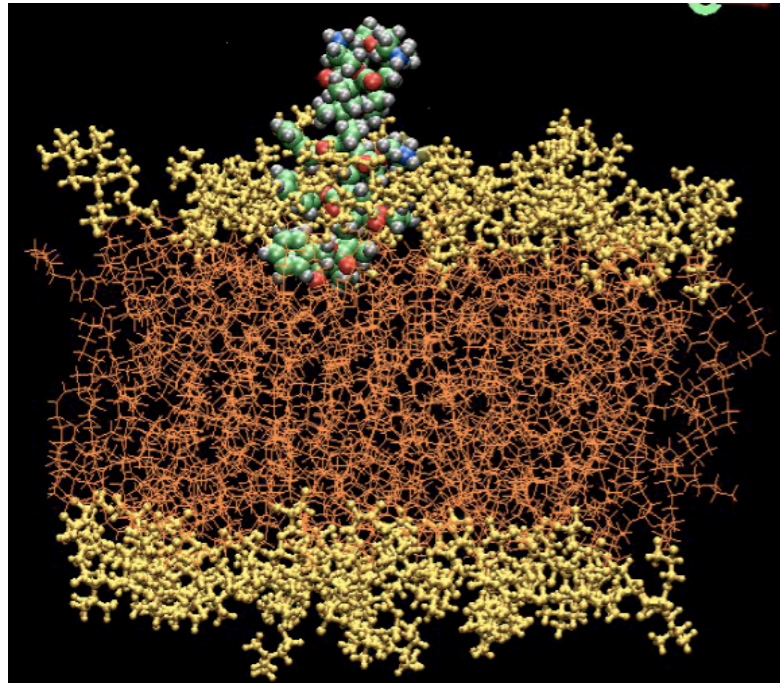


# Membrane insertion

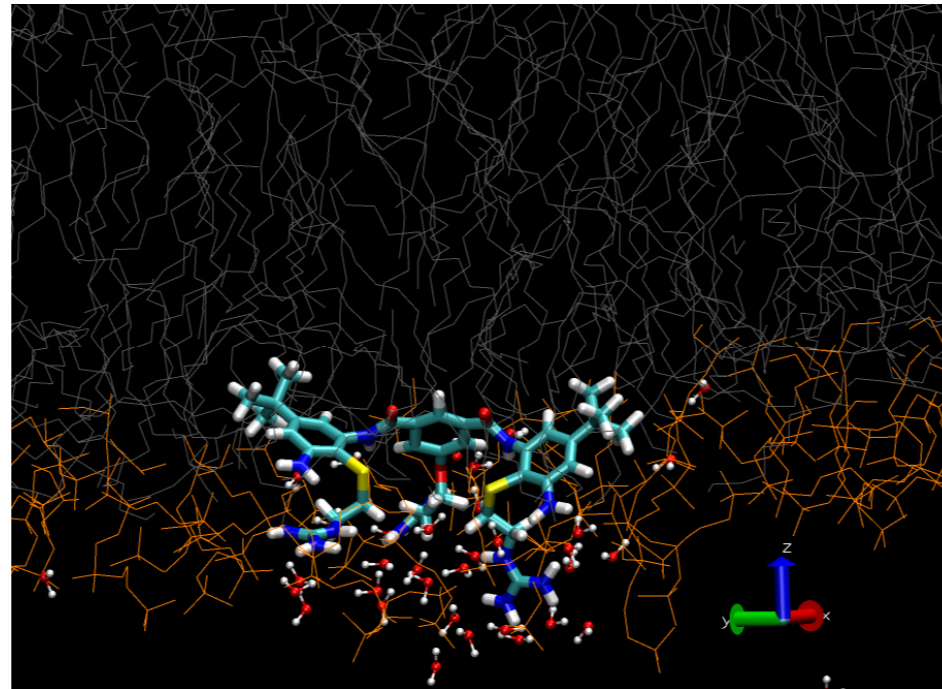


- More hydrophobic sequences undergo partial insertion
- Insertion: parallel to membrane normal
- amine groups play role in initial 'docking' to the membrane
- sequences with only charged groups remained in solution phase
- primary driving force: hydrophobic effect
- upon insertion, hydrophobic groups form favorable interactions with lipid tails
- dynamic flexibility allows polymer to adopt structures
- *structural properties depend on composition; hydrophobicity is primary determinant of activity and charged amine groups are important for selectivity*

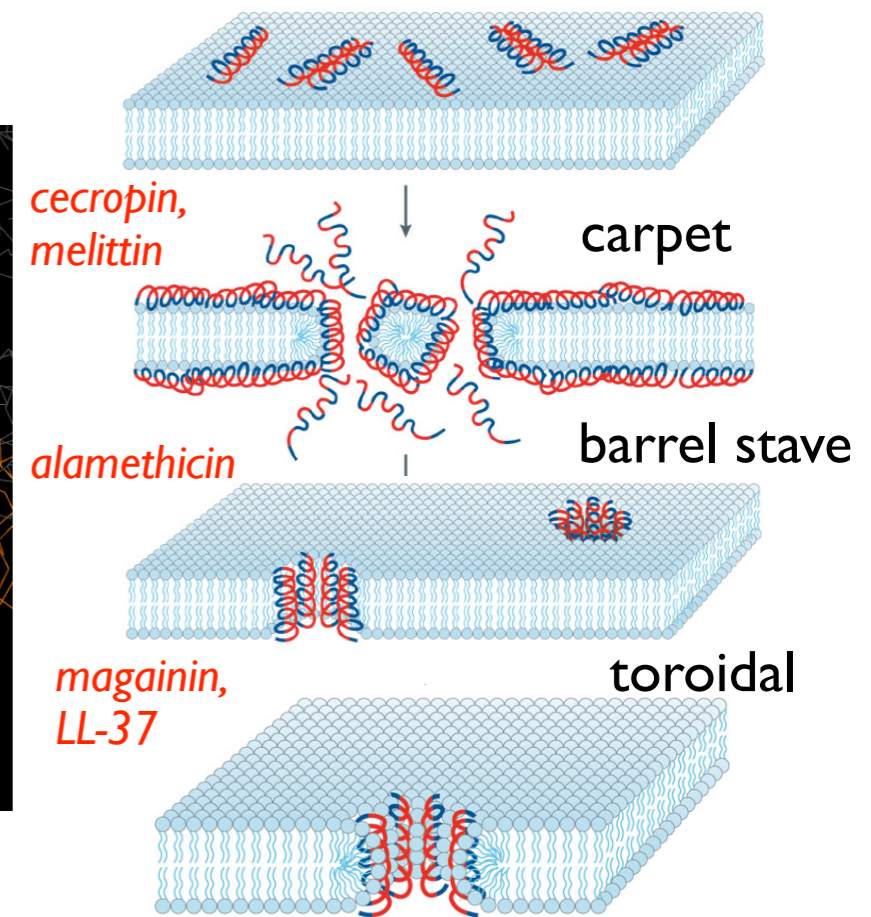
# Summary: antimicrobial polymers



Barrel stave?



Carpet mode?



- The composition, molecular weight and hydrophobicity of short co-polymers can be modulated to achieve structural diversity, which is crucial in controlling the antimicrobial activity.
- The primary driving force leading to insertion is confirmed to be the hydrophobic interaction with the membrane.
- Appropriate torsion backbone potentials have been developed for arylamide foldamers
- Our results indicate sensitive dependence of the overall shape on the sequence, suggesting that experimentally observed changes in activity can be correlated with particular sequences, providing an avenue for rational design.

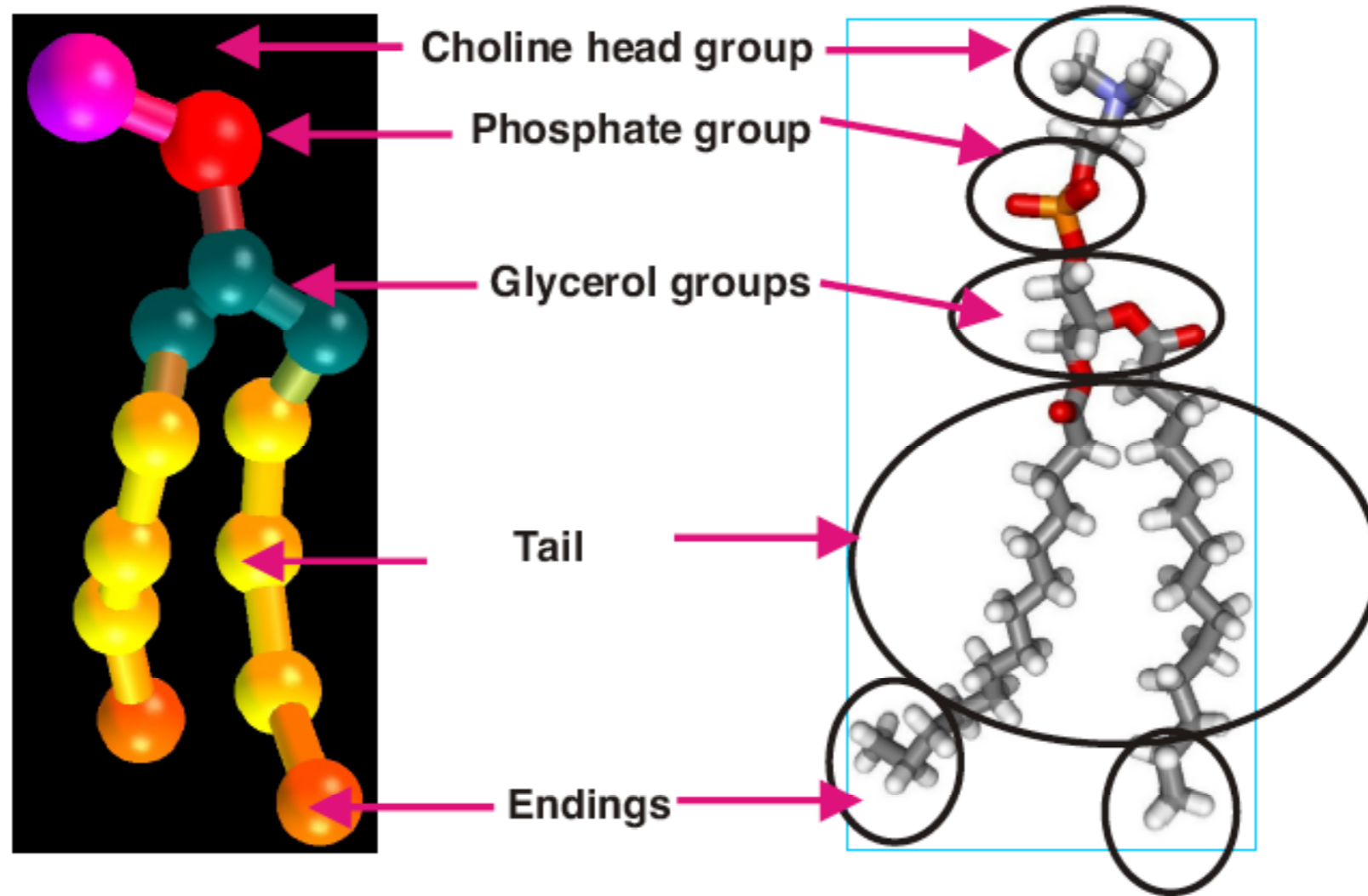
To understand more clearly the interplay of charged & hydrophobic groups and antimicrobial activity

To study antimicrobial activity in the context of aggregation

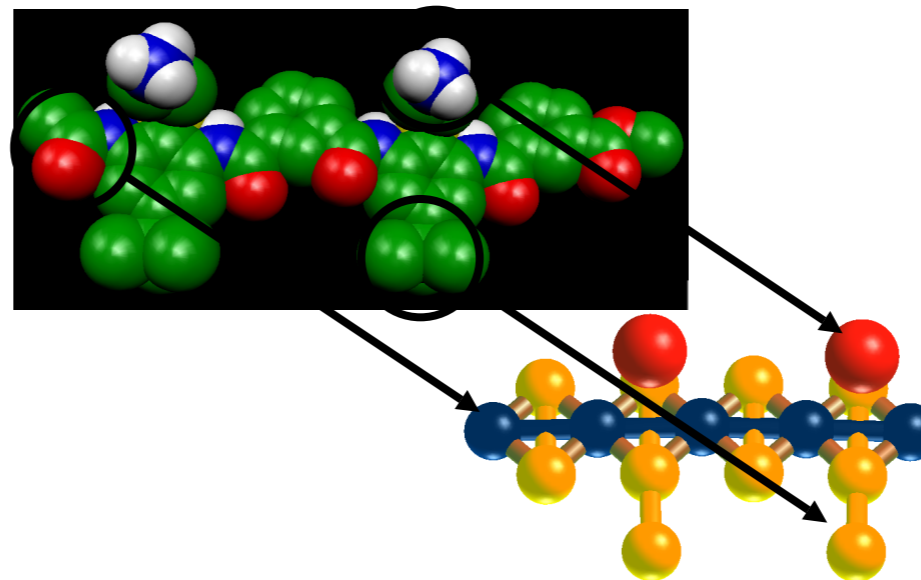
**coarse grain simulations**

# Coarse grained simulations?

Lipids:



AM polymers



# CG model: functional forms

$$U_{\text{intra}} = \sum_{\text{Bond}} k_b (r - r_0)^2 + \sum_{\text{Angle}} k_a (\theta - \theta_0)^2,$$

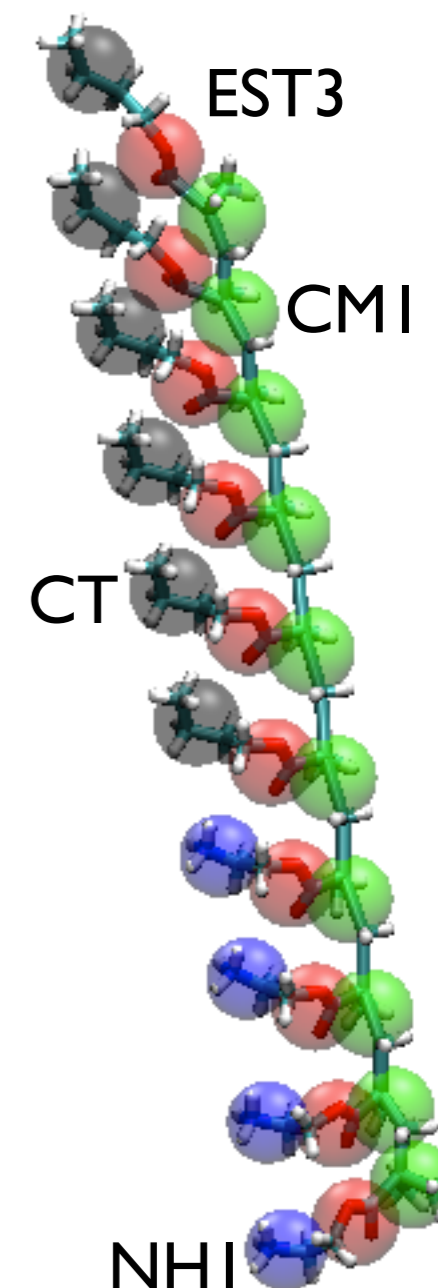
$$U_{\text{bond}}(r) \propto -k_B T \ln[P(r)/r^2]$$

$$U_{\text{angle}}(\theta) \propto -k_B T \ln[P(\theta)/\sin \theta],$$

$P(r), P(\theta) = \text{AA probability distributions}$

$$U_{\text{LJ}9-6}(r) = \frac{27}{4} \varepsilon \left\{ \left( \frac{\sigma}{r} \right)^9 - \left( \frac{\sigma}{r} \right)^6 \right\}, \text{ pairs involving water}$$

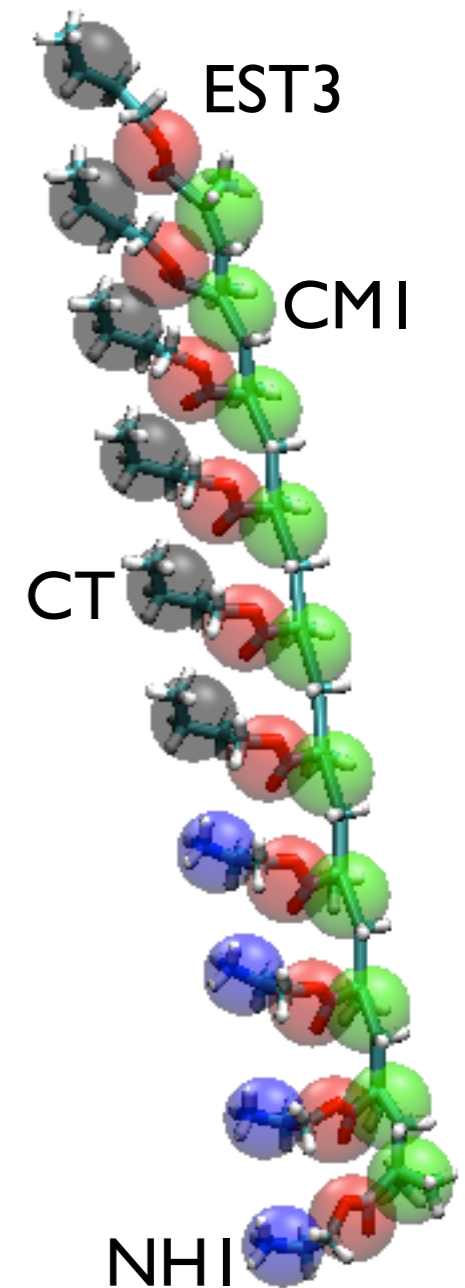
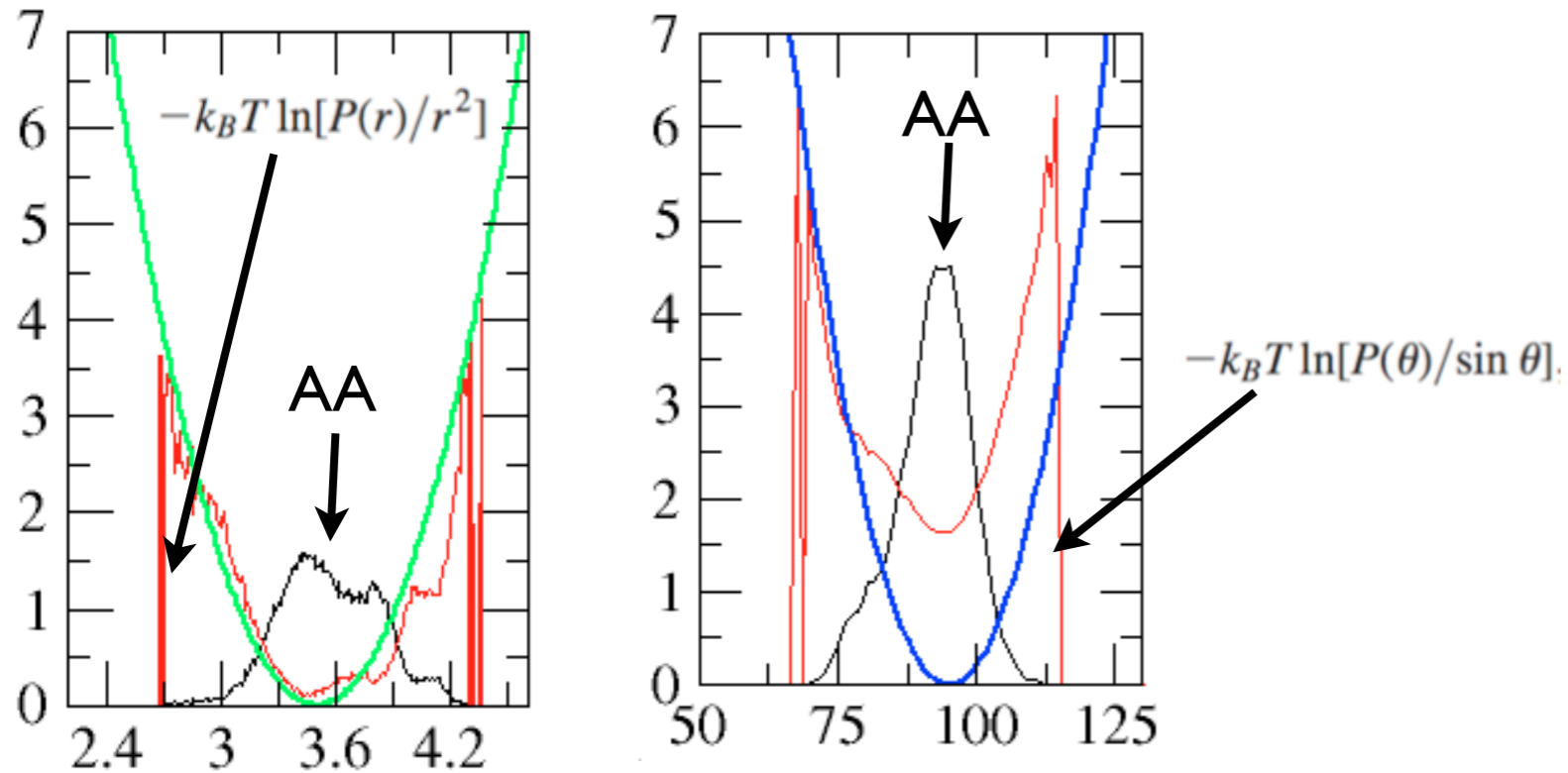
$$U_{\text{LJ}12-4}(r) = \frac{3\sqrt{3}}{2} \varepsilon \left\{ \left( \frac{\sigma}{r} \right)^{12} - \left( \frac{\sigma}{r} \right)^4 \right\}. \text{ all other pairs}$$





## Bonded Parameters:

- structural properties (pair distribution functions from AA-simulations)

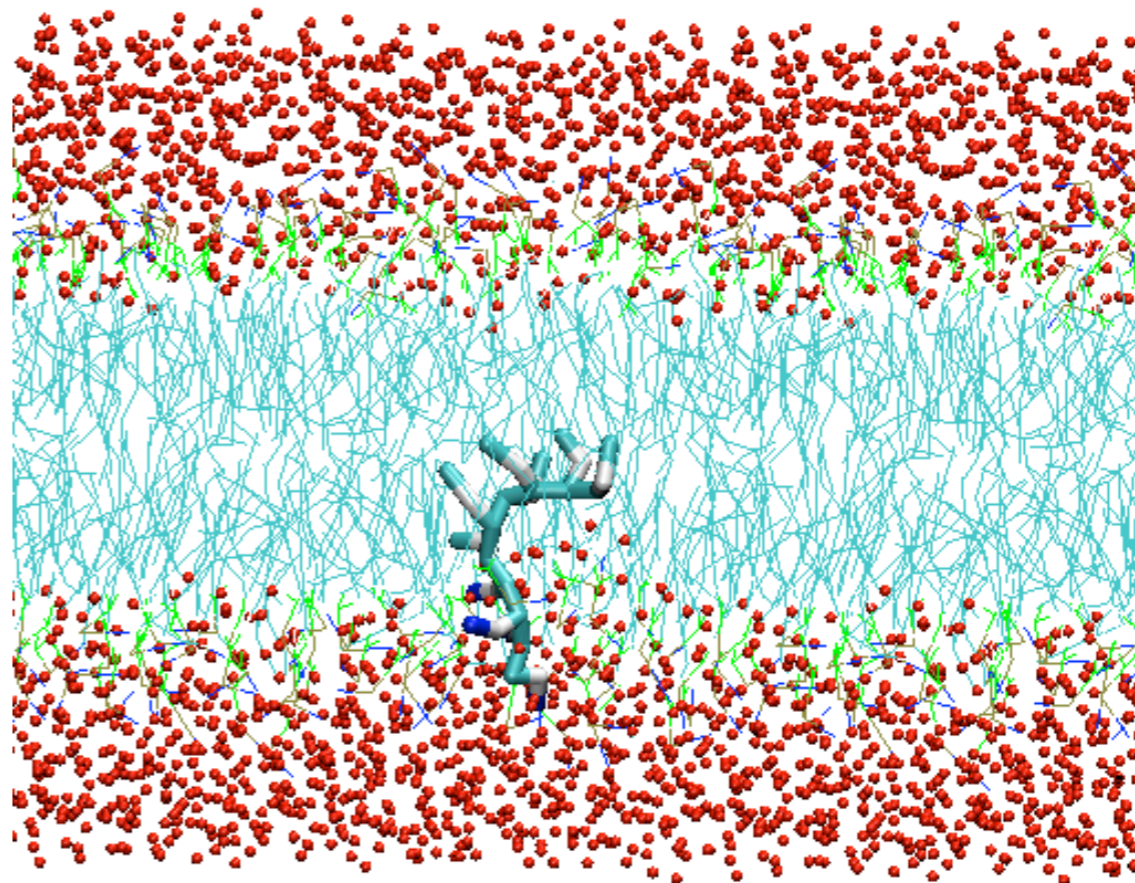
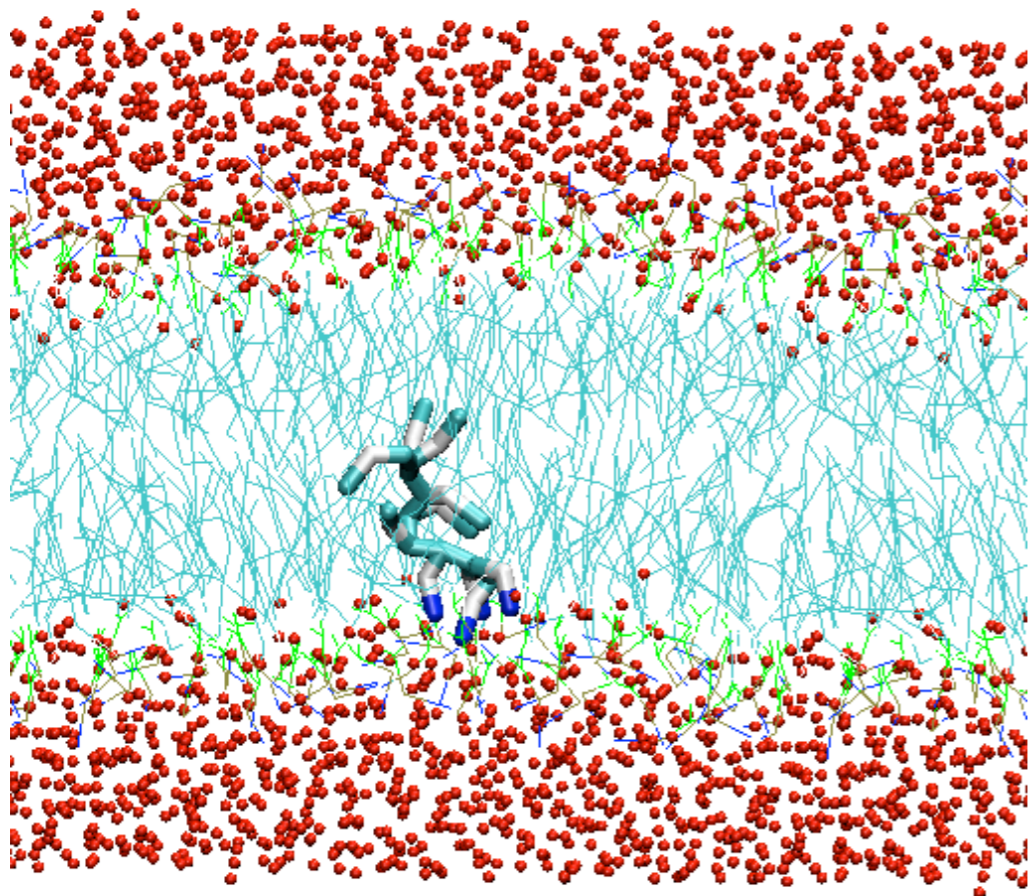


## Non-bonded Parameters:

- reproduce surface tension & density (if experimental numbers available)
- hydration free energy (X-water interaction)
- structural properties (pair distribution functions from AA-simulations)

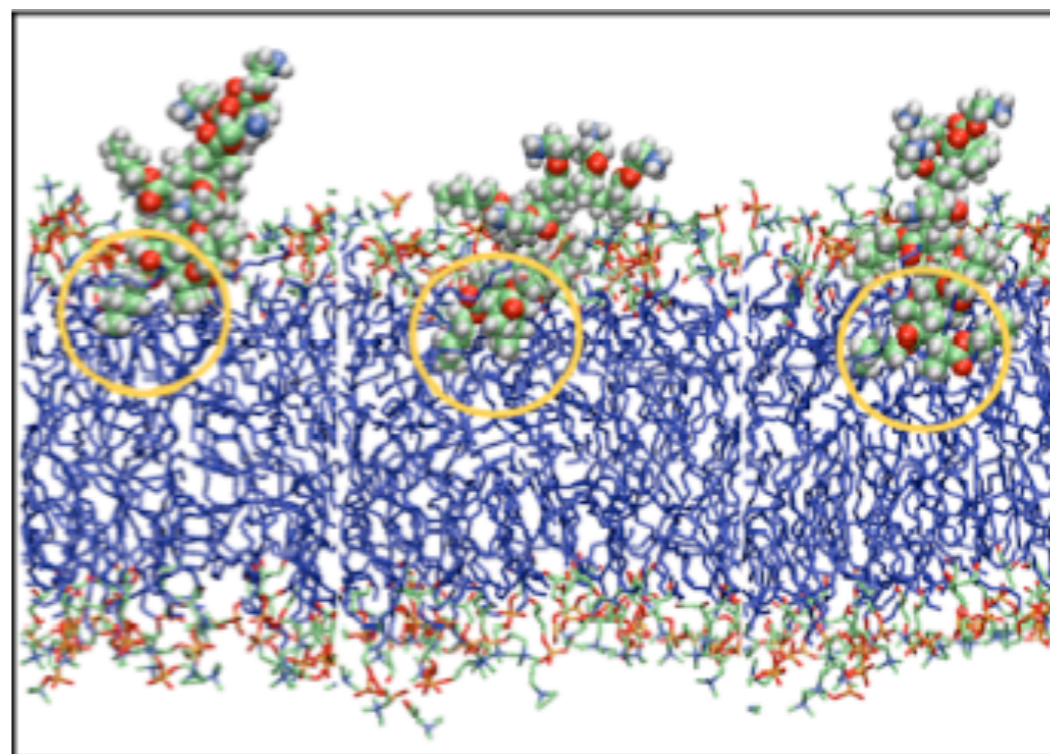
Initial parameters from Shinoda *et al.*,

# AMP in lipid (single)

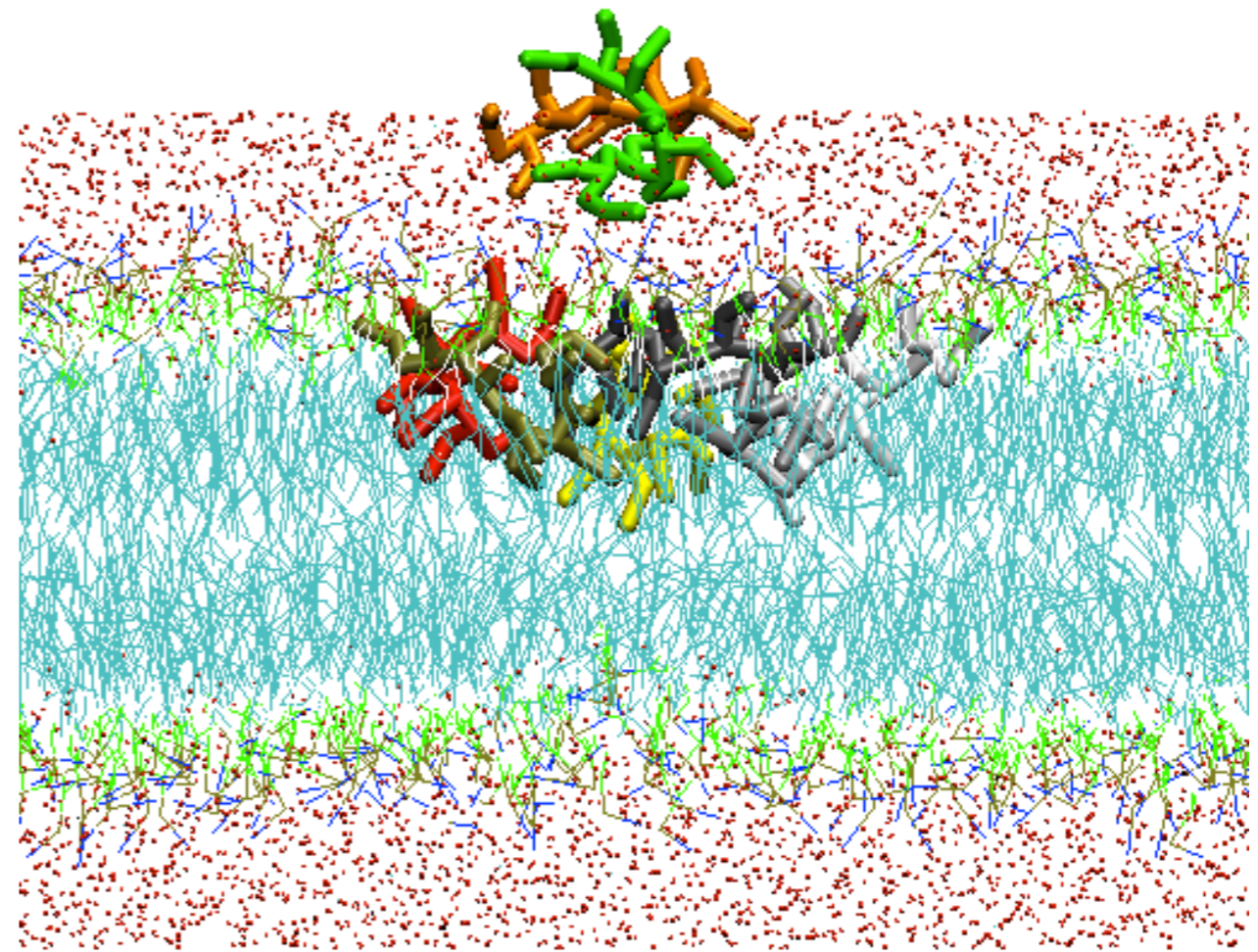
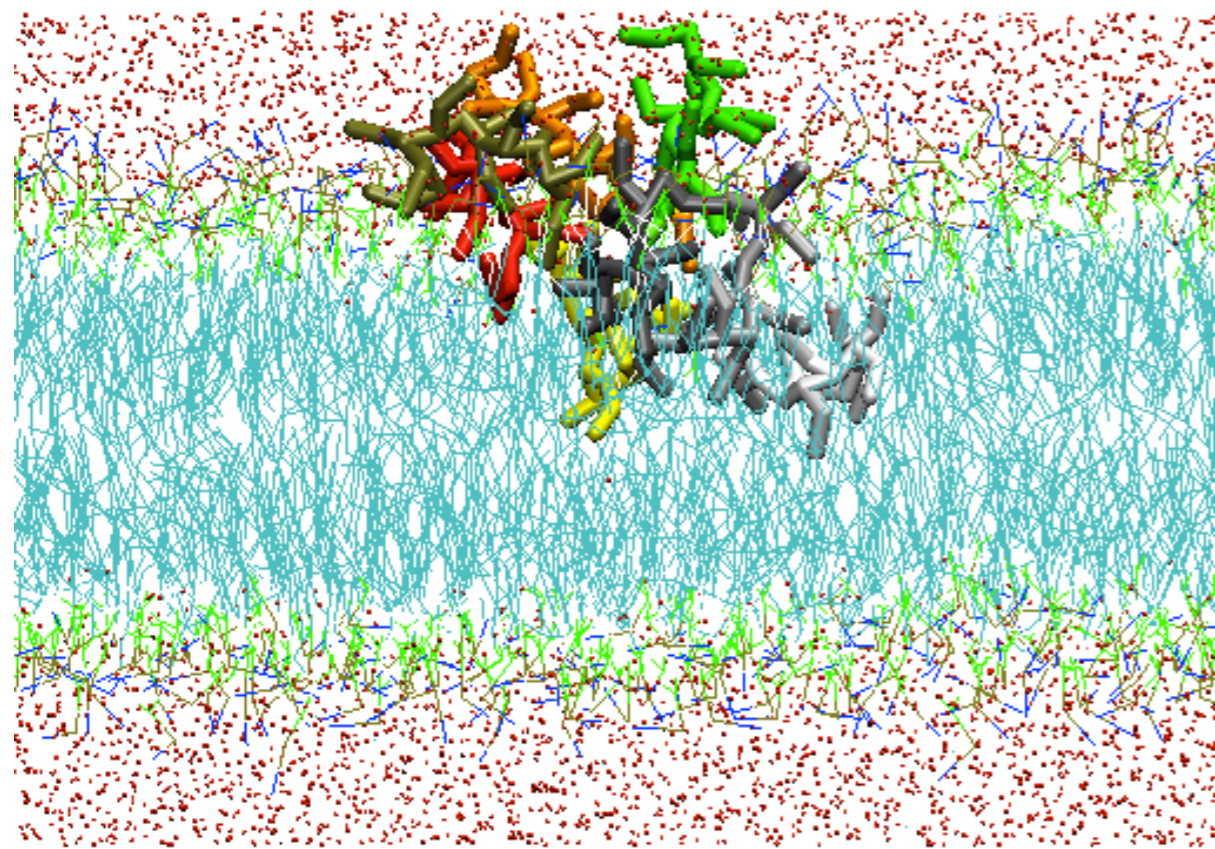


# CG sites: ~ 3400

# atoms in AA: ~25000



# AMP in lipid (aggregate)

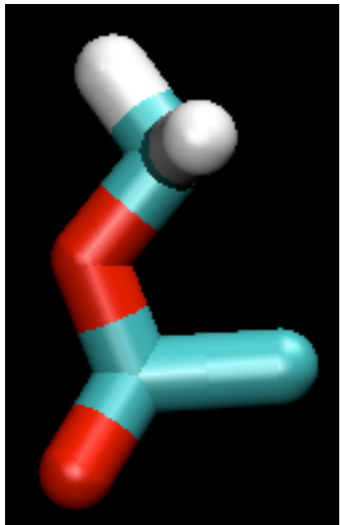


—————→  
10 ns

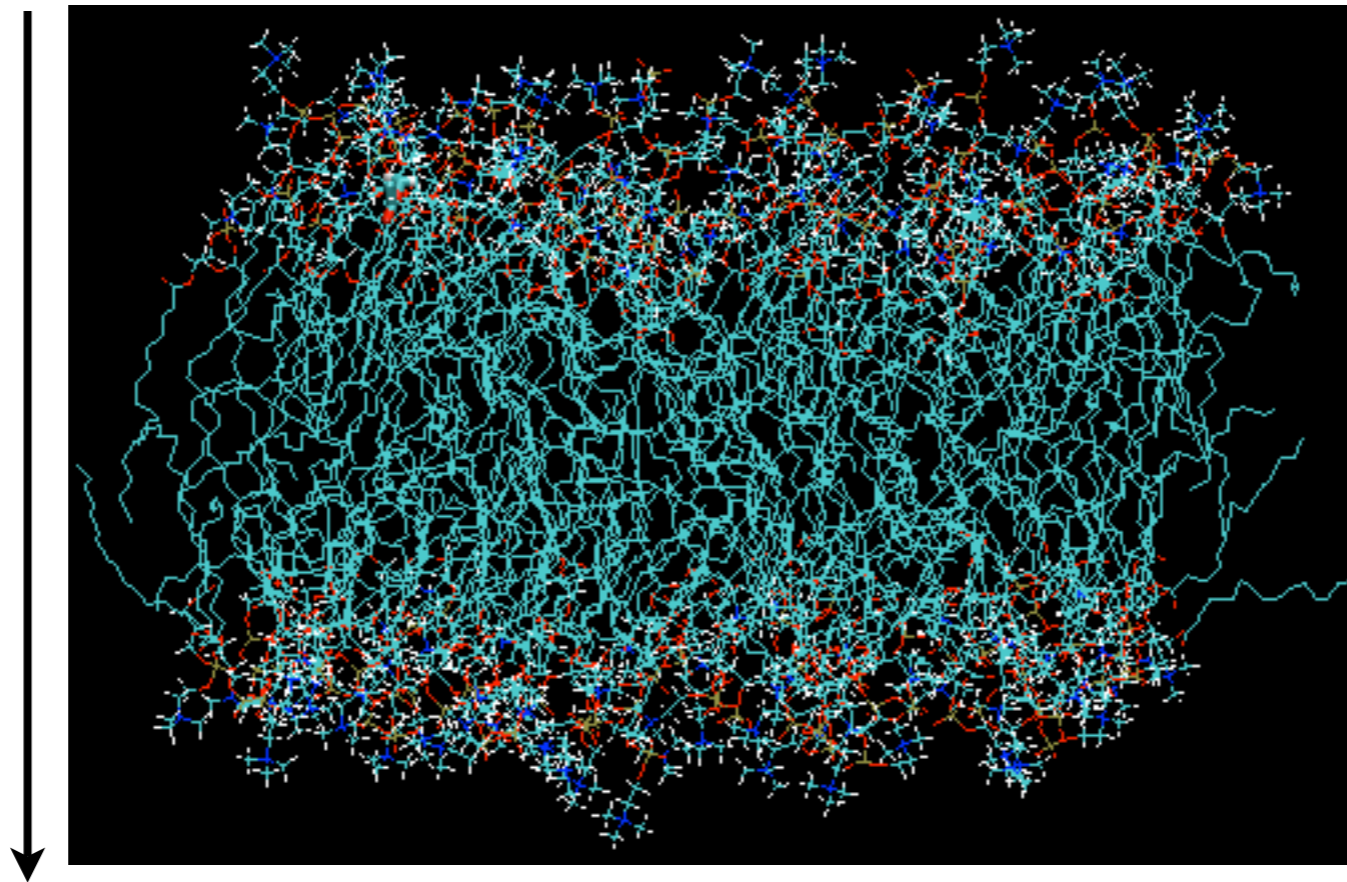
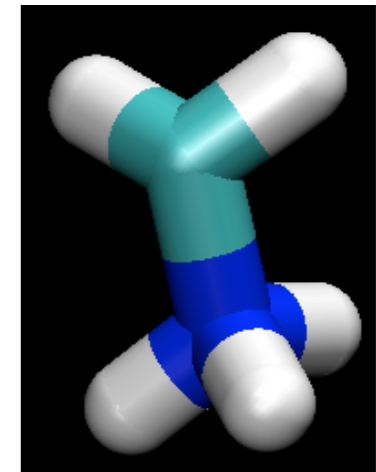
CG lipids: 512  
CG waters: 5000  
CG AMP: 8

further refinement:  
NB parameters based on PMF

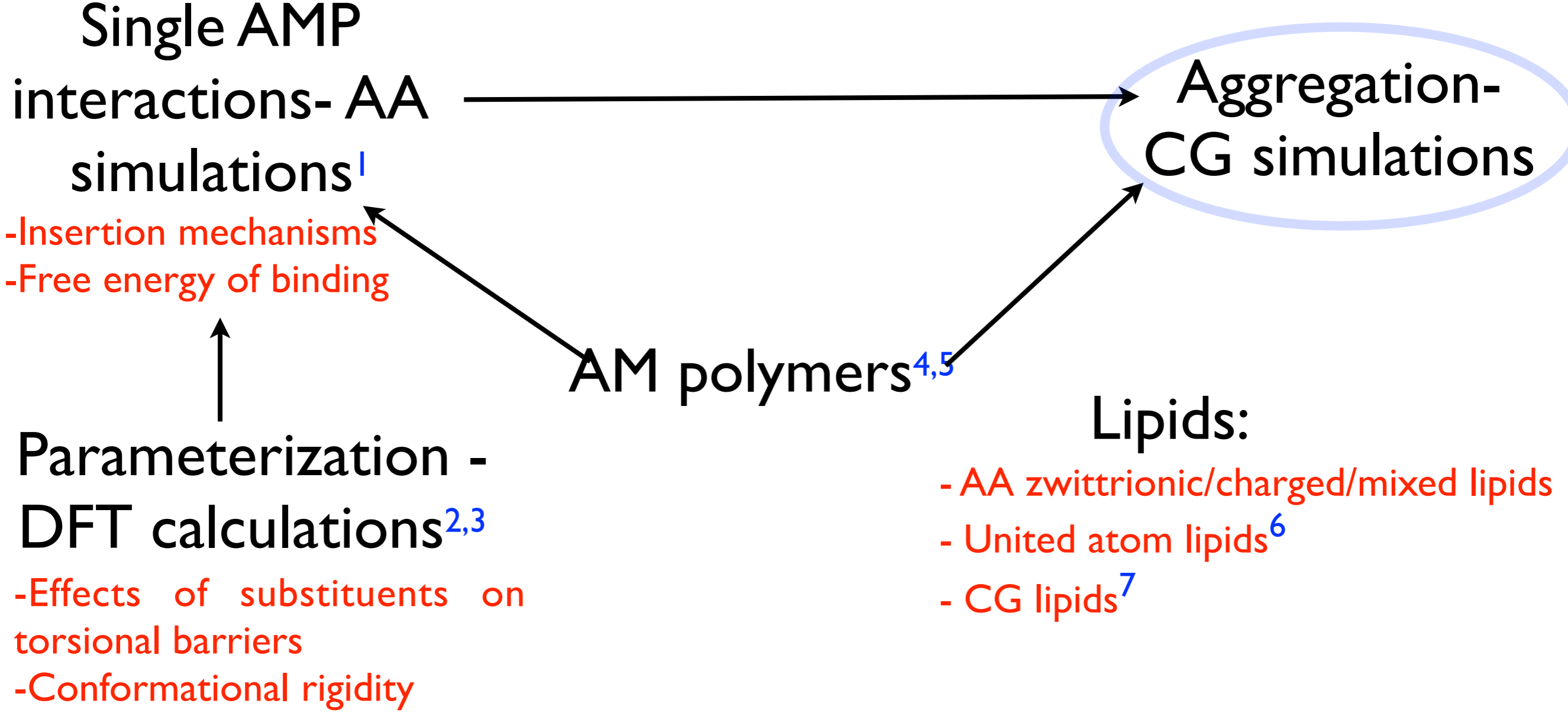
EST3



NH3 I



# Computational approach



1. JACS, 2006, 128, 1778-1779  
2. J Comp Chem, 2006, 27, 693-700  
3. J Phys Chem B, 2006, 110, 3517-3526  
4. Peptide Science (Review), 2008, 90, 83-93  
5. Acc. Chemical Res, 2009, ASAP article

Thank You